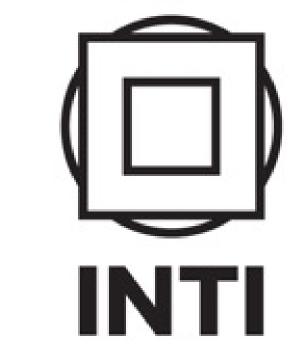


# **Development and Value Assignment of an Incurred Multi-Mycotoxin Reference Material**

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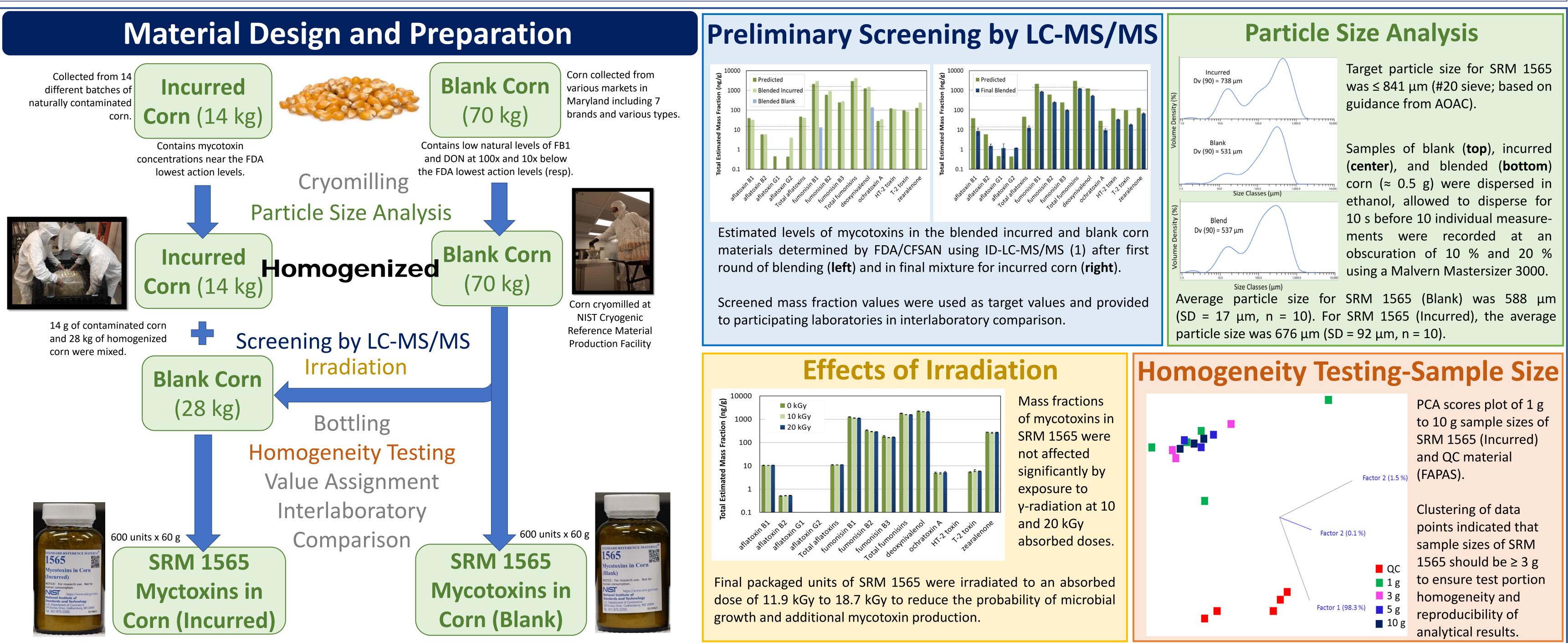
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### Why a Multi-Mycotoxin Material?

Fungi of the genera Aspergillus, Fusarium, and Penicillium produce mycotoxins, fumonisins, deoxynivalenol, zearalenone, ochratoxin A, HT-2 toxin, and T-2 toxin onto foods and feeds during harvests, storage, or onto finished products under warm or humid conditions. Although mycotoxin contamination can be minimized with the implementation of good agricultural and manufacturing practices, dietary exposure to humans and animals is unavoidable as these toxins are resistant to degradation by current food-processing procedures. Due to the numerous negative health impacts associated with the consumption of foodstuffs contaminated with mycotoxins, accurate determination is an international concern. Many commerciallyavailable certified RMs for mycotoxins mainly address a single mycotoxins, requiring the use of multiple RMs for multi-target methods. In addition, none of these RMs contain all of the FDA-regulated mycotoxins and mycotoxins of health significance. To address the increasing needs of laboratories moving toward LC-MS-based multimycotoxin analysis, the U.S. National Institute of Standards and Technology (NIST) collaborated with the U.S. Food and Drug Administration (FDA) to produce a naturally incurred RM for multiple mycotoxins in corn.





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### Interlaboratory Comparison

38 participating laboratories for NIST Health Assessment Measurements Quality Assurance Program (HAMQAP) Exercise 1. Participants were provided with blinded samples of SRM 1565 (Blank), SRM 1565 (Incurred), and a QC material (FAPAS). Asked to prepare 3 samples and report 3 results for each mycotoxin or group of mycotoxins in each sample, using their routine methods of analysis.

Control Target

- A stratified random sampling scheme was used by NIST and FDA to select 10 bottles of SRM 1565
- **Sample Preparation**

Value Assignment by NIST and FDA/CFSAN

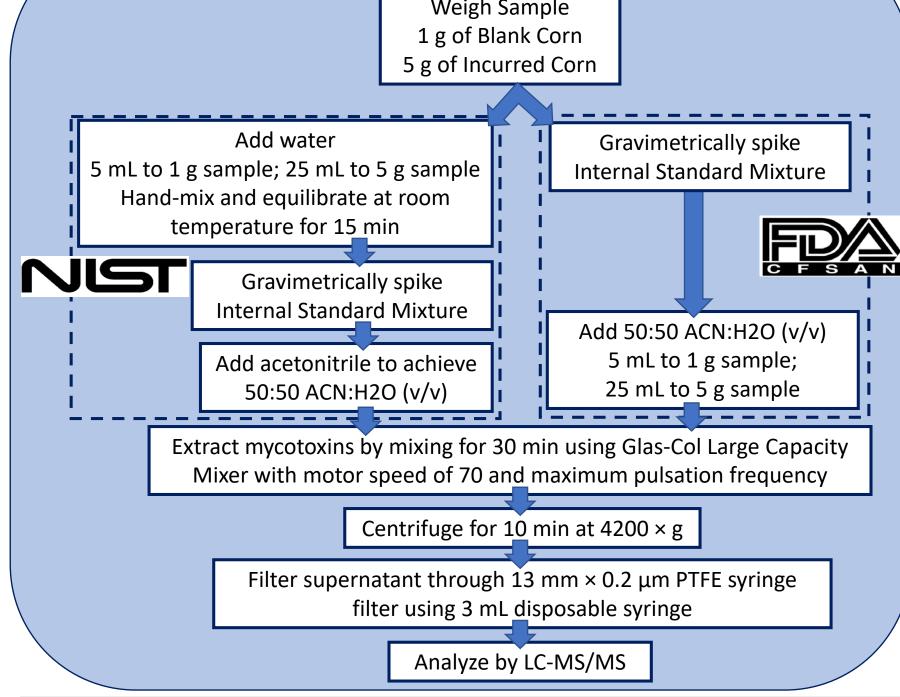
- (Incurred) and SRM 1565 (Blank) to prepare in duplicates.
- A QC material with know levels of mycotoxins was purchased from FAPAS to use as a control and was prepared in duplicates.
- Recovery studies revealed automated shaking was optimal for extraction and no significant increase in extraction yield was observed from additional extractions.
- NIST used a relative response factor approach to calibration.
- Homogeneity was evaluated for each determined mycotoxin as a function of packaging order, sample preparation order, and chromatographic run order.
- FDA evaluated sample size variability between 1 g and 5 g sample sizes.

#### Analysis of mycotoxins by NIST ID-LC-MS/MS Instrumentation: Agilent Series 1260 LC / Applied MS Source Parameters:

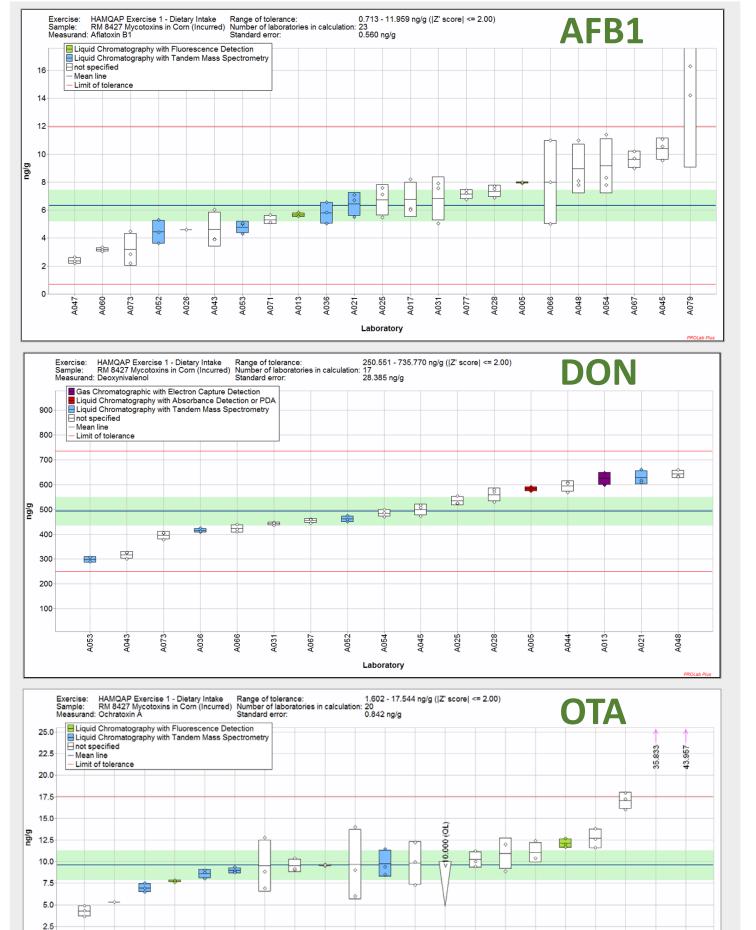
Collision Gas (CAD) Biosystems SCIEX API 5000 LC-MS/MS system, electrospray ionization in positive ion mode. Curtain Gas (CUR) MRM transitions and Compound MS Parameters: lon Source Gas 1 (GS1) 50

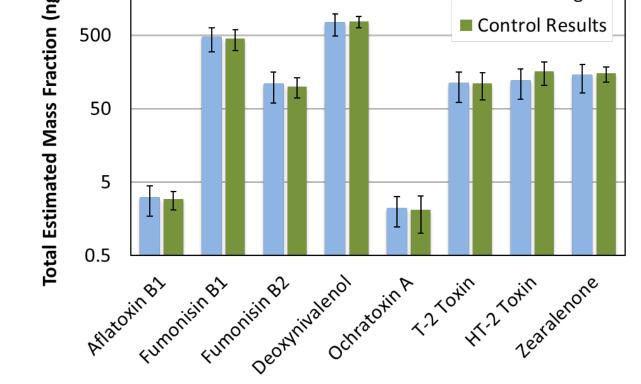
			<u>.</u>				•••••		
Mycotoxins	Molecular formula	Molecular weight	RT (min)	Adduct ion	MRM Transitions	DP (eV)	CE (eV)	CXP (eV)	Ion Source Gas (GS2)
Aflatoxin B1	C17H12O6	312.1	9.2	[M+H] <sup>+</sup>	313.1→241.0/285.0	107/85	53/35	13/15	
[13C17]-aflatoxin B1	$^{13}C_{17}H_{12}O_6$	329.1	9.2	[M+H] <sup>+</sup>	330.1→255.2/301.0	107/85	53/35	13/15	IonSpray Voltage (IS)
Aflatoxin B <sub>2</sub>	$\mathbf{C}_{17}\mathbf{H}_{14}\mathbf{O}_{6}$	314.1	9.0	[M+H] <sup>+</sup>	315.2→287.1/259.1	105/110	38/43	16/14	ionspiay voltage (15)
<sup>13</sup> C <sub>17</sub> ]-aflatoxin B <sub>2</sub>	${}^{13}C_{17} H_{14}O_6$	331.1	9.0	$[M+H]^+$	332.0→303.2/273.1	105/110	38/43	16/14	
Aflatoxin G <sub>1</sub>	$C_{17} H_{12}O_7$	328.1	8.7	[M+H] <sup>+</sup>	328.8→243.2/200.0	93/93	39/58	13/10	Temperature (TEM)
[ <sup>13</sup> C <sub>17</sub> ]-aflatoxin G <sub>1</sub>	${}^{13}C_{17} H_{12}O_7$	345.1	8.7	$[M+H]^+$	345.8→257.1/124.2	80/80	38/92	13.5/8	
Aflatoxin G <sub>2</sub>	C <sub>17</sub> H <sub>14</sub> O <sub>7</sub>	330.1	8.5	$[M+H]^+$	331.2→313.0/245.0	88/100	36/42	18/13	Entrance Detential
[ <sup>13</sup> C <sub>17</sub> ]-aflatoxin G <sub>2</sub>	${}^{13}C_{17} H_{14}O_7$	347.1	8.5	$[M+H]^+$	348.0→330.0/259.0	88/100	36/42	18/13	Entrance Potential
Deoxynivalenol	$C_{15}H_{20}O_{6}$	296.1	5.5	$[M+H]^+$	297.0→249.0/231.2	65/64	17/20	20/13	
[13C15]-deoxynivalenol	$^{13}C_{15}H_{20}O_6$	311.2	5.5	$[M+H]^+$	312.0→263.0/245.2	65/64	17/20	20/13	
Fumonisin B <sub>1</sub>	C34H59NO15	721.4	10.3	$[M+H]^+$	722.5→352.5/334.5	150/100	45/45	15/15	<b>Injection Volume:</b> 3 µL
[ <sup>13</sup> C <sub>34</sub> ]-fumonisin B <sub>1</sub>	13C34H59NO15	755.5	10.3	$[M+H]^+$	756.4→374.5/356.4	107/110	52/55	20/20	<u>injection volume.</u> 5 µL
Ochratoxin A	C20H18CINO6	403.1	11.0	$[M+H]^+$	404.0→239.0/102.0	74/73	36/101	13/17	
[ <sup>13</sup> C <sub>20</sub> ]-ochratoxin A	13C20H18CINO6	423.1	11.0	$[M+H]^+$	424.1→250.1/110.1	74/73	36/101	13/17	
T-2 toxin	$C_{24}H_{34}O_9$	466.2	10.6	$[M+NH_4]^+$	484.3→215.2/185.1	55/50	26/30	12/9	
[13C24]-T-2 toxin	$^{13}C_{24}H_{34}O_9$	490.3	10.6	$[M+NH_4]^+$	508.3→229.2/198.2	55/50	26/30	12/9	12344
Zearalenone	$C_{18}H_{22}O_5$	318.1	11.0	$[M+H]^+$	319.2→283.2/187.1	50/50	19/29	16/10	8
[13C18]-Zearalenone	$^{13}C_{18}H_{22}O_5$	336.2	11.0	$[M+H]^+$	337.2→138.2/124.0	45/35	70/77	10/21	1.15e4
									1.10+4

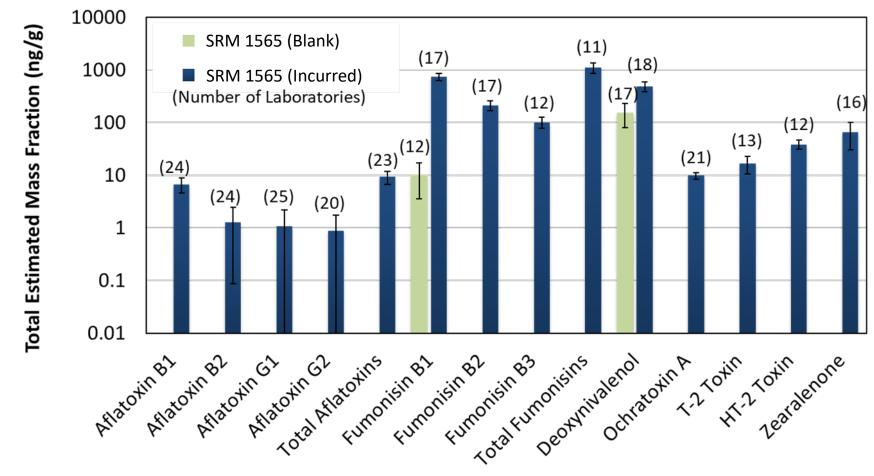
**Column:** Phenomenex Kinetex XBC18 LC column  $(100 \times 2.1 \text{ mm i.d.}, 2.6 \mu \text{m particles})$  with a  $10 \times$ 2.1 mm guard cartridge, 40 °C Mobile Phase A: 10 mmol/L ammonium formate in water containing 0.1 % formic acid (v/v)Mobile Phase B: 10 mmol/L ammonium formate in methanol containing 0.1 % formic acid (v/v)



- Summary of ID-LC-MS/MS Measurements by NIST and 30 **FDA/CFSAN**
- 50 • All values obtained for the QC material using the ID-LC-MS/MS 5000 500
  - method were within the value range assigned by the manufacturer.
    - NIST ID-LC-MS/MS measurement precision varied between 7.5 % and 41 % RSD in duplicate preparations of 10 samples of incurred and blank SRM 1565.
  - Based on FDA ID-LC-MS/MS measurement data (1), 1 g sample size is recommended for SRM 1565 (Blank). Using 5 g aliquots decreased measurement precision for deoxynivalenol (17 % to 35 % RSD) and fumonisin B1 (11 % to 23 % RSD). 5 g sample size is recommended for the remaining mycotoxins







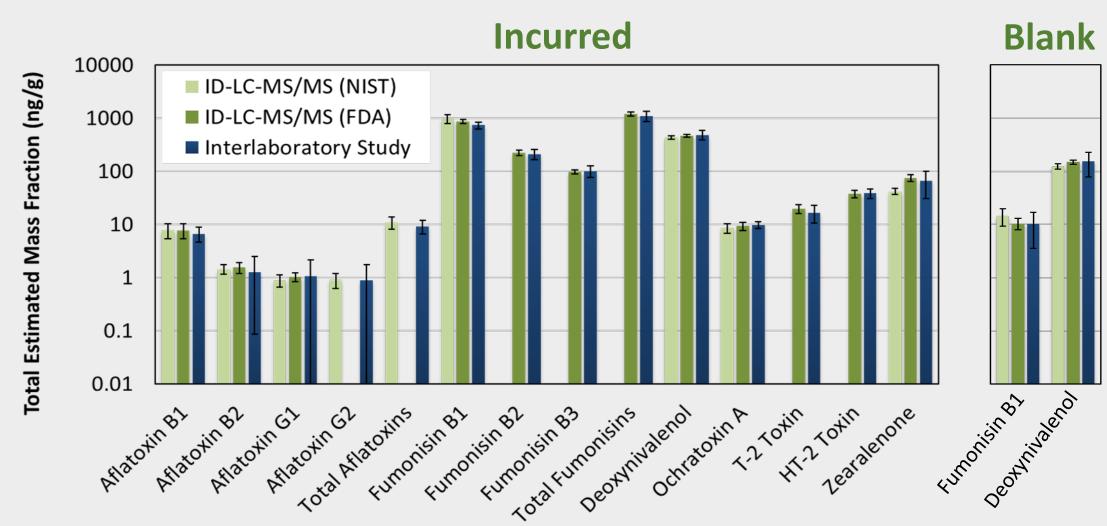
#### **Results from Interlaboratory Comparison**

- Data evaluated for quality based on the result provided for the control sample from FAPAS. Data from poor results of the control was excluded from the consensus data.
- Overall performance was good with between laboratory variabilities were comparable and ranged from 16 % to 49% for most mycotoxins in SRM 1565 (Incurred).
- Higher between-laboratory variability was observed for lower-level mycotoxins in the materials: aflatoxin G2 (82 %) in SRM 1565

	20.0	5				measurement challenges for some la
	12.0	5	NIST ID-LC-MS/MS.	identity criteria.		
	11.5	100	mycotoxins in SRM 1565 by	• FDA did not provide data for aflatoxin G2 based on failed		of the fumonisins in the blank r
	10.0	100	Sample chromatogram of	C		samples, except for total fumonisins
	2.0	40	000 // 60 60 70 80 90 100 110 120 Time (min)	• FDA data for ochratoxin A was not used for value assignment.	in SRM 1565 Mycotoxins in Corn (Incurred).	<ul> <li>Consensus means were within the tage</li> </ul>
	0	5	000 00- 1000 00-	fumonisins, and HT-2 toxin.	B1 (AFB1), Deoxynivalenol (DON), Ochratoxin A (OTA)	SRM 1565 (Blank).
LC Gradient:	Time (min)	<u>%B</u>	AFG2 0000 0000 00000	• NIST did not provide data for fumonisin B2, fumonisin B3, total	Results from interlaboratory comparison for Aflatoxin	
Flow Rate: 0.3 r	0		-ArG1 Ar	is recommended for the remaining mycotoxins.	PROLeb Plus	, (Incurred), and deoxynivalenol (46

## **Combined Results**

Assigned values for mycotoxins in SRM 1565 were determined by combination of the means from NIST ID-LC-MS/MS and FDA ID-LC-MS/MS measurements and the median from qualified laboratories from the interlaboratory comparison using a linear pool method (2). The uncertainty represents the SD of the aggregate probability distribution.



	Mass Fraction (ng/g)						
Mycotoxin	SRM 1565 (Blank)	SRM 1565 (Incurred)					
Aflatoxin B1	ND	7.5 ± 1.7					
Aflatoxin B2	ND	1.43 ± 0.34					
Aflatoxin G1	ND	0.98 ± 0.19					
Aflatoxin G2	ND	0.87 ± 0.24					
Total Aflatoxins	ND	10.2 ± 2.9					
Deoxynivalenol	142 ± 36	467 ± 67					
Fumonisin B1	10.4 ± 3.9	805 ± 190					
Fumonisin B2	ND	217 ± 30					
Fumonisin B3	ND	99.3 ± 8.4					
Total Fumonisins	ND	1150 ± 170					
Ochratoxin A	ND	9.4 ± 1.2					
HT-2 Toxin	ND	38.2 ± 6.0					
T-2 Toxin	ND	18.4 ± 4.2					
Zearalenone	ND	61 ± 36					

### Future Work

- 16 %) and fumonisin B1 (84 %) in
- target ranges for all analytes in all ins in SRM 1565 (Blank). Low levels material may have resulted in laboratories.

### • With the availability of reference standards

- Values for DON and ZON will be upgraded to certified with SI traceability
- Values will be assigned for fumonisin B2, fumonisin B3, HT-2 toxin, and T-2 toxin
- Development of other matrix-matched foods contaminated with mycotoxins

#### **References:**

(1) Zhang, K., et al. (2017) J. Agric. Food Chem. 65, 7138–7152. doi:10.1021/acs.jafc.6b04872 (2) Koepke, A., Lafarge, T., Possolo, A., & Toman, B. (2017) Metrologia 54, S34–S62 (3) Phillips, M.M., et al. (2019) J. AOAC Int. 102, doi:https://doi.org/10.5740/jaoacint.19-0109 Acknowledgements: AJ Moors, J Trevillian, DL Ellisor (blending, packaging, particle size determination); KM Morehouse, L Cumberland (irradiation); CA Barber, BA Benner, JB Thomas, CQ Burdette, J Camara, SE Long, JA Murray, BJ Place, CA Rimmer, LJ Wood (HAMQAP); CQ Burdette (technical); H-K Liu (statistics); G Hahm (poster assembly) Disclaimer: Certain commercial equipment, instruments, or materials are identified to specify the experimental procedure adequately. Such identification is not intended to imply

recommendation or endorsement by the NIST, nor is it intended to imply that the materials or equipment identified are necessarily the best available for the purpose.