

Collaboration on protein measurement between AIST- NIST

Synthetic protein particles as measurement standards for subvisible aggregates: Progress 2014

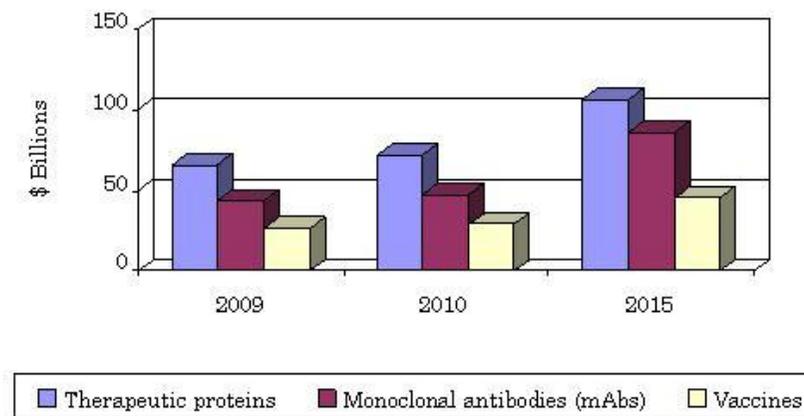
Shinya Honda

Molecular and Cellular Breeding Research Group

BioMedical RI, AIST, Japan

Growing market for pharmaceutical proteins

- The overall biologics market was **\$135 billion** in 2009, and was expected to grow to **\$149 billion** in 2010, a 9.4% growth rate.
- The total market is expected to grow to **\$239 billion** by 2015, a compound annual growth rate of **9.9% since 2010**.



Source: BCC Research (<http://bccresearch.wordpress.com/2012/09/25/>)

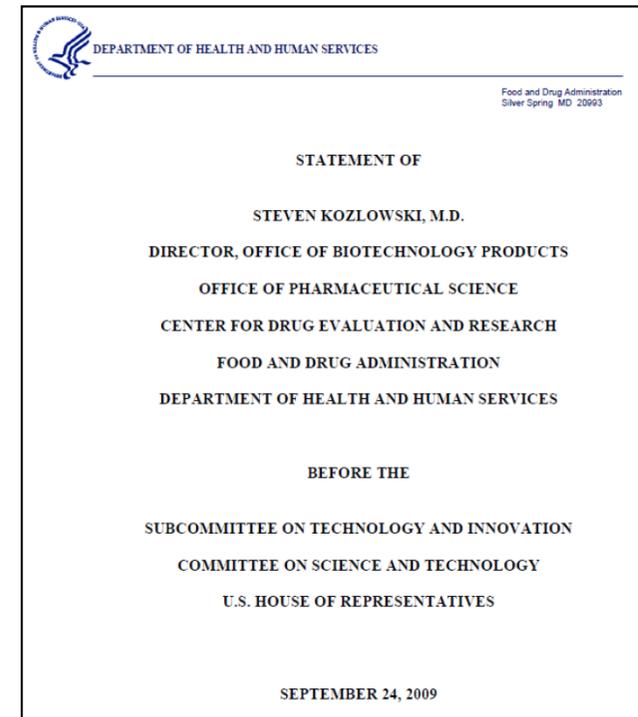
Three characteristics that should be analyzed

FDA statement before US House (2009)

- Advances in analytical tests during the last two decades have driven progress in biopharmaceutical manufacturing, but there is **still room for significant improvement**.
- FDA has identified **three properties** of therapeutic proteins that **cannot be sufficiently measured at this time** but that are very important for understanding the behavior of protein drugs.

Molecular properties on the frontier.

- Post-translation Modifications
- Three-dimensional Structure
- Protein Aggregation



Steven Kozlowski, M.D.
 Director, Office of Biotechnology Products
 Office of Pharmaceutical Science
 Center for Drug Evaluation and Research
 Food and Drug Administration

Inconsistency in measurements for protein aggregation

- Comparing the amount of aggregates from an antibody drug measured by three different methods
 - Gabrielson et al *J. Pharm. Sci.* 96, 268- (2007)

Table 1. Monomer and Soluble Aggregate Concentrations as a Mass Percentage of Total Protein for Unstressed and Acidified Antibody Samples

Aqueous Antibody Sample	Species	% by Mass of Total Protein		
		SV-AUC	SEC	AF4
Unstressed	Monomer	95.8 ± 0.5	99.6 ± 0.1	97.8 ± 1.1
	Total soluble aggregate	4.2 ± 0.5	0.4 ± 0.1	2.4 ± 1.1
Acidified	Monomer	84.6 ± 0.8	86.4 ± 1.5	80.3 ± 2.8
	Total soluble aggregate	15.4 ± 0.8	13.6 ± 1.5	19.7 ± 2.8

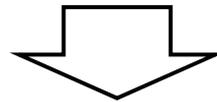
Reported values are mean ± one standard deviation ($n = 3$).



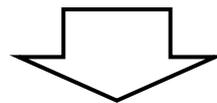
need for standardization

Aim and strategy

- Native proteins are generally unstable and smaller than aggregates.

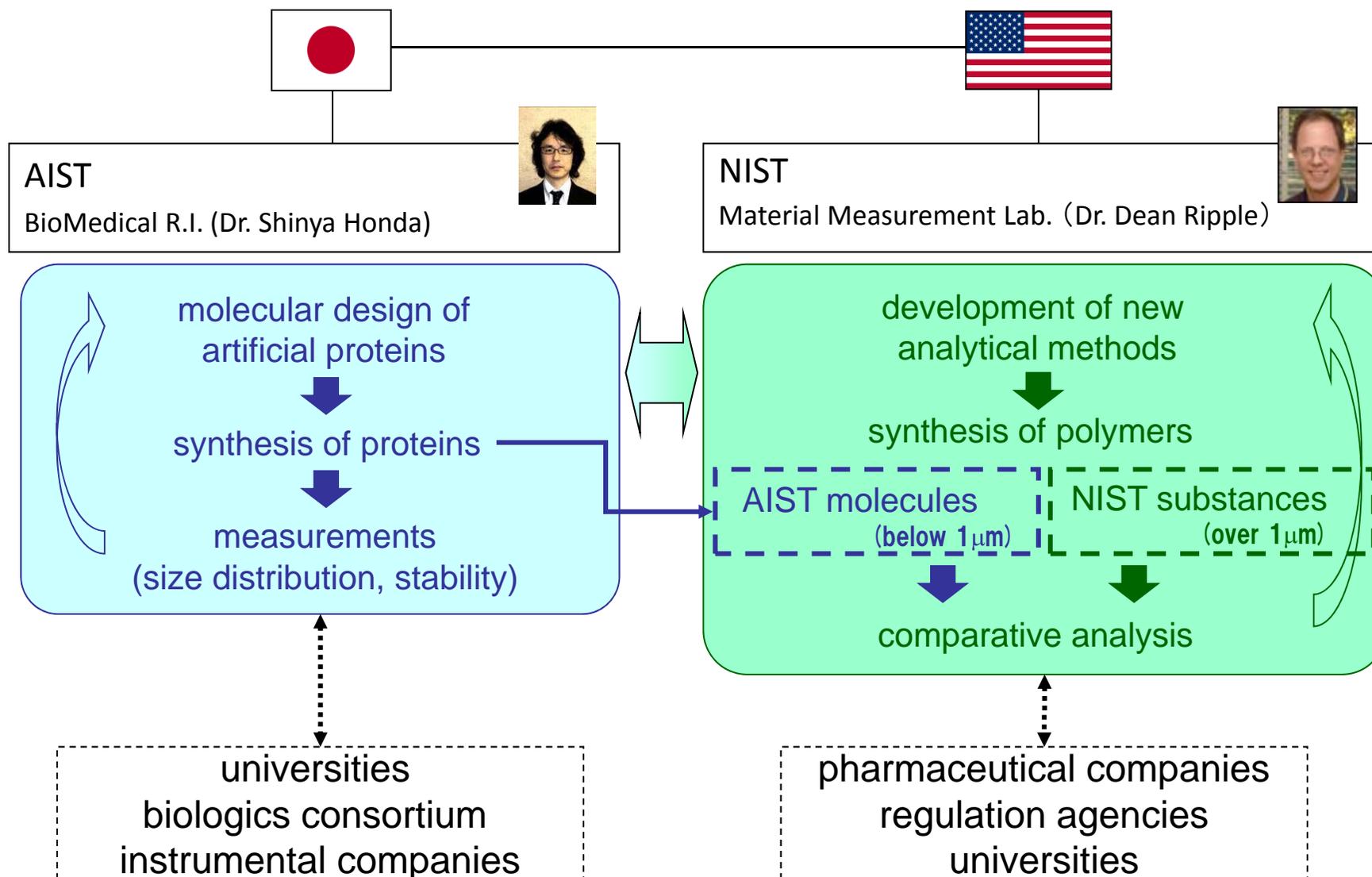


- We aim to synthesize **large protein particles** with enough stability by modifying native proteins using molecular design technique.

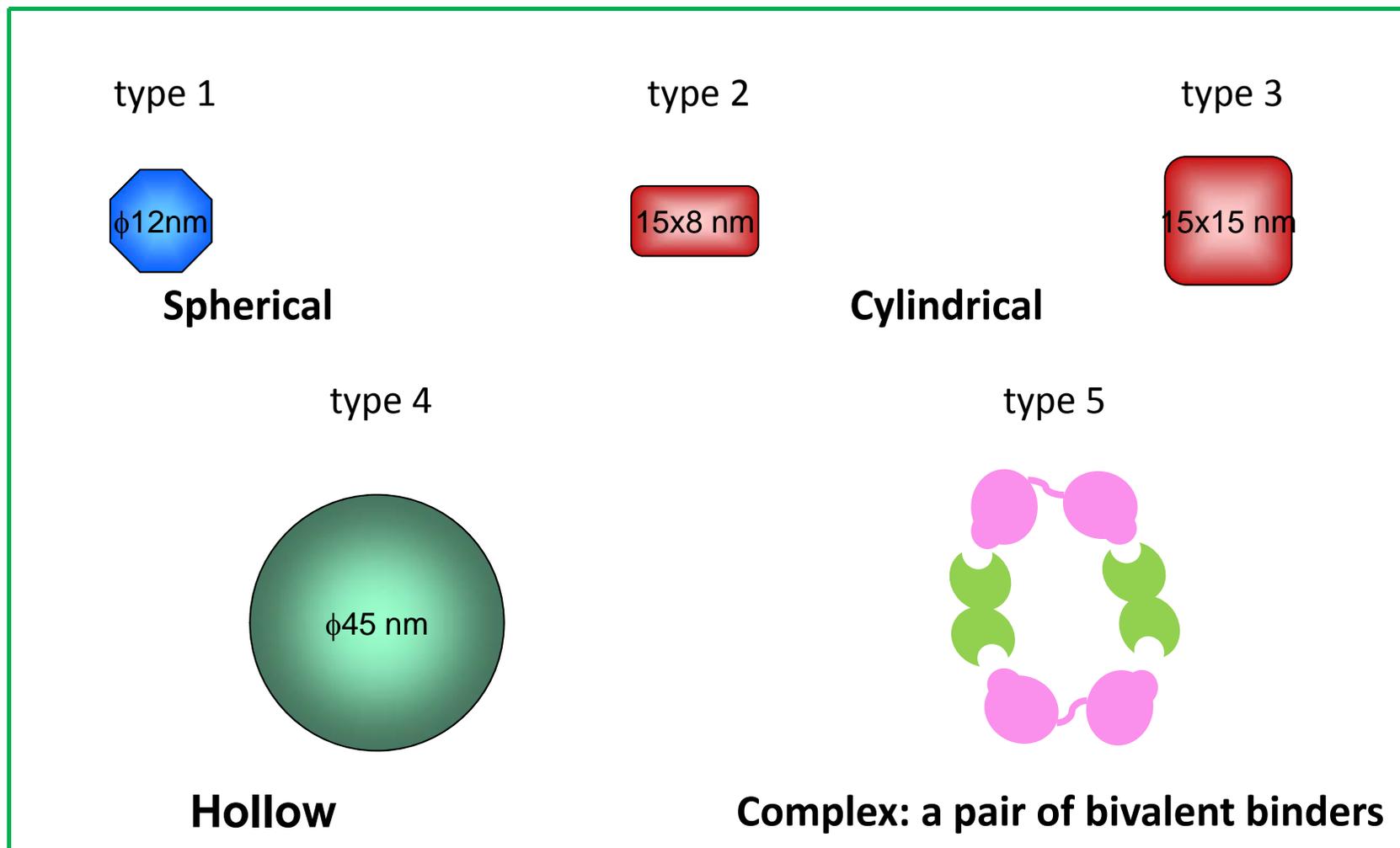


candidates for pre-authentication reference materials

Collaboration outline



Molecular candidates



Development flow

(Stage 1) Synthesis

- Gene synthesis
- Vector construction and cloning
- Bacterial expression
- Purification
- Identification
- Method optimization
- Large preparation
- Lyophilization

(Stage 2) Characterization

- Analytical size exclusion chromatography (SEC)
- MALDI-TOF-MS analysis
- Analytical ultracentrifuge
- Dynamic light scattering (DLS)
- FT-IR

(Stage 3) Homogeneity Assessment

- Analytical SEC
- DLS
- Circular dichroism
- Statistical analysis

(Stage 4) Stability Test

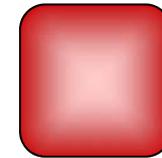
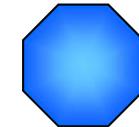
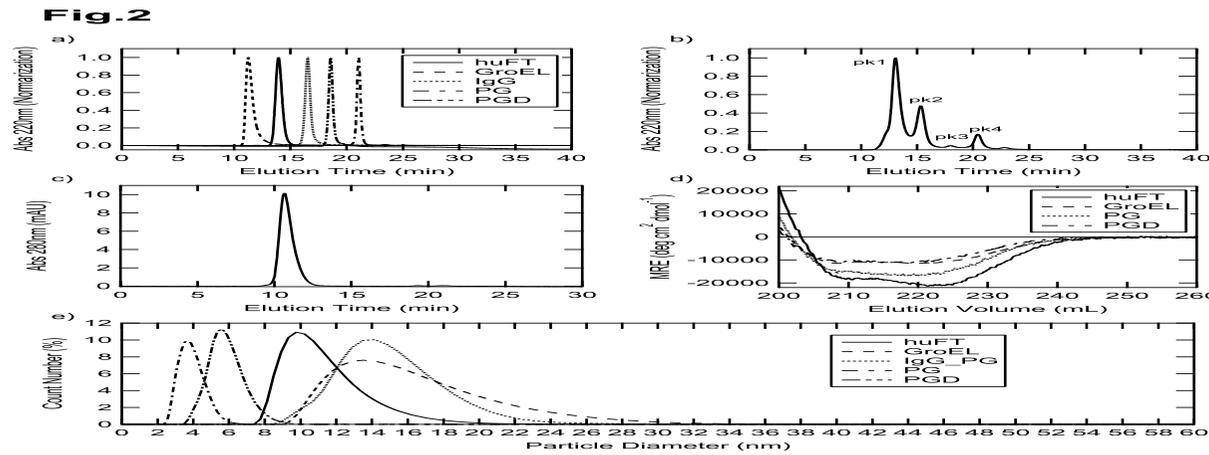
- Heat stress test
- Short-term storage stability
- Statistical analysis

(Stage 5) Documentation

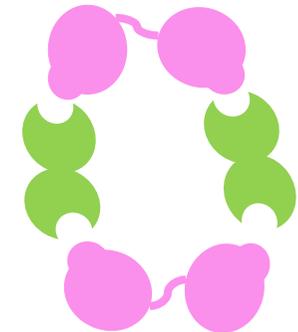
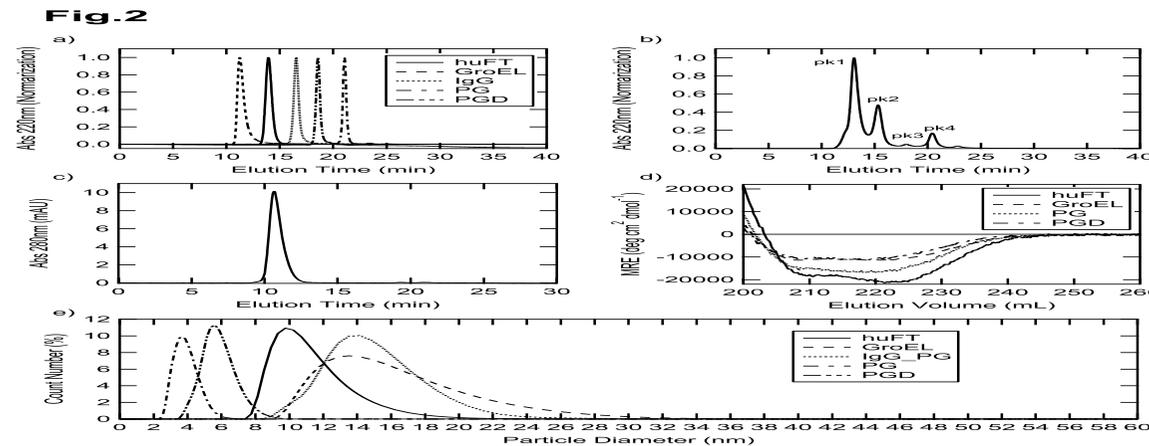
- Paper to journal
- Draft proposal to authority

Characterization: Type 1, 3, and 5

- SEC

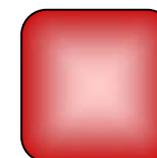
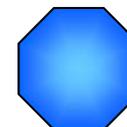


- DLS



Homogeneity: Type 1 and 3

- Between-bottle



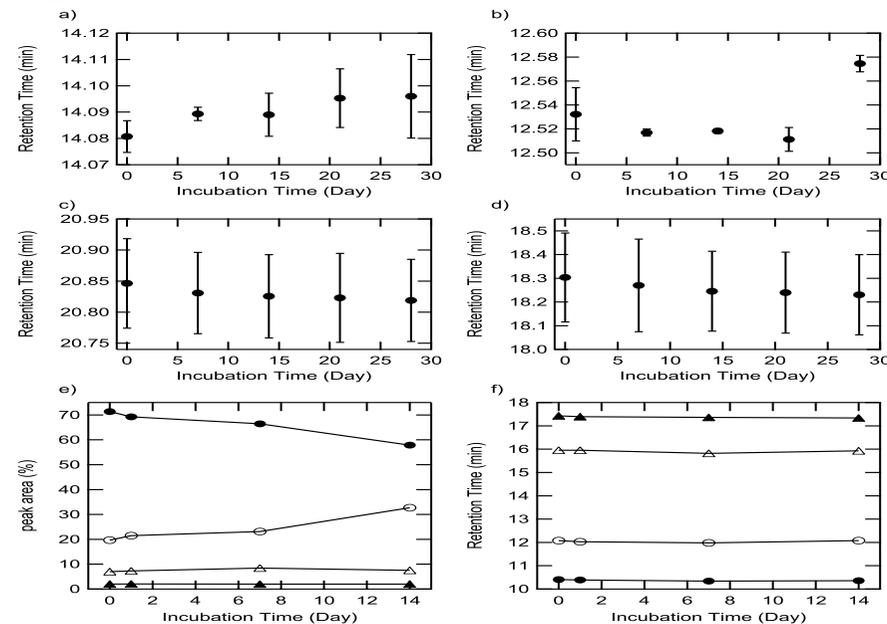
			Type 1	Type 3	
SEC	Retention Time	within-bottle (min)	14.17 ± 0.004	12.57 ± 0.006	
		between-bottle (min)	14.17 ± 0.011	12.56 ± 0.006	
		F-test	No	Yes	
		t-test	Yes	No	
	Purity	within-bottle (%)	99.5 ± 0.03	99.9 ± 0.05	
		between-bottle (%)	99.4 ± 0.09	99.9 ± 0.07	
			F-test	Yes	Yes
			t-test	No	Yes
CD _{222nm}		within-bottle (MRE)	-20348 ± 355	-8222 ± 283	
		between-bottle (MRE)	-20614 ± 668	-8712 ± 322	
		F-test	Yes	Yes	
		t-test	Yes	No	
DLS		within-bottle (nm)	12.76 ± 0.32	17.26 ± 0.14	
		between-bottle (nm)	12.55 ± 0.17	17.37 ± 0.11	
		F-test	Yes	Yes	
		t-test	Yes	Yes	

Stability: Type 1 and 3

- Storage stability by SEC retention time
 - 40, 25, 4, -30°C; 75, 60% RH; 0-4 weeks

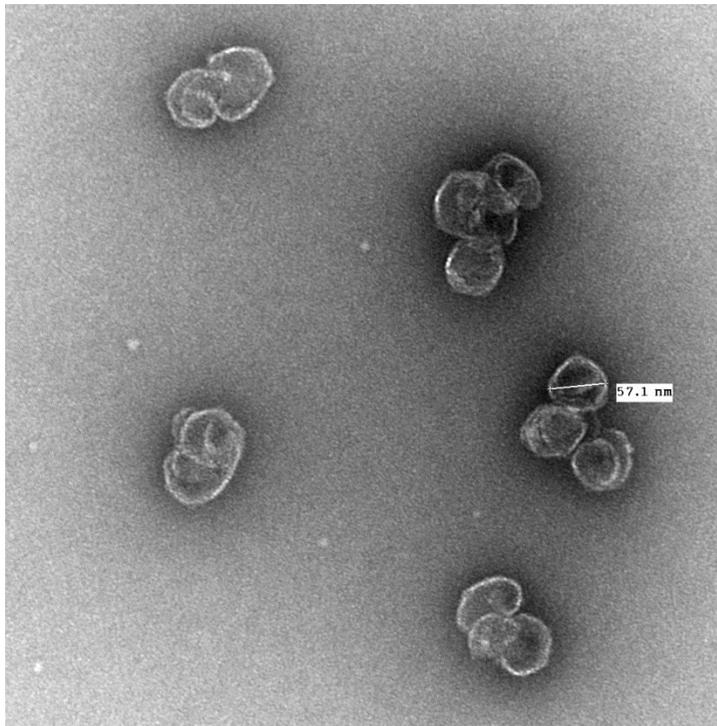
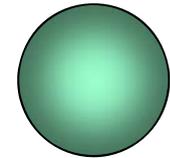


Fig. 4



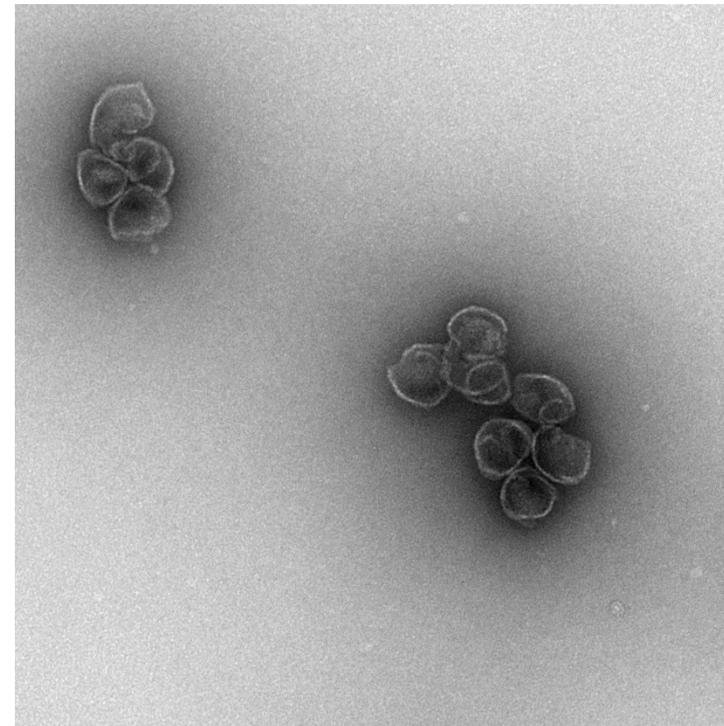
Characterization: Type 4

- Electron microscope (negative staining)



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AMT Camera System



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Documentation

- Paper to journal
 - H Takahashi, Y Feng, A Ooishi, H Watanabe, and S Honda, “Towards the development of a reference material for protein particle”, *submitted*.
- Draft proposal to authority
 - “General requirements of the protein-related standard material to use for aggregation measurement analysis”, *in preparation*

2014 Summary and future plans

- Synthetic methods for all five types of candidates were established.
- Extensive characterizations for Type 1, 3, and 5 candidates were carried out.
- Homogeneity assessments and storage stability analyses for bottled samples (Type 1, 3, and 5 candidates) were performed.
- Purification method for Type 4 candidate was improved.
- Large amount of Type 1 and 3 samples was shipped to NIST.
- Paper and draft proposal were prepared.

- Storage stability analyses for Type 4 candidate will be conducted.
- Inter-laboratory validation study will be continued with NIST.

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