

# OSAC 2022-S-0011 Standards for Construction of Multilocus Databases

Wildlife Forensics Biology Biology Scientific Area Committee Organization of Scientific Area Committees (OSAC) for Forensic Science





## **Draft OSAC Proposed Standard**

## OSAC 2022-S-0011 Standards for Construction of Multilocus Databases

Prepared by Wildlife Forensics Biology Version: 1.0 October 2021

#### **Disclaimer:**

This OSAC Proposed Standard was written by the (subcommittee) of the Organization of Scientific Area Committees (OSAC) for Forensic Science following a process that includes an <u>open comment period</u>. This Proposed Standard will be submitted to a standards developing organization and is subject to change.

There may be references in an OSAC Proposed Standard to other publications under development by OSAC. The information in the Proposed Standard, and underlying concepts and methodologies, may be used by the forensic-science community before the completion of such companion publications.

Any identification of commercial equipment, instruments, or materials in the Proposed Standard is not a recommendation or endorsement by the U.S. Government and does not imply that the equipment, instruments, or materials are necessarily the best available for the purpose.

To be placed on the OSAC Registry, certain types of standards first must be reviewed by a Scientific and Technical Review Panel (STRP). The STRP process is vital to OSAC's mission of generating and recognizing scientifically sound standards for producing and interpreting forensic science results. The STRP shall provide critical and knowledgeable reviews of draft standards or of proposed revisions of standards previously published by standards developing organizations (SDOs) to ensure that the published methods that practitioners employ are scientifically valid, and the resulting claims are trustworthy.

The STRP panel will consist of an independent and diverse panel, including subject matter experts, human factors scientists, quality assurance personnel, and legal experts, which will be tasked with evaluating the proposed standard based on a comprehensive list of science-based criteria.

For more information about this important process, please visit our website at: <u>https://www.nist.gov/topics/organization-scientific-area-committees-forensic-science/scientific-technical-review-panels</u>.



## **Standards for Construction of Multilocus Databases**

#### 2 Foreword

- 3 This standard defines the minimum requirements that shall be met when developing allele
- 4 frequency and population genetic databases for wildlife forensics.
- 5 The composition of a database intended for use in population genetic analyses is critical for
- 6 accurate comparison among the individual subjects as well as statistically sound group
- 7 assignment (e.g. individual, relatedness, population, geographic source, taxonomic grouping).
- 8 Analysts must use their expert knowledge in assessing the scientific merit of results obtained
- 9 from analysis of allele frequency and population genetic data, and in the subsequent reporting of
- 10 these results.
- 11

1

- 12 Keywords: wildlife forensics, population database, population genetics, multilocus, DNA
- 13



14

#### **Table of Contents** 15

16	1	Scope	5
17	2	Normative References	5
18	3	Terms and Definitions	5-6
19	4	Requirements	6-8
20	5	Conformance	8
21	6	Annexes	8-9
22	Aı	nnex A (informative) Bibliography	9

#### 23 **1 Scope**

- 24 This document provides minimum standards to guide the construction of multilocus population
- 25 genetic databases. This document covers criteria for the identification and collection of samples,
- 26 inclusion of associated biological data, choice and evaluation of genetic markers, and standard
- 27 statistical evaluation of the reference database. This document does not cover specific
- 28 applications such as individual matching, familial matching, geographic assignment, or other
- 29 wildlife forensic techniques to evidence in wildlife forensic casework. This document only
- 30 applies to databases generated from reference samples and does not include evidence items.
- 31 These minimum standards are not intended to replace standards in ISO 17025 or additional
- 32 forensic laboratory standards, but are designed to guide laboratories that are working toward
- 33 meeting those standards. Notes throughout this document offer clarifications and examples of
- 34 how a lab may meet a specific standard.

#### 35 2 Normative References

- 36 ANSI/ASB Standard 019, Wildlife Forensics General Standards<sup>1</sup>
- 37 ANSI/ASB Standard 046, *Wildlife Validation Standards-STR Analysis*<sup>1</sup>
- 38 ANSI/ASB Standard 047, Wildlife Validation Standards-Validating New Primers for
- 39 Sequencing<sup>1</sup>
- 40 ANSI/ASB Standard 048, Wildlife Forensic DNA Standard Procedures<sup>1</sup>
- ANSI/ASB Best Practices Recommendations 114, Best Practice Recommendations for Internal
   Validation of Software used in Forensic DNA Laboratories<sup>1</sup>

#### 43 **3 Terms and Definitions**

44 For purposes of this document, the following definitions apply.

#### 45 **3.1**

- 46 assignment
- 47 a method for assigning individuals to predefined categories, based on a suite of characters (e.g.
- 48 multilocus genotype) measured for the individual and for samples from each category (e.g.
- 49 potential source populations).
- 50 **3.2**

#### 51 autocorrelation

- 52 phenomenon where samples closer in geographic space or time tend to be more similar or
- 53 dissimilar to each other than expected by chance alone, for a given variable such as allele
- 54 frequencies.
- 55

56 **3.3** 

#### 57 co-ancestry

58 identical alleles that are copies of the same ancestral allele without mutation; this is a subset of

- 59 identical by state (IBS).
- 60

### 61 **3.4**

#### 62 familial matching

a method for determining genetic family relationships such as paternity, siblingship and otherkinships.

#### 65 **3.5**

#### 66 individual matching

a method for comparing one DNA profile to another to determine if the DNA profiles are

- 68 consistent at the level of interest
- 69

#### 70 **3.6**

#### 71 **population**

a group of organisms of the same species in a defined geographic area, such that any pair of
 members can interbreed.

#### 74 **3.7**

#### 75 probability of identity

- the probability that two randomly drawn individuals within a given population will have
- identical genotypes at multiple loci (reference Waits et al 2013 or Peakall and Sykes 1996)

#### 78 79 **3**

## 79 3.880 probability of siblings

- 81 the probability that two randomly drawn individuals within a given population will have
- 82 identical genotypes at multiple loci when relatives are included in the sample.

#### 83 84 **3.9**

#### 85 statistical power

- 86 the probability of a hypothesis test finding an effect if there is an effect to be found.
- 87

#### 88 4 Requirements

- 89 The following requirements and recommendations address criteria for construction and
- 90 evaluation of population databases, including identification of database components, choice of
- 91 genetic markers, procedures for statistical analysis, and evaluation and interpretation of results
- 92 for general population genetic analyses. Species differ based on demographic, ecological, and
- 93 evolutionary factors, so quantitative values for the minimum number of individuals and genetic
- 94 markers needed for a reference database are expected to vary according to the species and
- 95 populations of interest. These criteria shall be addressed in laboratory validation studies
- 96 according to established population genetic theory and practice.

#### 97 4.1 Inclusion Criteria for Genetic Database Samples

99 100 101 102 103 104 105	<ul> <li>a. sample acquisition;</li> <li>b. establishment of parameters for inclusion of samples;</li> <li>c. validation process for use of genetic markers;</li> <li>d. criteria for individual sample data quality;</li> <li>e. quality control/curation of sample information and genetic data</li> </ul> 4.1.2 Quality control shall include adherence to standards in ANSI/ASB 019, ANSI/ASB 046.		
106	ANSI/ASB 047, and ANSI/ASB 048		
107	<b>4.1.3</b> In determining database composition the laboratory shall assess, at minimum:		
108 109 110 111 112 113	<ul> <li>a. sample size needed to accurately represent source population genetic diversity;</li> <li>b. related taxonomic information, including <ol> <li>presence of subspecies</li> <li>evolutionary significant units (ESU)</li> <li>hybrids in the species group of interest</li> <li>geographic range of the taxa in question</li> </ol> </li> </ul>		
114 115	<b>4.1.4</b> Criteria shall be established for metadata associated with sample acquisition, to include at minimum:		
116 117 118 119 120 121 122	<ul> <li>a. geographic location of source samples (e.g. sampling location, breeding location, location of death);</li> <li>b. sex of individual, if known;</li> <li>c. age class of individual, if known;</li> <li>d. type of tissue sampled (e.g. fresh tissue, blood, bone, hair, antler, keratin, feces, etc);</li> <li>e. collection information (i.e. date, collector - both name and agency/institution, method of collection).</li> </ul>		
123	4.1.5 At minimum, genetic markers shall be evaluated for the:		
124	a. number of loci required, as determined by laboratory validation.		
125 126	NOTE: The number of loci needed will vary by species/population and forensic application (e.g. individual matching, population assignment, paternity)		
127 128 129 130 131 132 133 134	<ul> <li>b. genotyping error rate;</li> <li>c. genetic diversity measures, including but not limited to: <ol> <li>Hardy-Weinberg Equilibrium,</li> <li>linkage disequilibrium,</li> <li>allelic richness,</li> <li>allelic diversity,</li> <li>heterozygosity measures within and among populations;</li> </ol> </li> <li>d. presence of null alleles.</li> </ul>		

**4.1.1** Protocols for constructing genetic databases shall include:

- 4.1.6 Quality criteria shall be established for sample inclusion when adding genetic data tospecies/population databases. This shall include, at minimum:
- 137 a. minimum acceptable completeness of genotype per sample;
- b. minimum genotype quality measures depending on genotyping platform [e.g. capillary
- electrophoresis Relative Fluorescence Units (RFU); Next-generation sequencing genotype quality score and read depth].
- 141 **4.2** Once constructed, the database shall be evaluated for:
- 142 a. representative geographic coverage;
- b. power to discriminate species/population boundaries;
- c. power to identify natural groupings that are ecologically or biologically meaningful;
- 145 d. population level allele frequencies;
- e. presence of spatial or temporal autocorrelation;
- 147 f. sex-related bias;
- g. estimates for statistical power (i.e. probability of identity, probability of siblings);
- h. presence of duplicated samples;
- i. level of co-ancestry
- Note: Database composition will vary based on forensic application (e.g. individual matching,familial matching, geographic assignment techniques).
- 153
- **4.3** Laboratories shall have protocols for evaluation of software intended for use in statistical
- analysis, including commercial programs and programs developed in-house.
- 4.4 Once initially validated, databases augmented with new samples or subsetted shall be re-evaluated as in 4.2.
- 158 **4.5** Laboratories shall have protocols for database archival and version control.

#### 159 5 Conformance

- 160 Conformance to the standards outlined in this document is measured by the availability of
- 161 written documentation in the form of formal protocols and methods available for examination.
- 162
- 163
- 164
- 165
- 166
- 167
- 168
- 169
- 170
- 171
- 172

173	Annex A
174	(informative)

174 175

#### 175 176

## Bibliography

177

178	The following information	provides a list of the literature	resources that may assist in defining
1,0			

179 the breadth and scope of this standard. This list is not meant to be all inclusive. The laboratory

180 shall develop a list tailored to its specific needs. Updated references shall be added to the

181 laboratory's list as new methods or technologies are incorporated into the laboratory's protocols.

182	1]	J.M. Butler. Fundamentals of Forensic DNA Typing. Elsevier Academic Press, San Diego, CA, 2010.
183	2]	Legendre P and Legendre L (1998) Numerical Ecology. 3rd ed. Elsevier, Amsterdan.
184	3]	Ogden, R and A. Linacre 2015, Wildlife forensic science: A review of genetic
185		geographic origin assignment. Forensic Science International: Genetics, 18:152 - 159.
186		https://doi.org/10.1016/j.fsigen.2015.02.008
187	41	Rousset F., 2004. Genetic Structure and Selection in Subdivided Populations. Princeton
188	L	University Press, Princeton, NJ.
189	5]	Slatkin, M, and H. E. Arter. Spatial Autocorrelation Methods in Population Genetics. The
190		American Naturalist, 138(2): 499–517. JSTOR, www.istor.org/stable/2462484. Accessed
191		11 Mar. 2021.
192	6]	Smouse, P., R. Peakall. Spatial autocorrelation analysis of individual multiallele and
193		multilocus genetic structure. Heredity 82:561–573 (1999).
194		https://doi.org/10.1038/sj.hdy.6885180
195	7]	Waits, L. P, G Luikart, P Taberlet 2001 Estimating the probability of identity among
196	-	genotypes in natural populations: cautions and guidelines. Molecular Ecology, 10(1):
197		249-256. https://doi.org.10.1046/j.1365-294x.2001.01185.x
198	8]	Waples, R. S. 1995. Evolutionarily significant units and the conservation of biological
199		diversity under the endangered species act: Evolution and the aquatic ecosystem: defining
200		unique units in population conservation. American Fisheries Society Symposium, vol. 17:
201		8-27. https://ci.nii.ac.jp/naid/10021852122/en/
202		
202		
203		
204		
201		
205		
206		
207		
207		