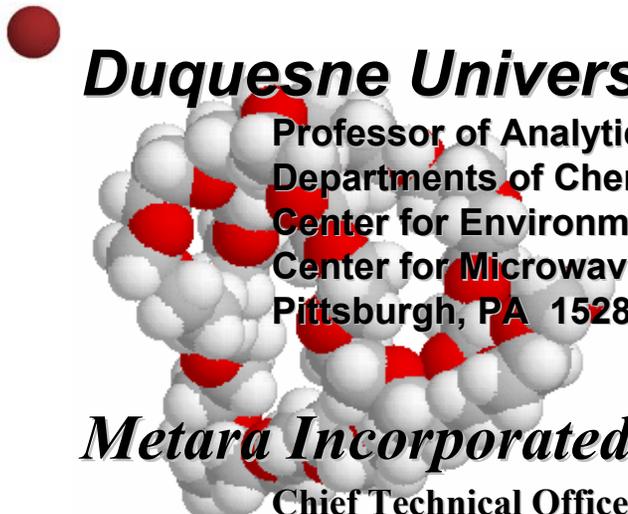


# Automated and Near Real-time, Trace Contamination and Chemical Species Analysis for the Semiconductor Industry

H.M. 'Skip' Kingston  
March 27, 2003

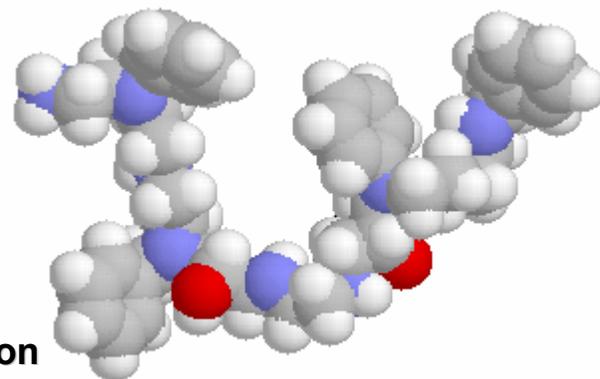


***Duquesne University,***

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# Overview

- The “problems” as seen by a metrologist
- Metrology for contamination analysis in Semiconductor Fabrication
- New enabling technology needed
- Inventions required to impact the problem
- Analytical chemistry metrology tools
  - Standard methods and new methods
  - IDMS, IPMS, SIDMS
- Speciation and why it is important
  - “Chemically Significant Data”
    - Bob Helms, SEMATECH – ULSI 3/25/02
    - Thomas Theis, IBM-ULSI 3/25/02
- Time relevant inclusive data – initial tests



# The Problem:

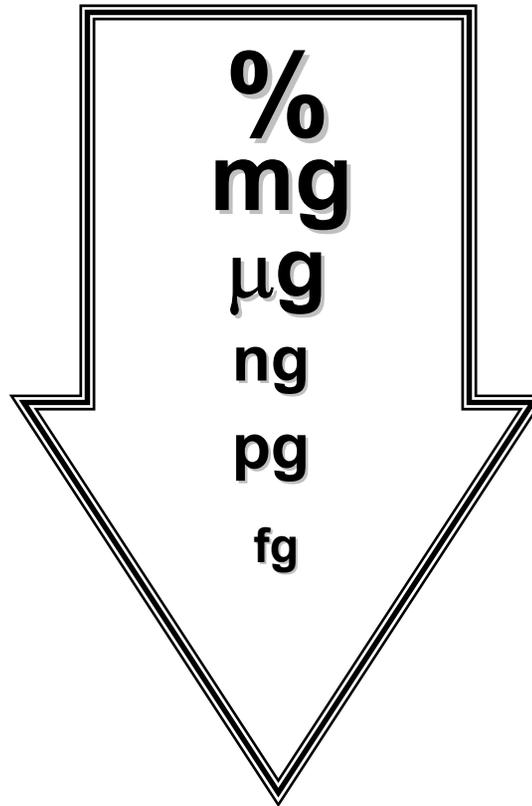
**At these dimensions Contamination is a critical parameter in the function and assembly of Semiconductors and Nanotechnology devices**

- ***Nanotechnology*** - As devices become 3 to 20 atoms in size, nanotechnology and micromachines predict contamination determines the viability of the device
- ***Semiconductor*** - As semiconductor line widths go to  $<0.13$  micrometers contamination become a critical limitation of the performance of these devices



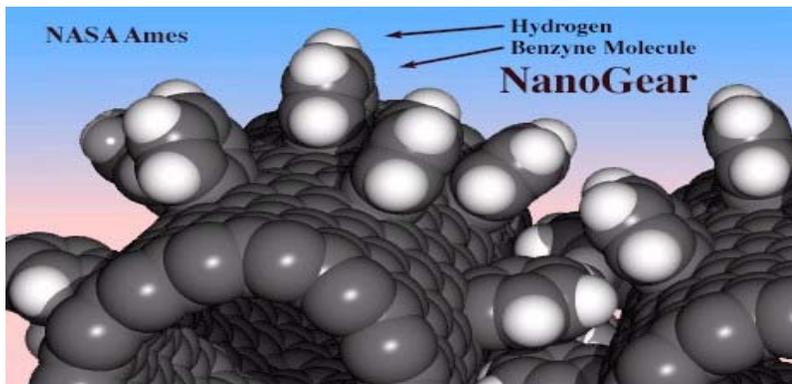
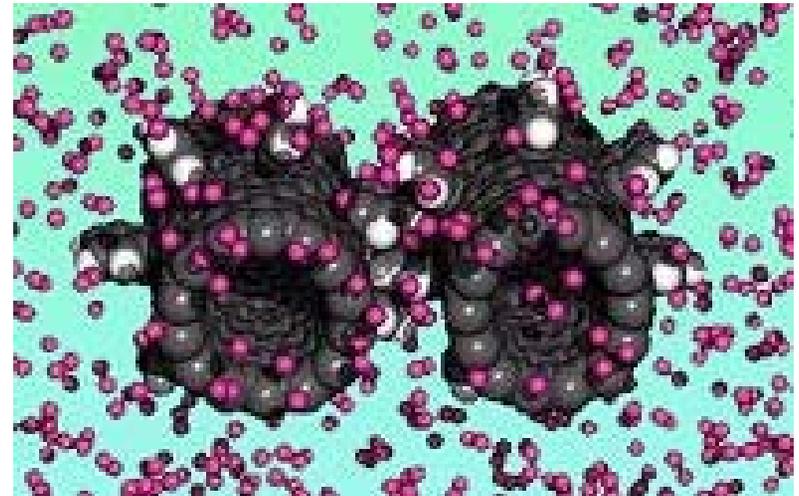
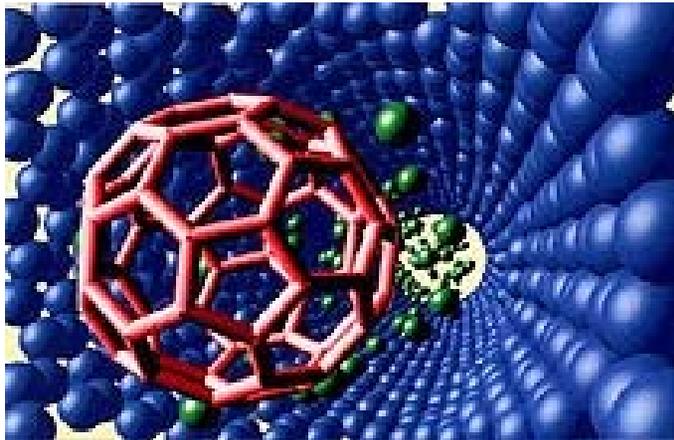
**The Relationship:** As trend in ultra-trace element analysis go lower, previous techniques become inadequate and new methods are required to maintain or achieve accuracy & precision

**“New Thresholds Require New Methods”**



# Nanotechnology

How much contamination does it take to act as a monkey wrench in the gears at the molecular level?



What is Nanotechnology?

NanoMachines

...are so small, they "look" bumpy - the "bumps" are individual atoms.

## ***Semiconductor Road Map for Development*** **Important Industry Directions!**

**“Key Area of the 2001 edition of the ITRS focus on the yield model and defect budget, defect detection and characterization, yield learning, and wafer environment contamination control.”**

Reference: “Examining upcoming yield enhancement challenges in the 2001 roadmap” Micro, February 2002



# The Challenge?

**“The most critical challenge is to find ways to determine the effects of trace impurities on device performance and yield.”**

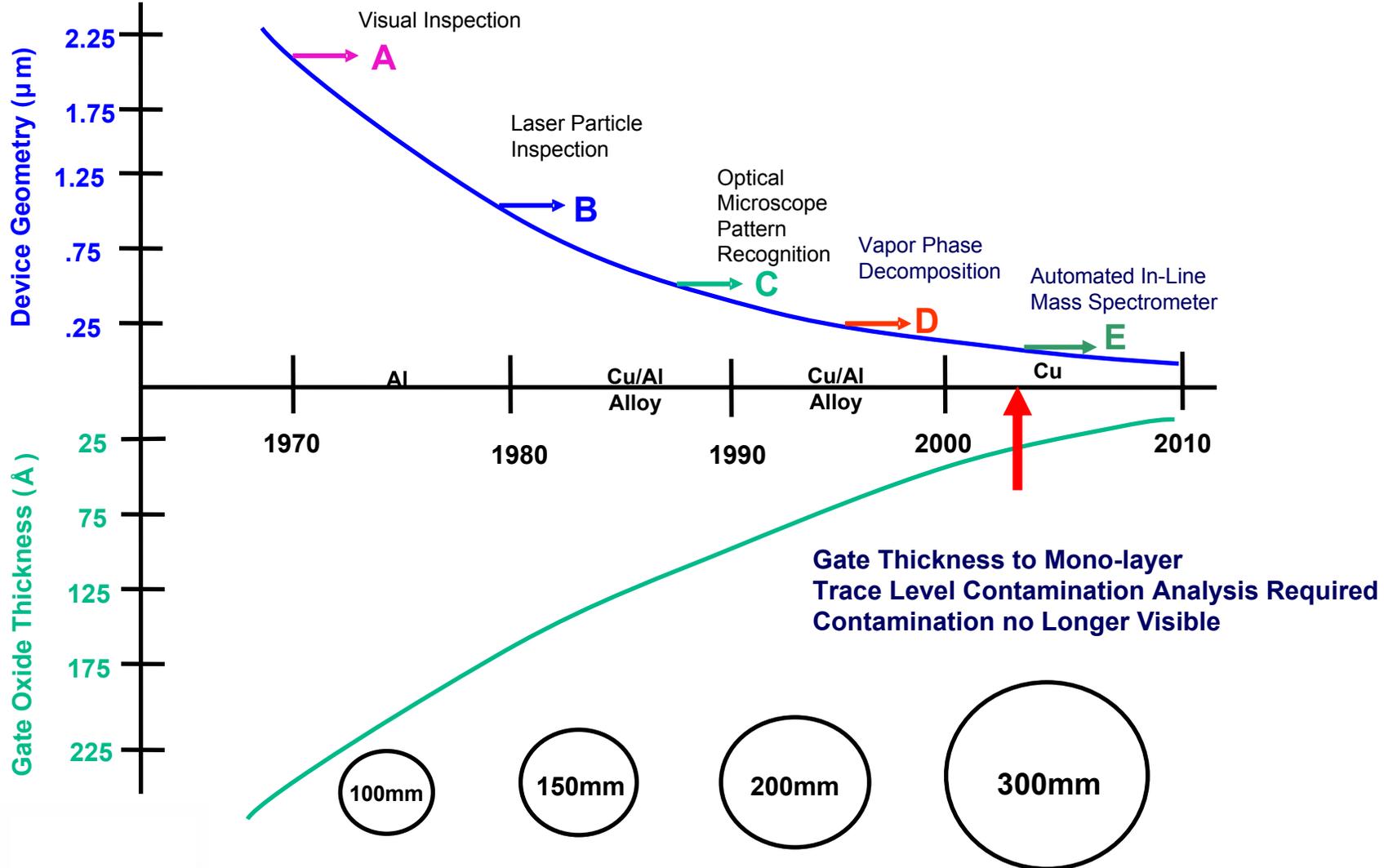
Reference: “Examining upcoming yield enhancement challenges in the 2001 roadmap”  
- *Micro*, February 2002

Authors: Christopher Long, IBM;  
Milton Godwin, Applied Materials;  
Manuela Huber, Sematech/Infineon;  
Richard Jarvis, Sematech/AMD  
Fred Lakhani, Sematech



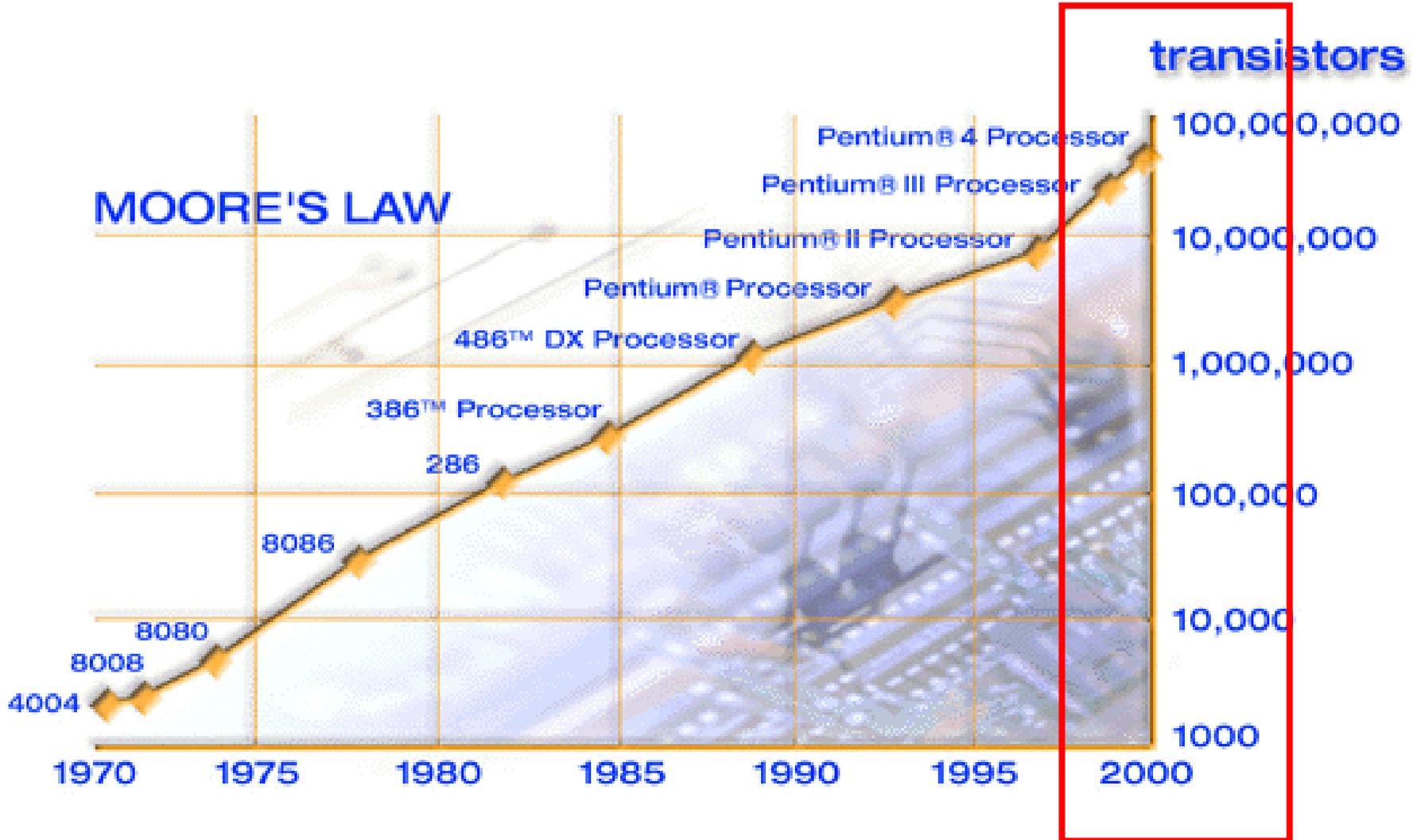
# Moore's Law & Drives Contaminate Analysis

Device dimensions increase importance and implementation of contamination measurement metrology

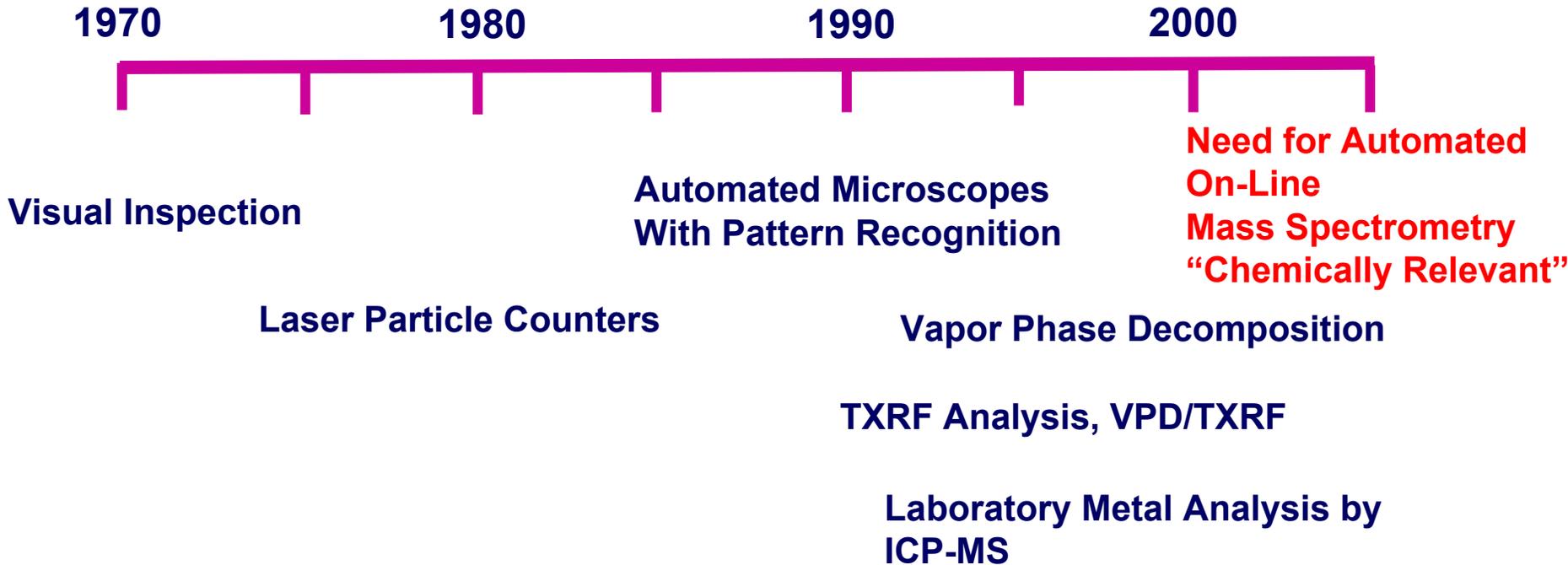


# Moore's Law

Analytically  
Significant  
Threshold



# History of Contaminate Analysis for Semiconductors



critical nature of contamination metrology



# State-of-the-art: mid 2000

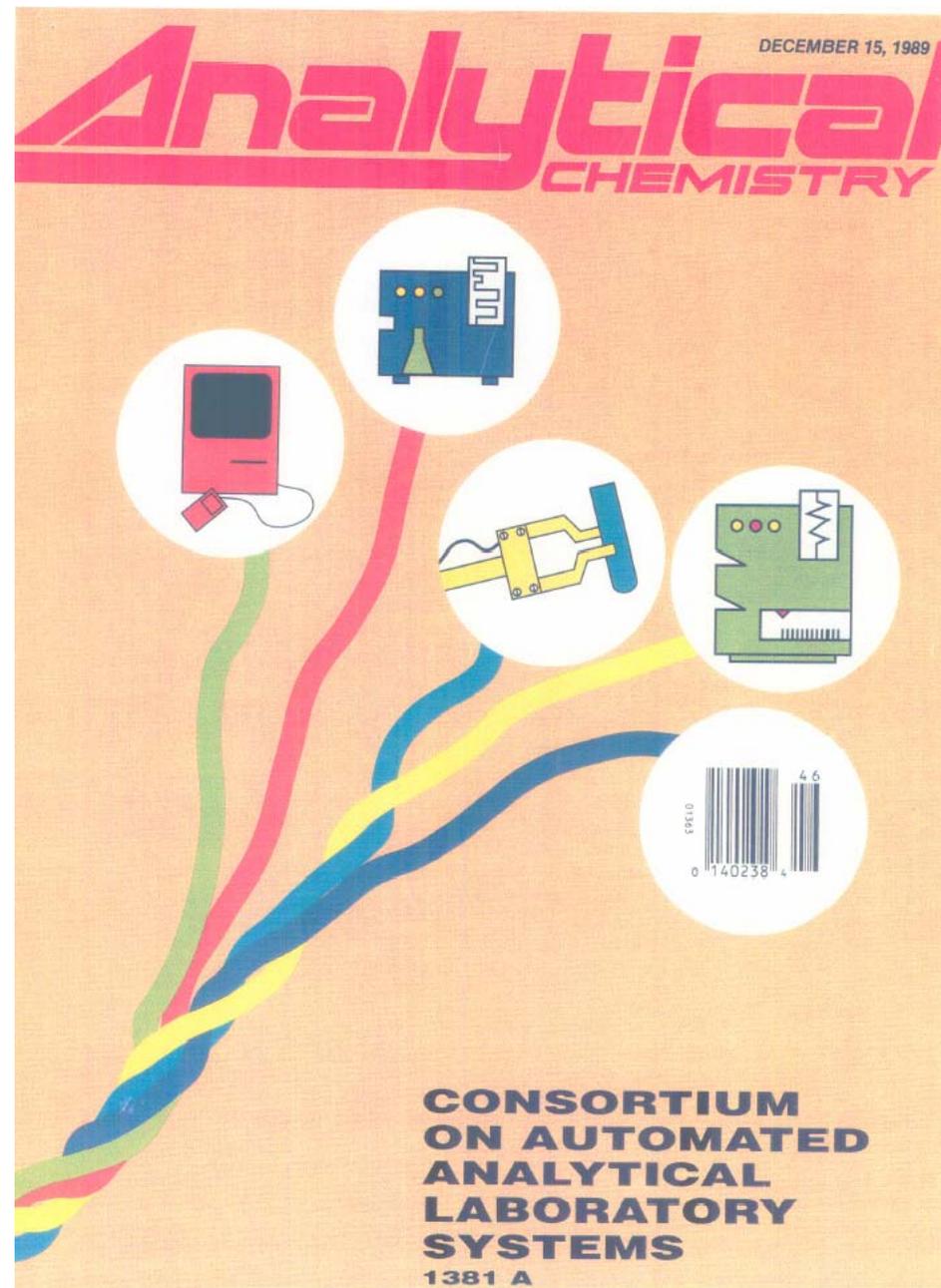
## Project Begins

- **Elemental metal analysis primarily**
  - Few anion, organic and no species information
- **Species evaluation not considered feasible**
- **6-12-24-48 hour sample to data turnaround is usual for metal analysis in semiconductor fabs**
- **Lack of on-line metrology for liquids**
- **Up to 60 liquid cleaning steps required in some advanced wafer processing**
- **Constituent analysis needs emerging eg. Cu ECD**



U.S. Industries had before addressed a selected set of these critical metrology issues in an industry government consortium known as CAALS:

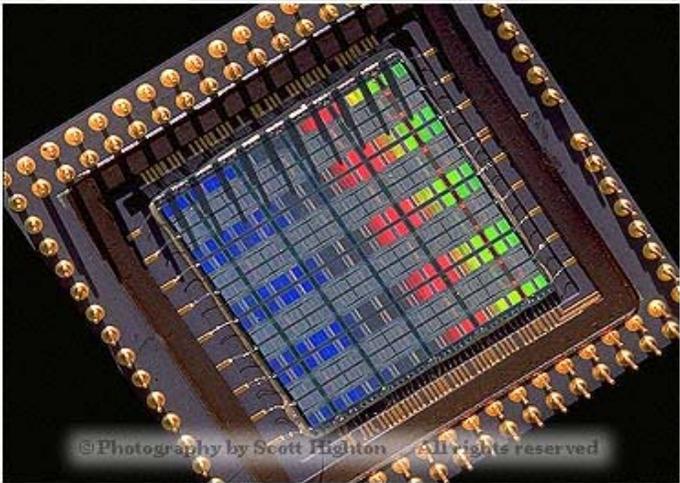
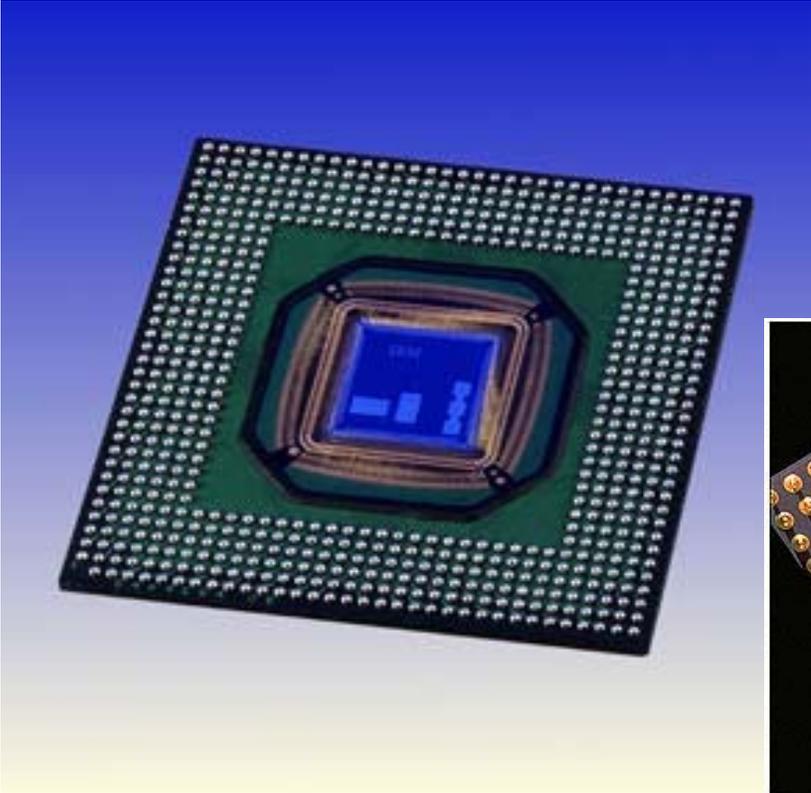
- NIST
- Hewlett-Packard
- Perkin-Elmer
- Dupont
- Union Carbide
- Kodak
- Department of Energy
- Zymark
- et. al.



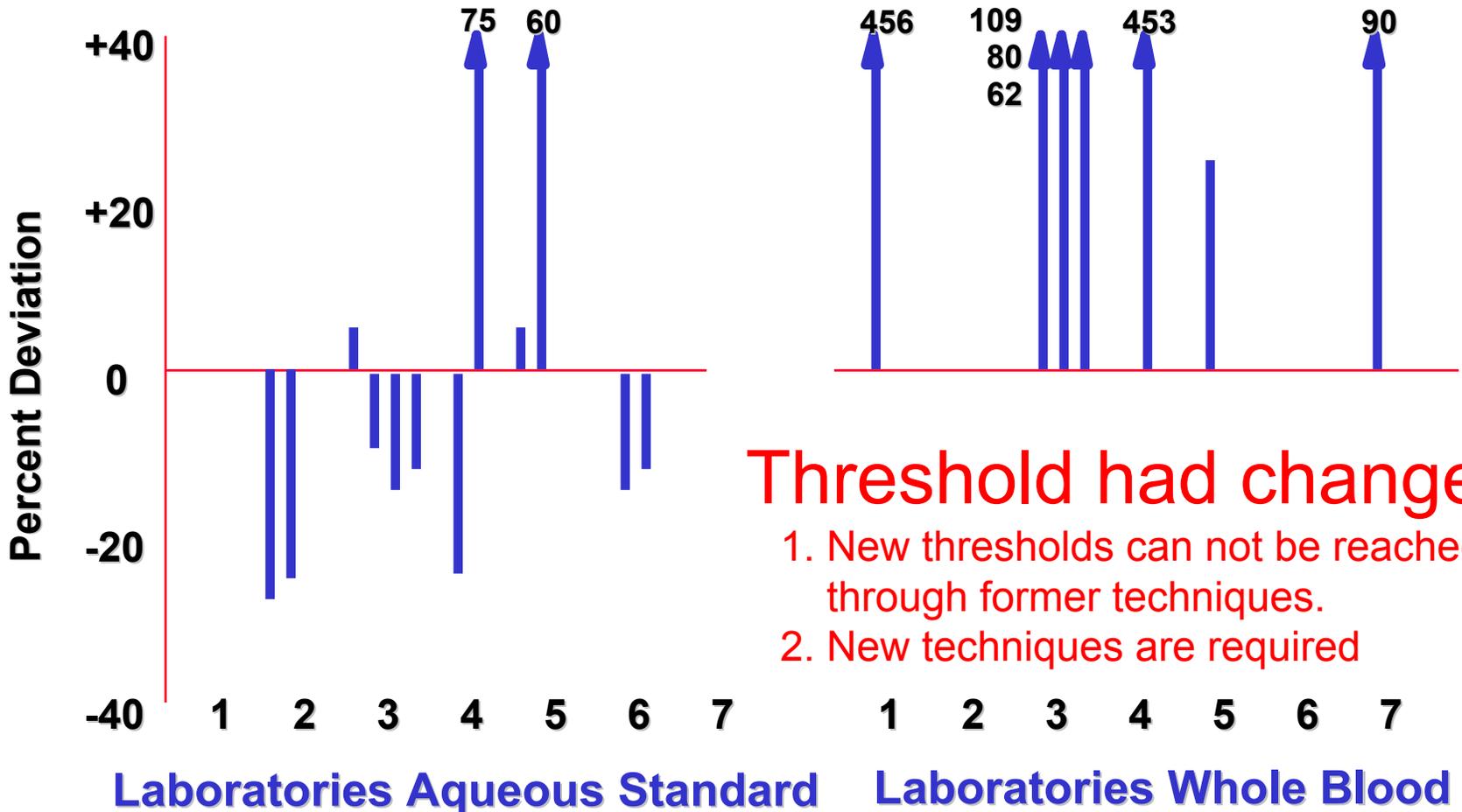
| Item                                   | ICP-MS  | Desired Technology  |
|--|---|---|
| Target Application                     | Lab applications  | Real-time, In-line, In-fab: Trace contamination monitoring of standard aqueous semiconductor solutions      |
| Sample Matrix                          | UPW, SC1, SC2, IPA, DHF, HNO <sub>3</sub> , H <sub>2</sub> O <sub>2</sub> | UPW, SC1, SC2, IPA, DHF, HNO <sub>3</sub> , H <sub>2</sub> O <sub>2</sub>                                   |
| Monitors                               | Depends Upon Available Standards<br>Cations (+)<br>Elements<br>Inorganics | 20 - 28 Trace Contaminants<br>Cations (+) and anions (-)<br>Elements and species<br>Inorganics and Organics |
| Sample Preparation                     | Manual  | Automated   |
| Limit of Detection                     | < 20 ppt  | < 20 ppt  |
| Calibration and Quantitation Standards | Calibration Curves  | Direct Analysis, Automated Calibration<br>NIST traceable  |
| Sample Volume                          | 5 mL  | <5 mL   |
| Technology                             | ICP-MS  | Based on Mass Spectrometer  |
| Throughput                             | 24 to 72 hours  | 5-10 sample/hr (5 min/sample)   |
| Accuracy                               | ± 25% at the quantitation limit   | <± 25% at the quantitation limit  |



# Demanding Technology Requires Demanding Metrology



# Example: Comparison of Interlaboratory results of an Aqueous Standard and Whole Blood



**Threshold had changed!**

- 1. New thresholds can not be reached through former techniques.
- 2. New techniques are required

Reproduced from "The Role of the Analytical Blank in Accurate Trace Analysis" by T. J. Murphy, National Bureau of Standards Special Publication 422, Accuracy in Trace Analysis: Sampling, Sample Handling, and Analysis, Proceedings of the 7th IMR Symposium, Oct. 7-11, 1974, Pub. 1976.



# ***New Technology Required***

## **An Unconventional Instrument:**

**An Automated real-time integrated sample preparation and analysis system for inorganics, organics, and species, for the majority of cleaning and process solutions**

**What to do?**

**Invent and Engineer a New Technology**



# Recall – ITRS Challenge #2

“The most critical challenge is to find ways to determine the effects of trace impurities on device performance and yield.”

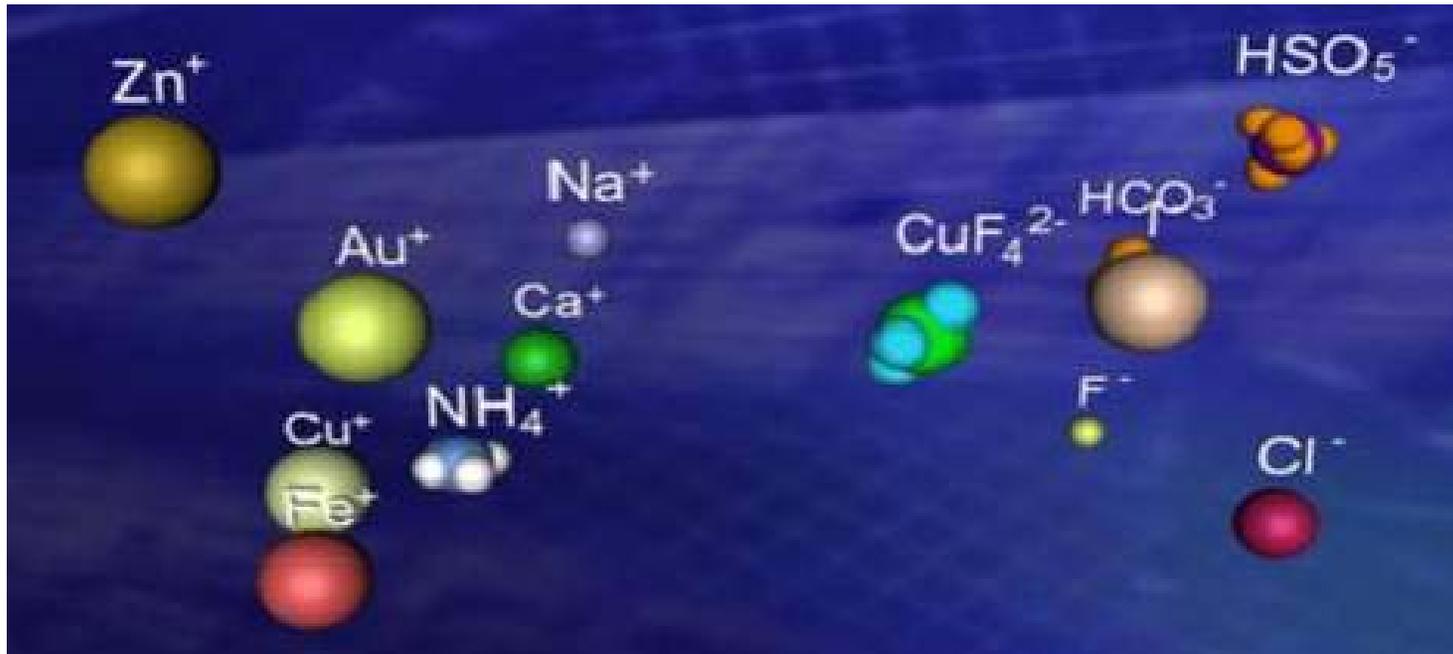
i.e. Statistically valid correlation and modeling  
i.e. “Chemically” significant determinations

Reference: “Examining upcoming yield enhancement challenges in the 2001 roadmap”  
- *Micro*, February 2002

Authors: Christopher Long, IBM;  
Milton Godwin, Applied Materials;  
Manuela Huber, Sematech/Infineon;  
Richard Jarvis, Sematech/AMD  
Fred Lakhani, Sematech



# Cations(+) & Anions(-)



Cations

Anions

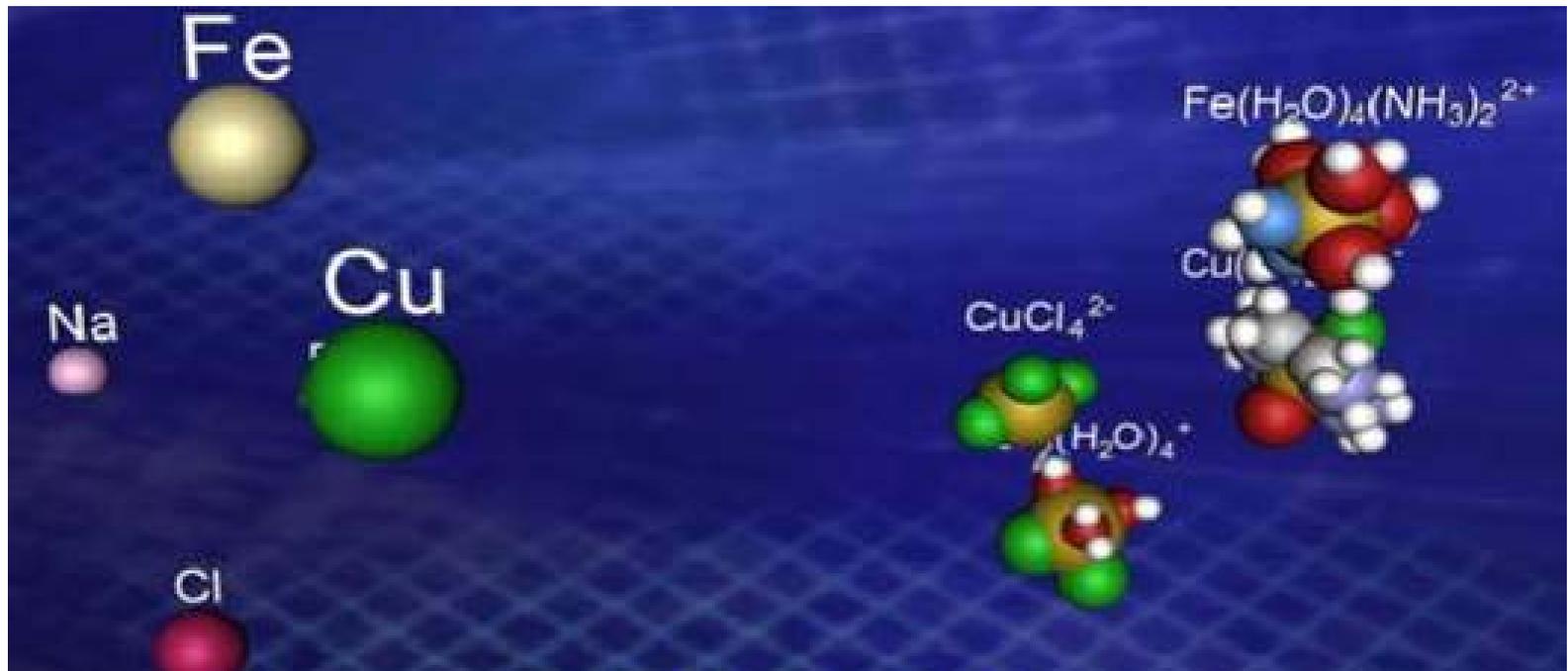
# *Elements & Species:*

**Elemental Species**

**Organic Molecules**

**Inorganic Complex Species**

**Organic Complex Species**



Elements

Species

# Why Have Mass Spectrometers Not Been Used in On-Line, Real Time, Multi-matrix Rapid Sequential Analysis?

- **Manual calibration requirement**
- **Lack of long term stability**
- **Matrix interferences**
- **Lack of instrument sensitivity in some matrices**
- **Inappropriate and unstable MS ionization sources**
- **Unavailability of fully automated systems**
- **Lack of applicable sample preparation integrated and automation with the mass spectrometer**

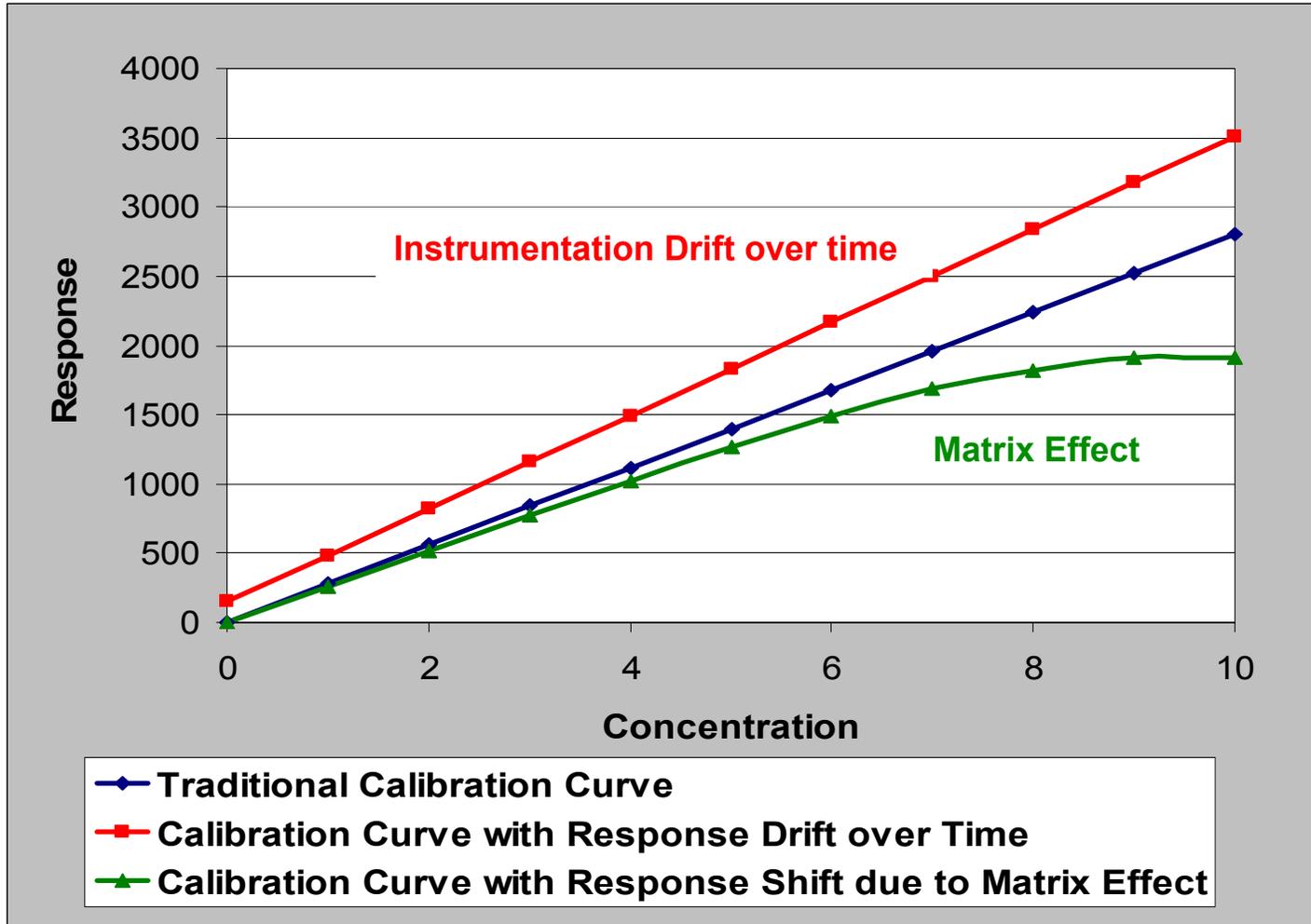


# Mass Spectrometer Calibration

- **Mass Spectrometer Drift**
- **Requires Continual Calibration**
- **Method of Calibration Must be Reliable**
- **Method Desired to be Matrix Independent**
- **Calibration Solutions Should Inexpensive**
- **Method must be very fast**



# Quantitation: Traditional Calibration Curve



# **Example, Examination of One Approach: On-Line, Real-Time, Automated, Direct Quantization for Elements, Species and Complex Molecular Species**

- **Direct Quantization through On-Line **IDMS** and **SIDMS** as a new capability:**
  - In Process Mass Spectrometry or “**IPMS**”
- **Fundamental enabling technology**
- **Alternative and simultaneous measurement of elements, molecules, and molecular species**



## Questions to consider:

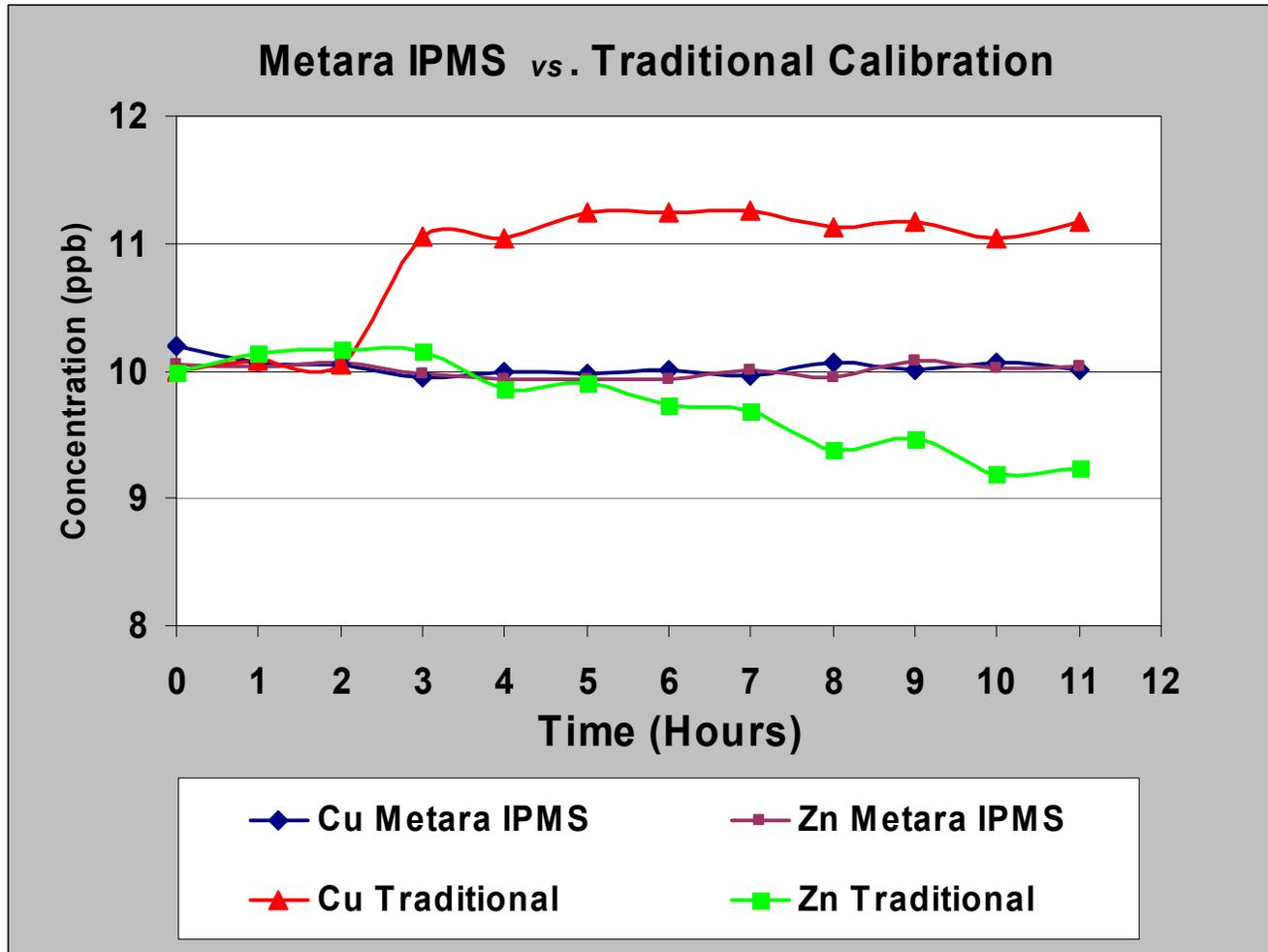
1. What new technology do you see that you do not expect?
2. Does technology Changes the way we do things?
3. Does it change what we call “normal”?
4. Do you think his son will use a drum?



# Mass Spectrometer Drift

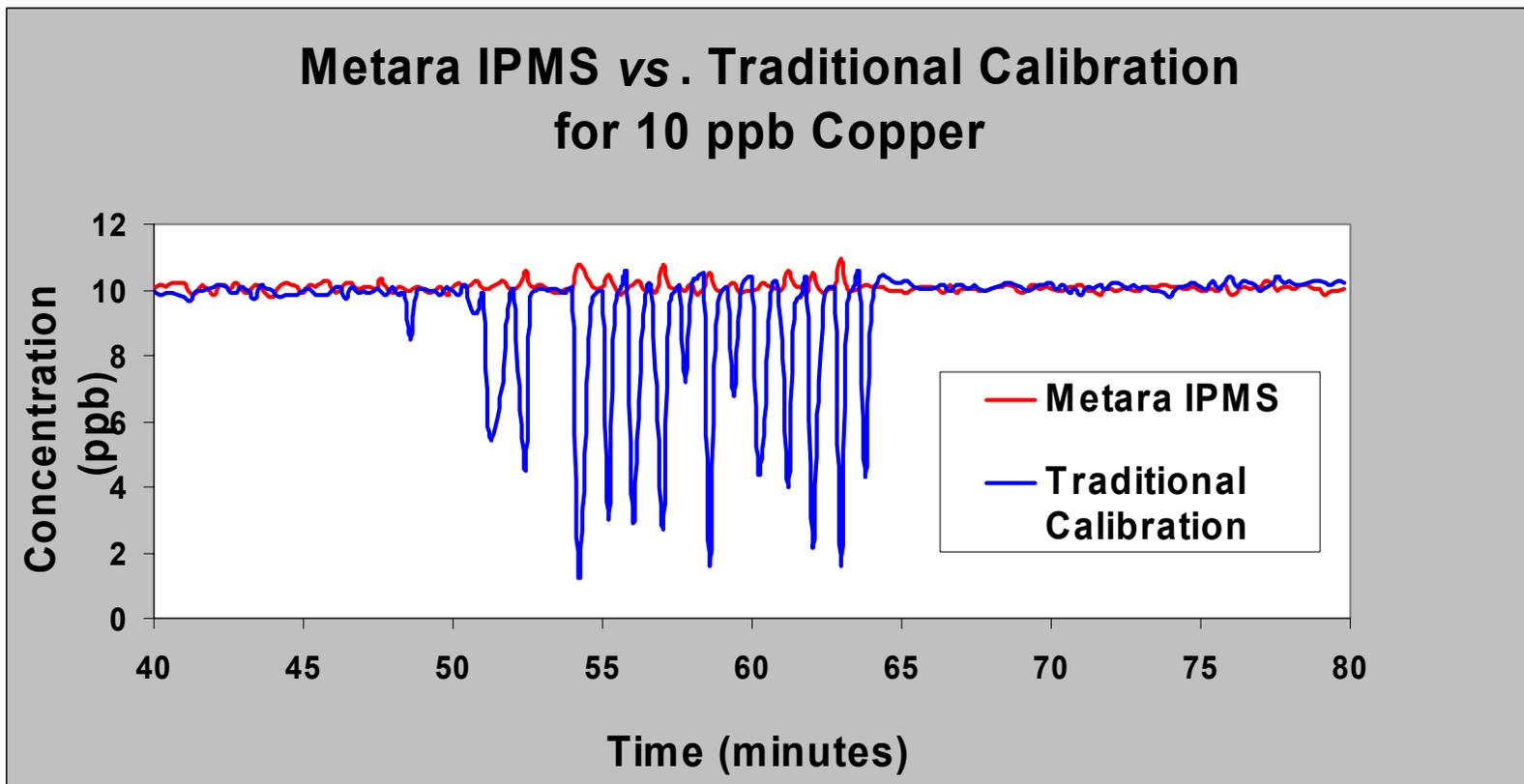
## Example: IPMS vs. Traditional -

### IPMS reduces or eliminates effects of mass spectrometer drift, enabling automation (patent pending)



# Mass Spectrometer Signal Stability

**Example: IPMS compensating for MS stability, retaining accuracy, and enabling automation** (patent pending)

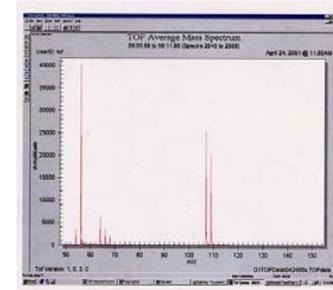
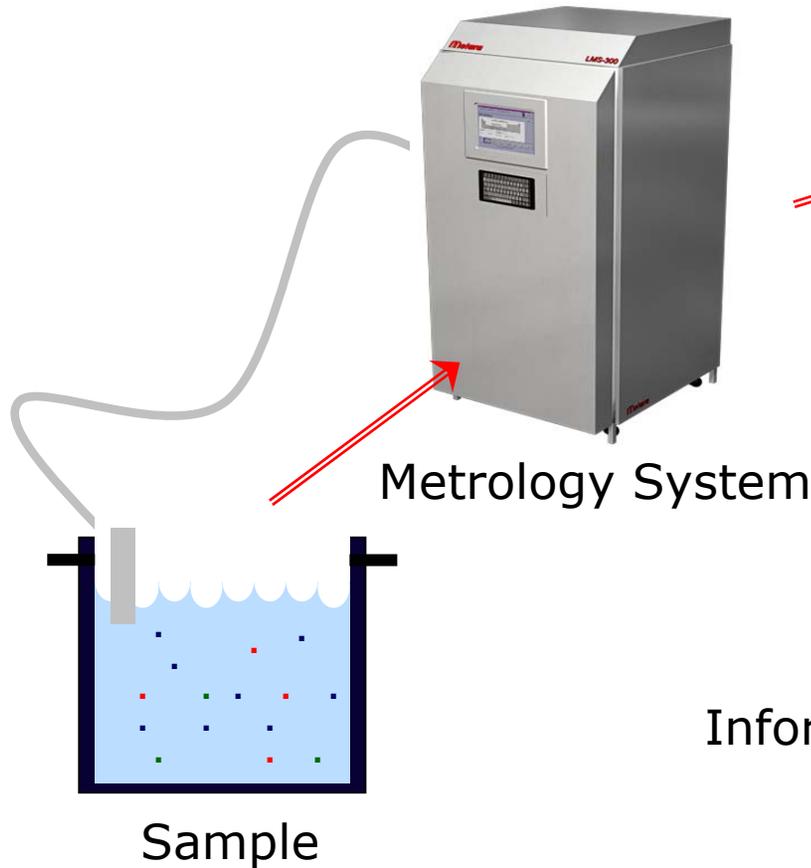


With a typical analytical instrument, air bubbles in the sample introduction system may cause a dramatic instability and/or decrease in signal.

*However,  $^{63}\text{Cu}$ :  $^{65}\text{Cu}$  isotope ratio measured by the IPMS Method remain constant.*



# Technology Overview



Data

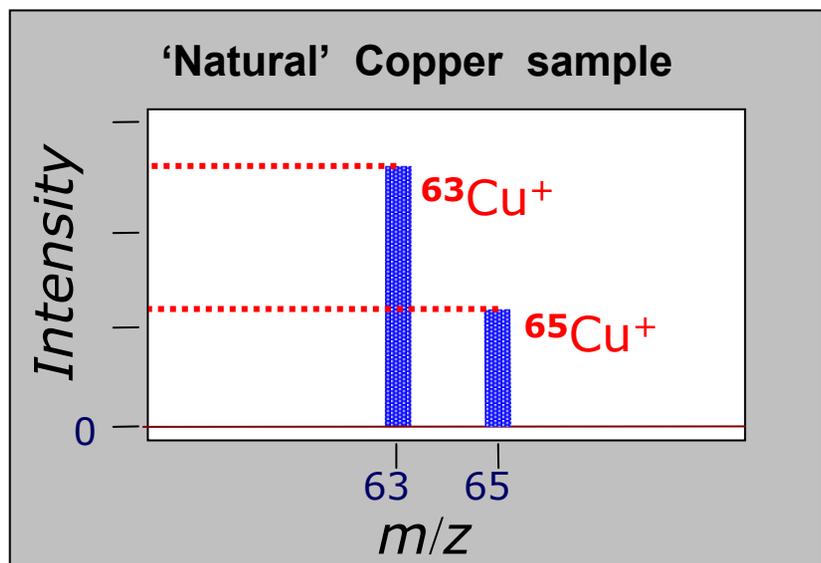
Information

| <u>Element</u>           | <u>Conc.</u>   |
|--------------------------|----------------|
| Ionic & Metallic         | 1.23 ppb       |
| <b>Organic Molecules</b> | <b>190 ppt</b> |
| <b>Species</b>           | <b>640 ppt</b> |

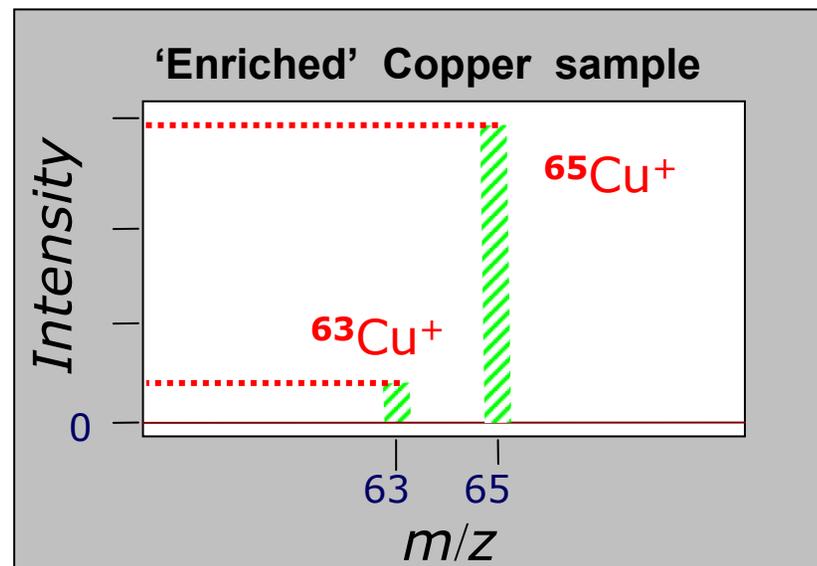
# Quantification Method

## Metara In-Process Mass Spectrometry (IPMS)

*e.g. Mass-Spectra of Copper Samples*



$^{63}\text{Cu}$  &  $^{65}\text{Cu}$  occur naturally in a 69:31 abundance ratio.

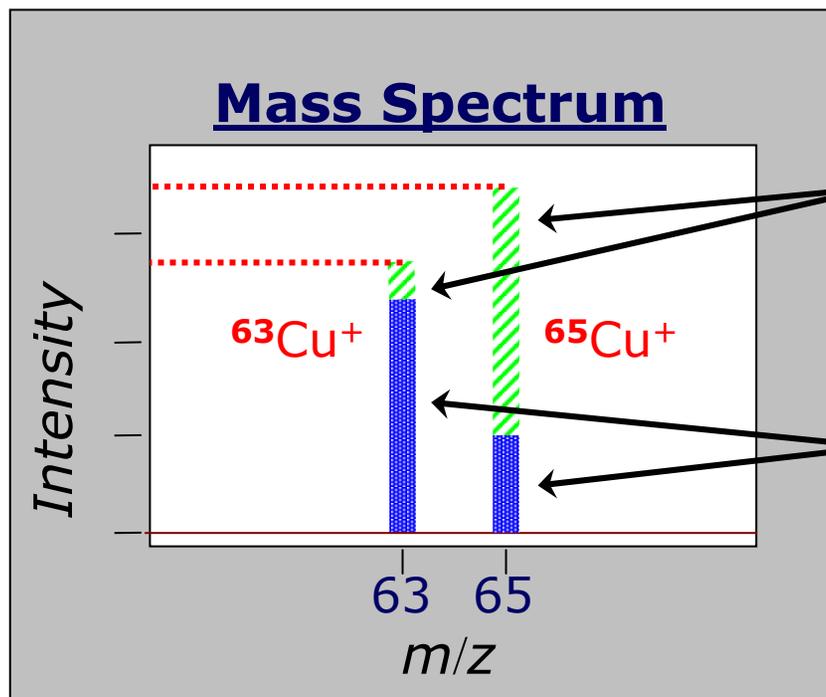


Isotopically-enriched solutions are commercially available (e.g. 5:95).

# Quantification Method

## Metara IPMS

*A sample of copper of unknown concentration is 'spiked' with a known amount of an isotopically-enriched standard, and introduced into the LMS-300 TCA.*

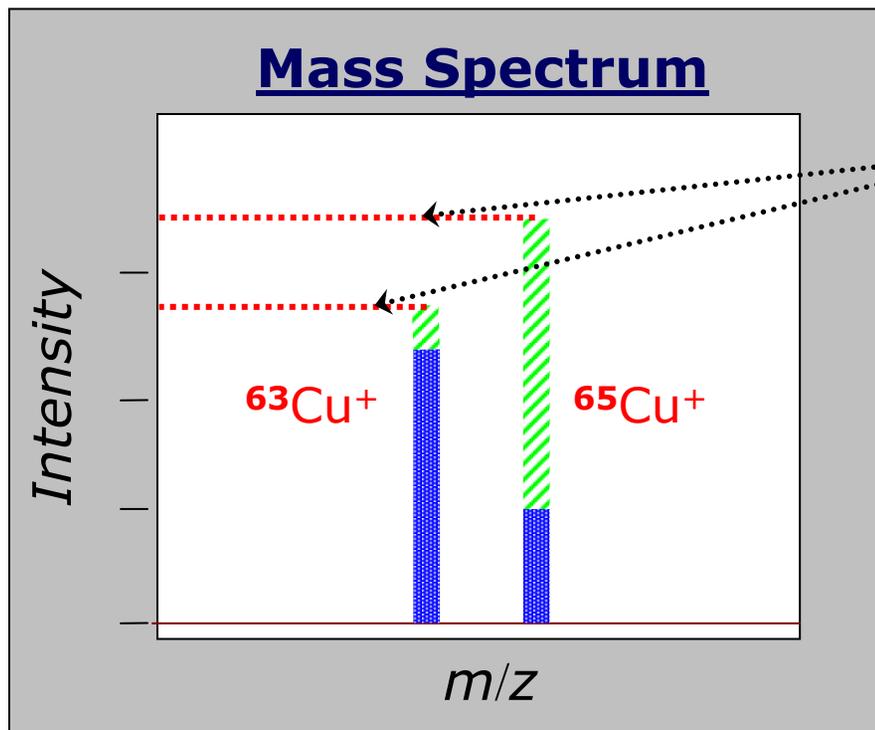


Contributions to signals from the spike.

Contributions to signals from the sample.

# Quantification Method

## Metara IPMS



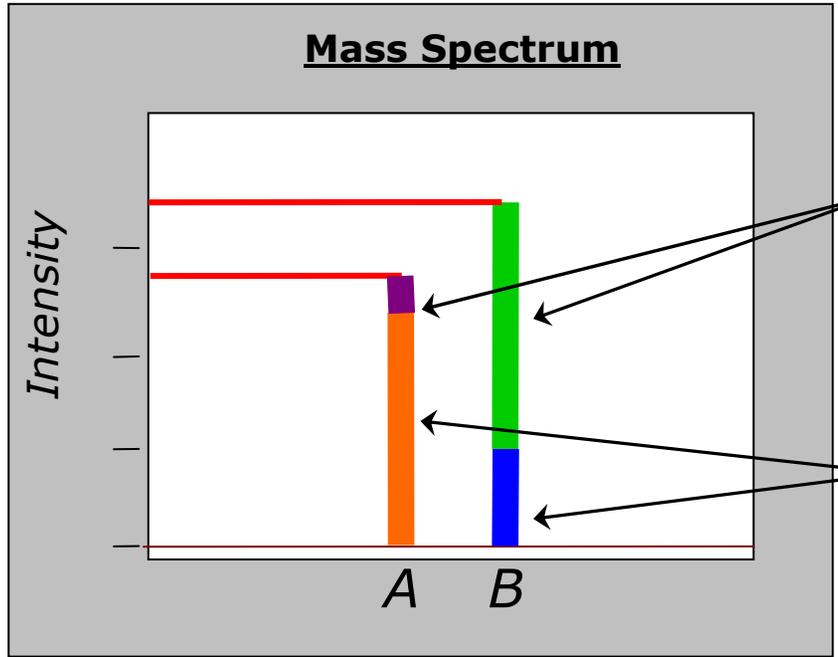
*The concentration of copper in the sample is deduced from the **RATIO** of the isotope signals...*

*This calculation is exemplified in a later example.*

| <u>Element</u> | <u>Conc.</u>   |
|----------------|----------------|
| Fe             | 1.23 ppb       |
| <b>Cu</b>      | <b>190 ppt</b> |
| Ni             | 640 ppt        |

# Metara IPMS Calculation

$$\text{Ratio} \left( \frac{\text{Isotope A}}{\text{Isotope B}} \right) = \frac{(\text{Amount of A from Sample} + \text{Amount of A from Spike})}{(\text{Amount of B from Sample} + \text{Amount of B from Spike})}$$



Contributions to the signals from the *Spike*.

Contributions to the signals from the *Sample*.

... cntd



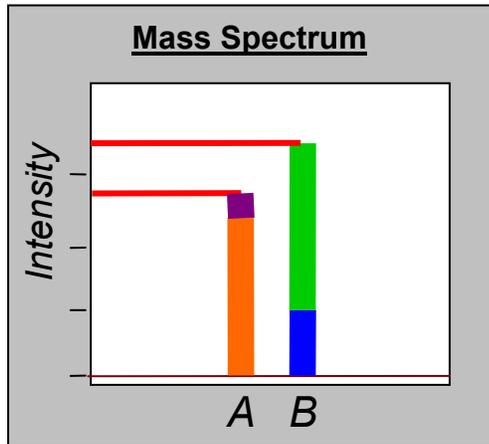
# Calculation & Formula

...cntd

$$\text{Ratio} \left( \frac{\text{Isotope A}}{\text{Isotope B}} \right) = \frac{(\text{Amount of A from Sample} + \text{Amount of A from Spike})}{(\text{Amount of B from Sample} + \text{Amount of B from Spike})}$$

So ...

$$\text{Ratio} = \frac{(A_s C_s V_s + A_{sp} C_{sp} V_{sp})}{(B_s C_s V_s + B_{sp} C_{sp} V_{sp})}$$



Where:

- $A_s$  = Fraction of isotope A in sample (natural)
- $B_s$  = Fraction of isotope B in sample (natural)
- $A_{sp}$  = Fraction of isotope A in spike (altered)
- $B_{sp}$  = Fraction of isotope B in spike (altered)
- $C_s$  = **Concentration of element in sample**
- $C_{sp}$  = Concentration of element in spike
- $V_s$  = Volume of the sample
- $V_{sp}$  = Volume of the spike

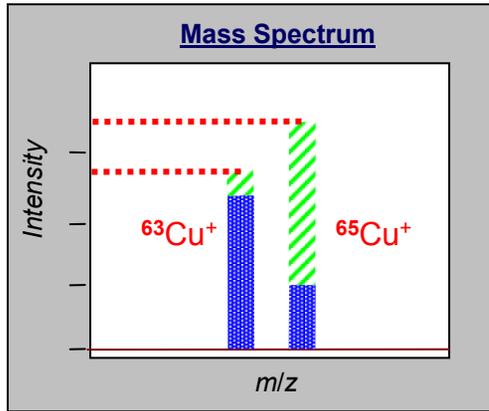
Known?

- ✓
- ✓
- ✓
- ✓
- ✗
- ✓
- ✓
- ✓

**Solve for  $C_s$ , (the concentration of the element in the sample) ...**



# Quantitation of Cu



## Known & Measured Values

- 1) The isotope ratio in the enriched spike standard,  
= 0.05 : 0.95 ( $^{63}\text{Cu}$  :  $^{65}\text{Cu}$ )      ( $A_{sp}$  &  $B_{sp}$ )
- 2) The concentration of the spike standard solution,  
= 0.88 ppb = 13.6 nmol/L      ( $C_{sp}$ )
- 3) The relative volumes of the spike and sample.  
= 0.13 (spike / sample)      ( $V_{sp} / V_s$ )
- 4) Measured Isotope Ratio  
= 0.82 ( $^{63}\text{Cu}$  /  $^{65}\text{Cu}$ )      (*Ratio*)
- 5) Natural Isotope Ratio  
= 0.692 : 0.308 ( $^{63}\text{Cu}$  :  $^{65}\text{Cu}$ )      ( $A_s$  &  $B_s$ )

... cntd

$$\text{Ratio} = \frac{(A_s C_s V_s + A_{sp} C_{sp} V_{sp})}{(B_s C_s V_s + B_{sp} C_{sp} V_{sp})}$$

Rearrange the equation to solve for  $C_s$

$$C_s = C_{sp} \left( \frac{V_{sp}}{V_s} \right) \frac{(A_{sp} - \text{Ratio} \times B_{sp})}{(\text{Ratio} \times B_s - A_s)}$$

$$C_s = 13.6 \text{ nmol/L} \times 0.13 \times \frac{(0.05 - 0.82 \times 0.95)}{(0.82 \times 0.308 - 0.692)}$$

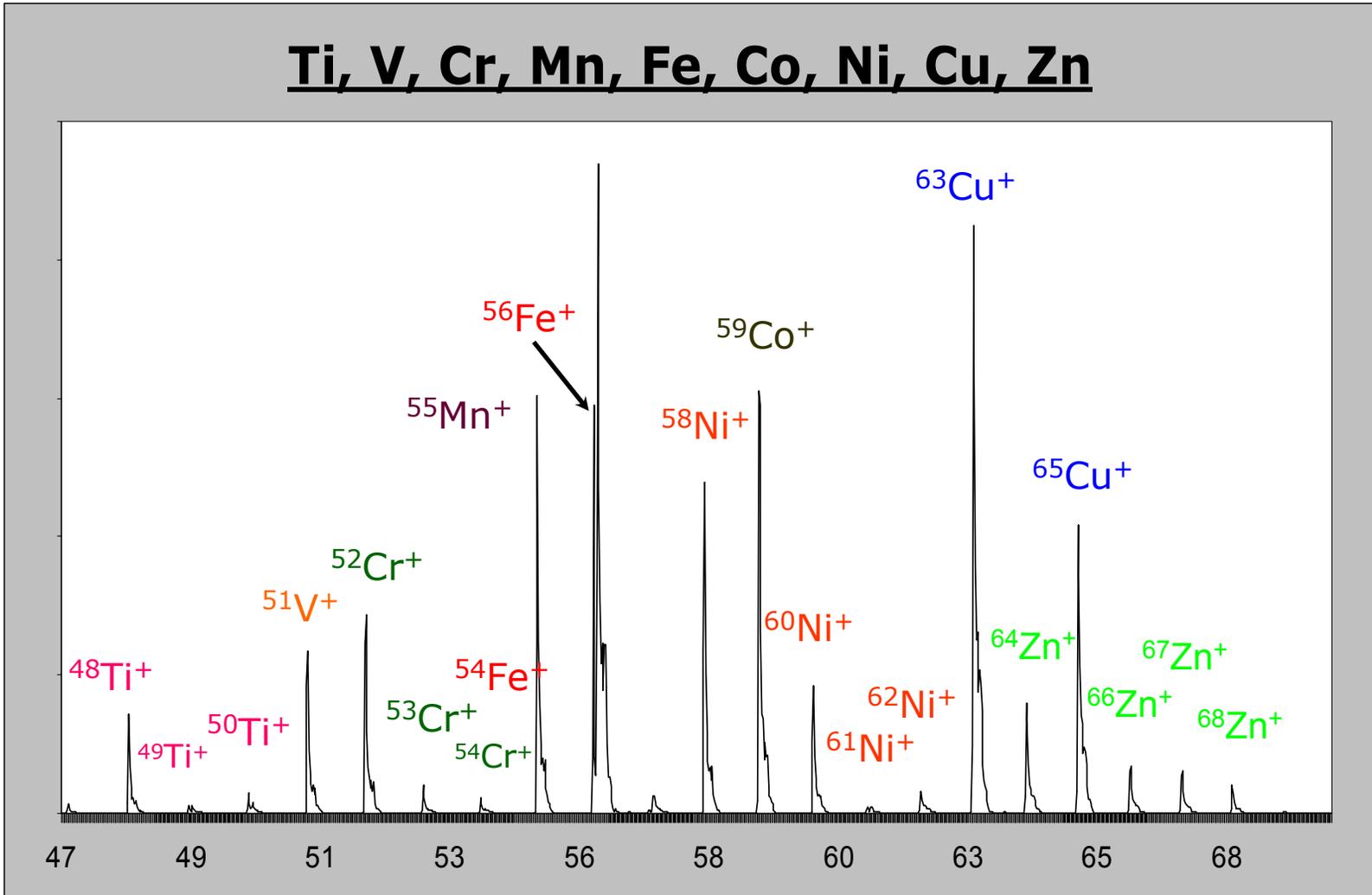
$$\underline{C_s = 2.93 \text{ nmol/L} = 0.186 \text{ ppb}}$$

So ...

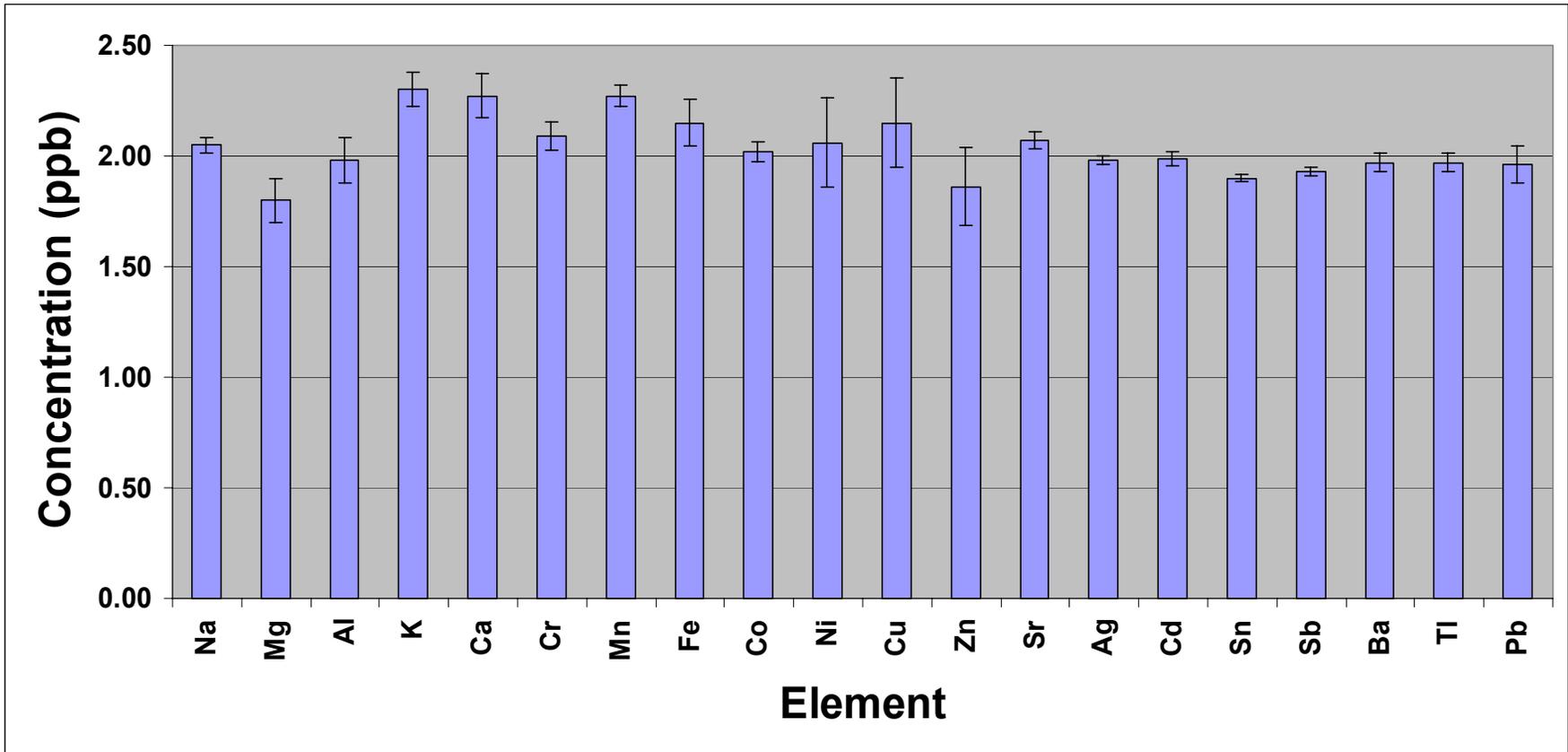
| <u>Element</u> | <u>Conc.</u>   |
|----------------|----------------|
| <b>Cu</b>      | <b>186 ppt</b> |

# First Row Transition Metals In **Hard Ionization Mode**

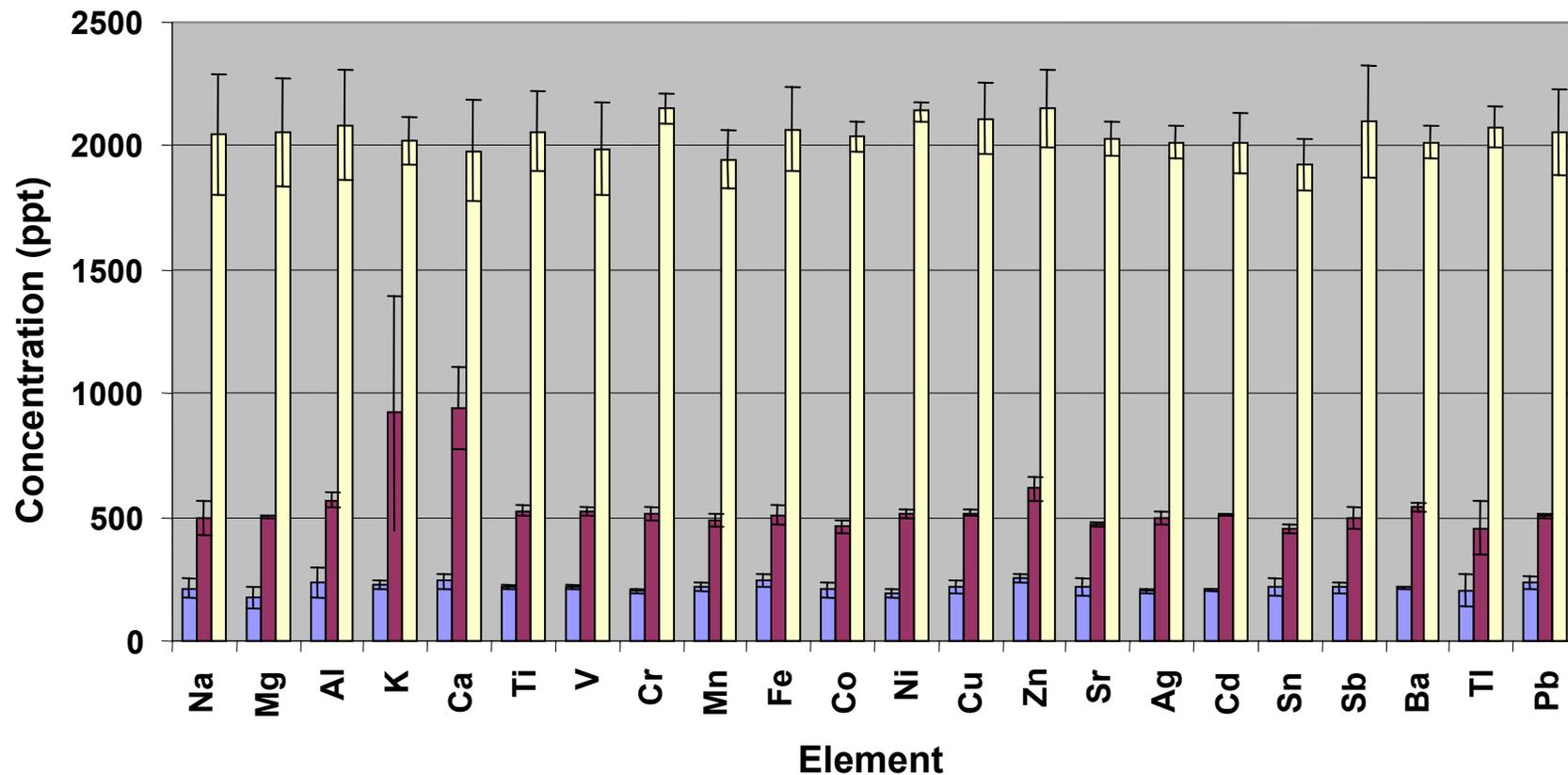
Ti, V, Cr, Mn, Fe, Co, Ni, Cu, Zn



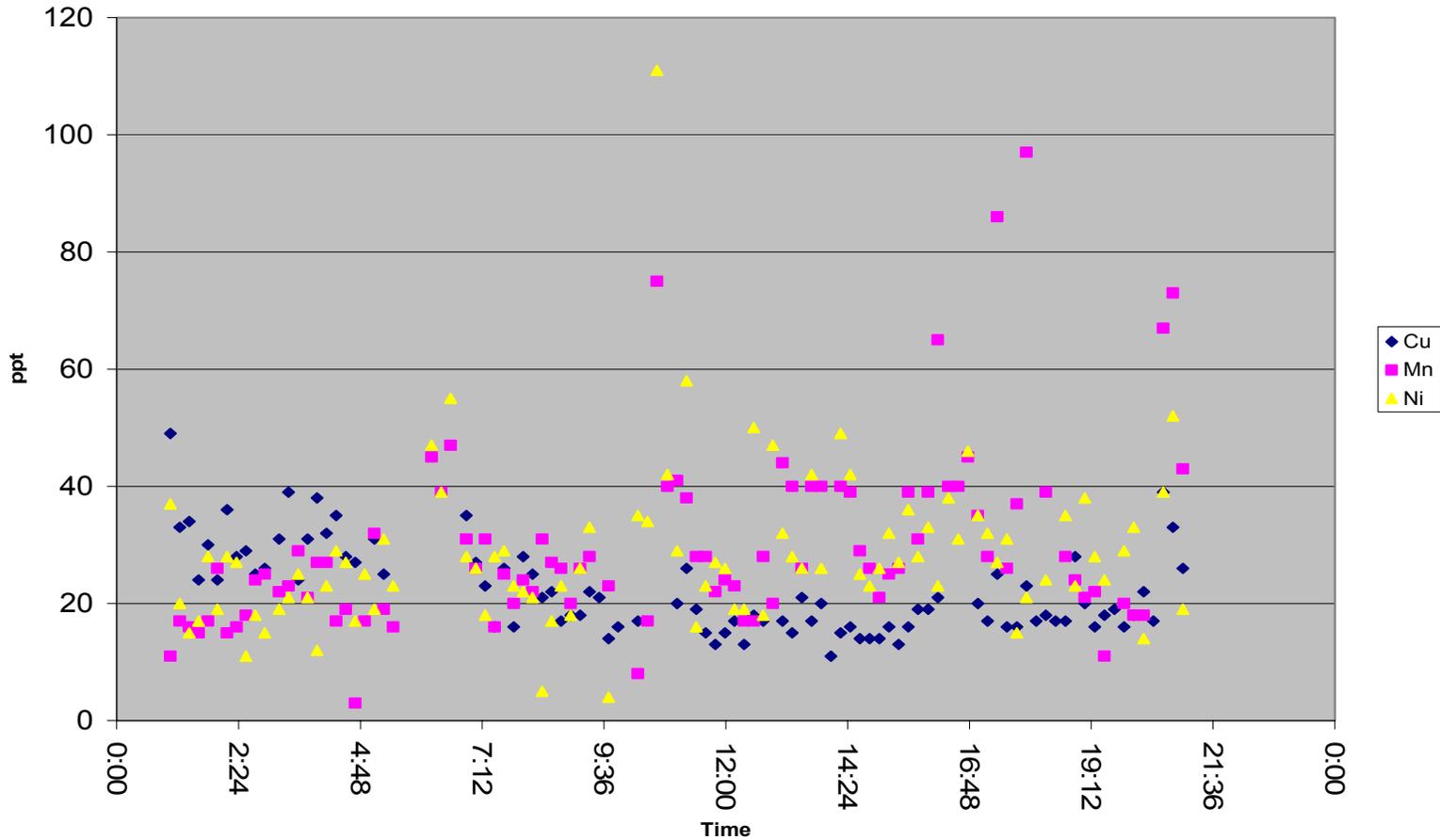
# IP-MS Performance for SC1 at 2 ppb 90% Confidence Limits, n = 3



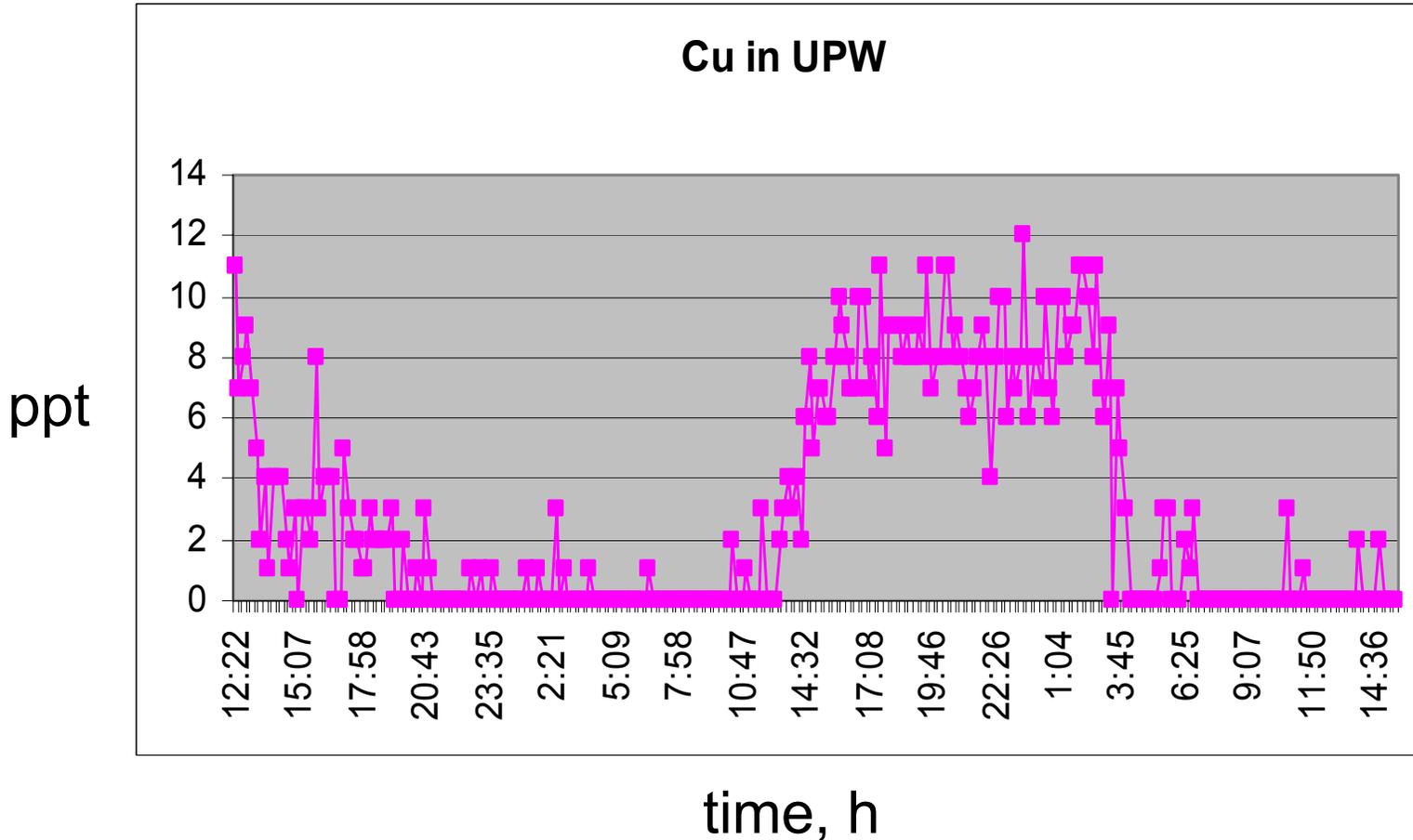
# IPMS, TCA analysis of 200, 500, 2000 ppt Solutions UPW Matrix, n=3, 90% Confidence Limit



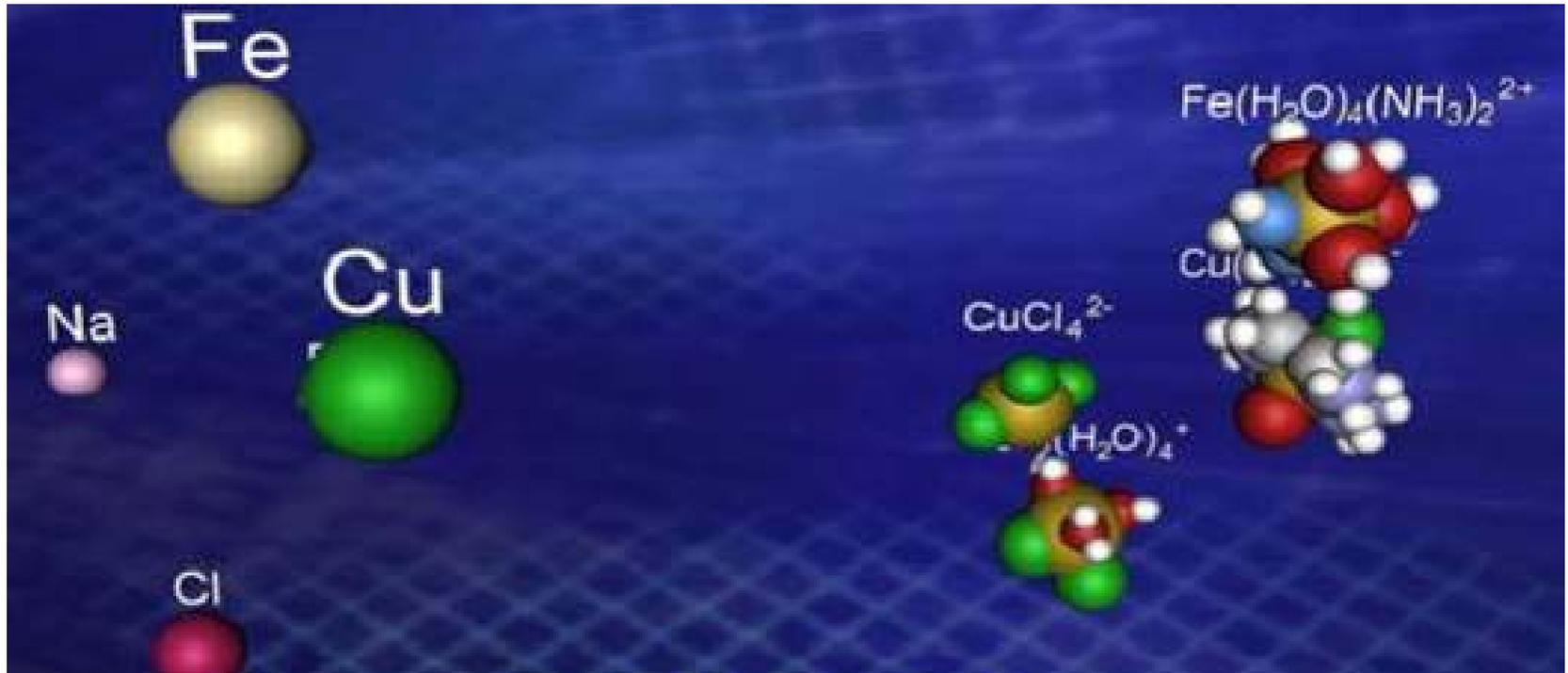
# On-line Monitoring: Cu, Ni and Mn Concentrations vs. Time in a SC1 Cleaning Bath



# Trend Data for Cu in UPW



# Elements & Species



Elements

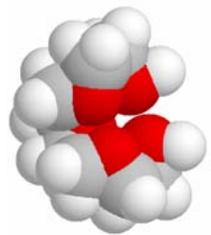
Species

# Ionization: Cations, Anions, Species

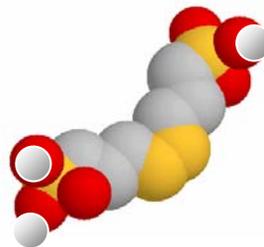
The instrument can detect *cations*, *anions*, and *molecular species* by reversing the polarity of appropriate voltages in the mass-spectrometer and using soft or hard ionization.

➤ Positive Mode

➤ Negative Mode



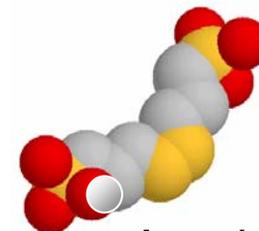
Suppressor



Accelerator  $[SPS \cdot 3H]^+$



Sulfate



Accelerator  $[SPS \cdot H]^-$

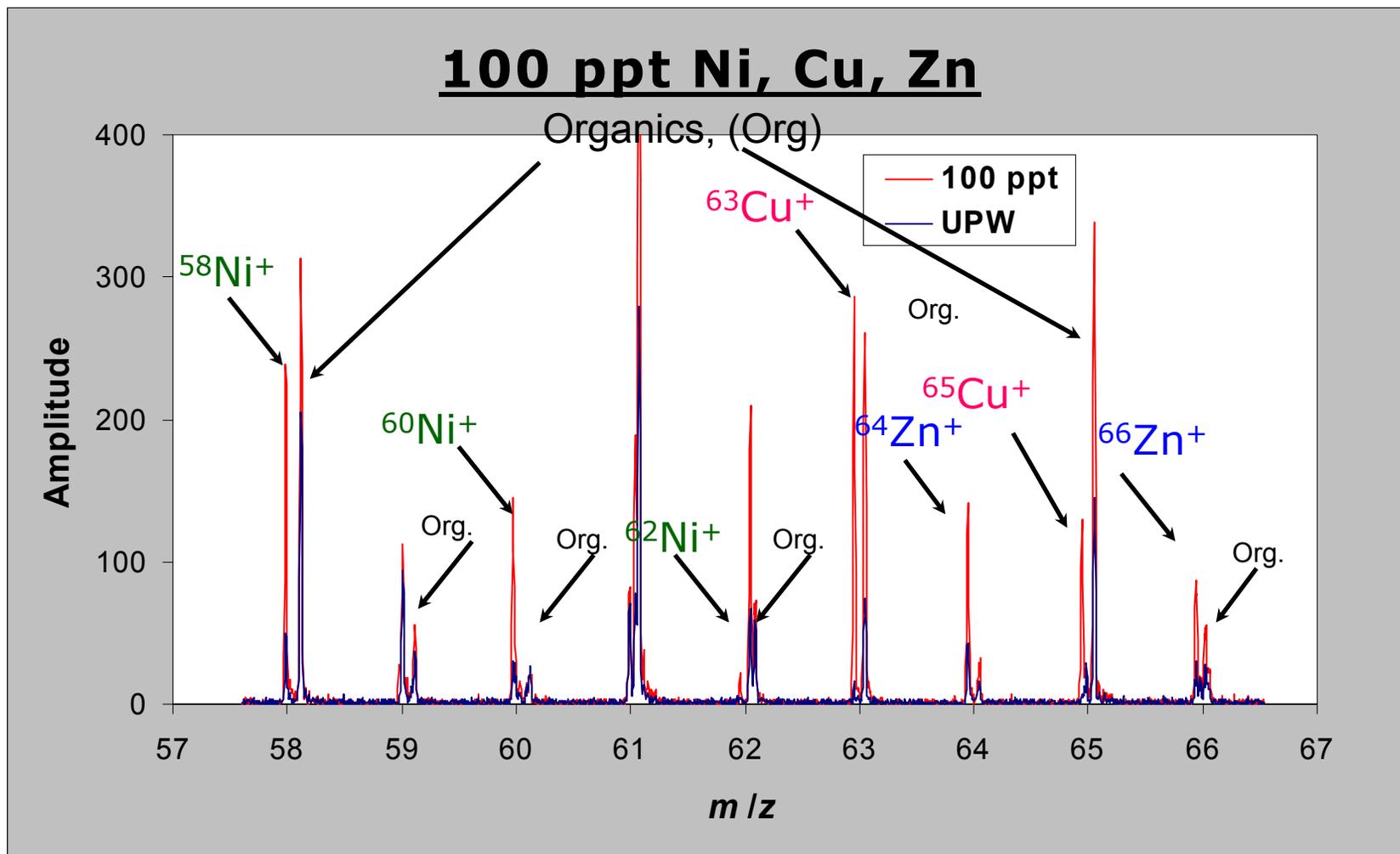


# Chemical Species Measurement Reveal Fundamental Molecular Structure and Chemical Mechanisms

- Contamination control depends directly on **molecular species** and **mechanisms** causing the contamination or controlling the process
- Species reveal the chemical processes involved and contain information relating the
  - Origin
  - Effect
  - Prevention
  - Remediation
  - System

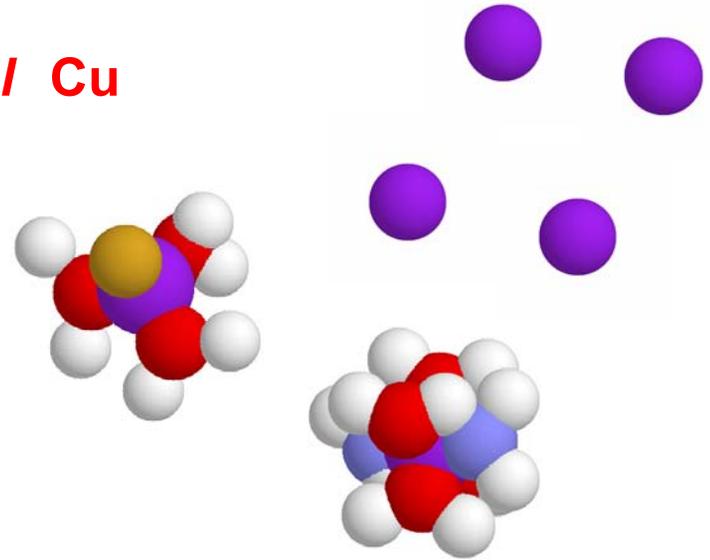


# Ni, Cu, Zn In **Soft Ionization Mode** Include Organic Molecular Ions



# Speciation Example: Cu Species Identification in Semiconductor Solutions

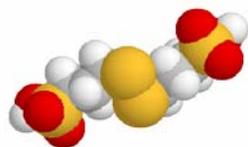
- Elemental quantitative mode: *total Cu*
- Speciation mode – Process info
- Example
  - UPW bath:  $[\text{Cu}(\text{H}_2\text{O})_4]^{2+}$
  - HF bath:  $[\text{CuF}(\text{H}_2\text{O})_3]^+$
  - SC-1 bath:  $[\text{Cu}(\text{NH}_3)_2(\text{H}_2\text{O})_4]^{2+}$
  - SC-2 bath:  $[\text{Cu}(\text{OOH})\text{Cl}_2]^-$
- Identifying Cu species in the monitored process, reveals sources, pathways and mechanisms of the Cu contamination.



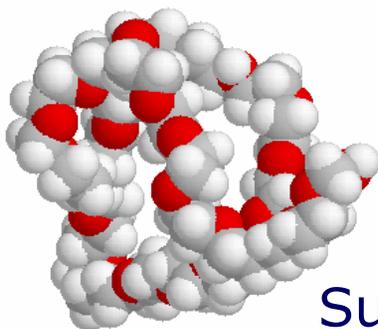
# Ionization: Elemental vs. Speciation

## ➤ Speciation Mode of Analysis for Organics & Inorganic Complexes

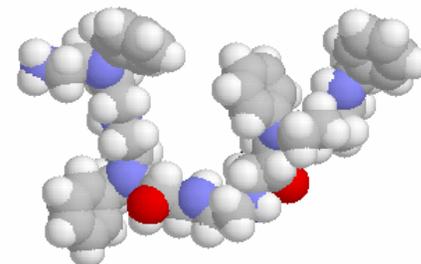
- 'Soft' mode – the molecular ions are imparted with a low kinetic energy in the gas-phase.
- Gentle collisions minimize fragmentation and ligand loss.
- The integrity of the solution-species is preserved:



Accelerator



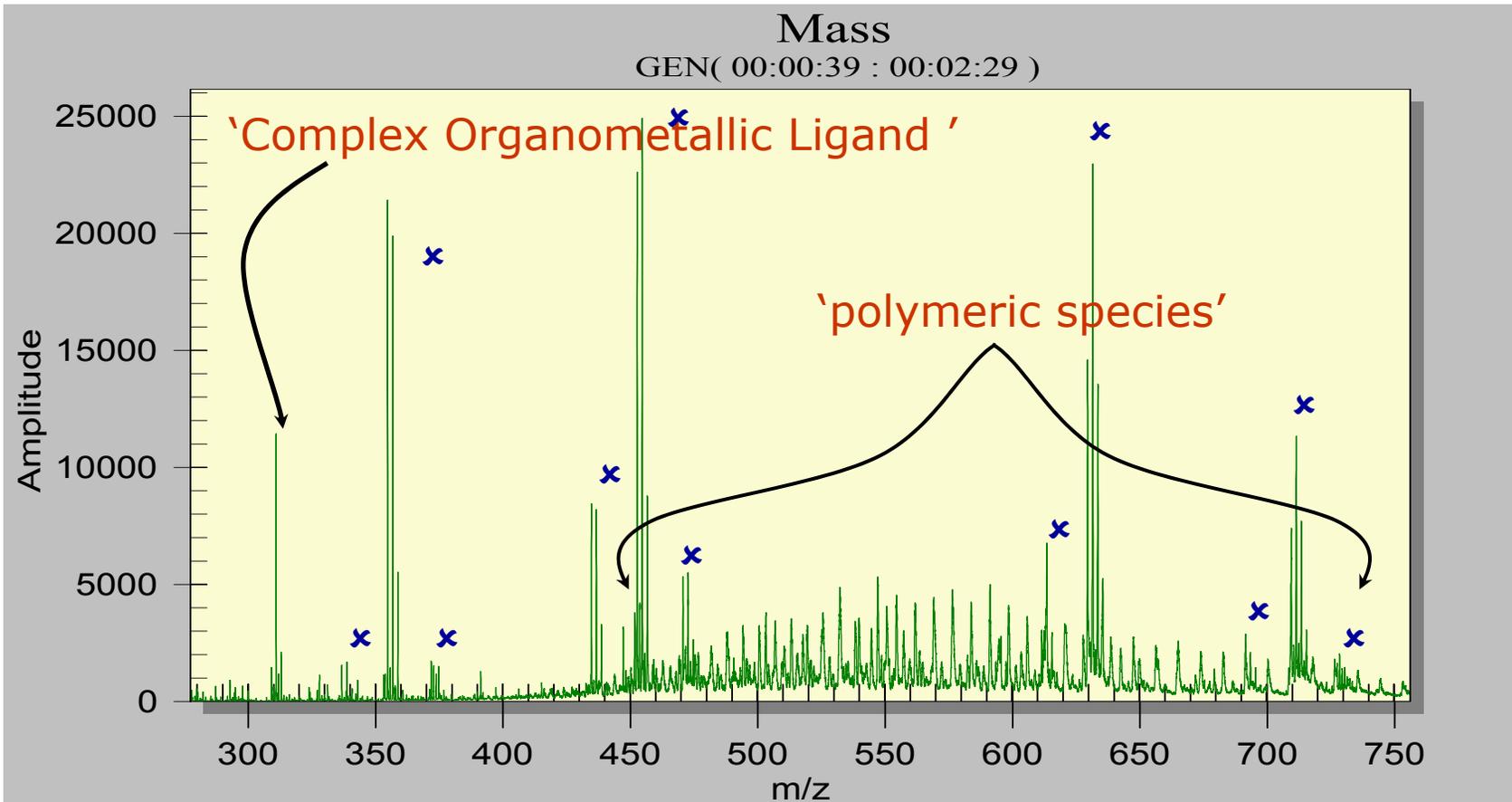
Suppressor



Leveler

- Diagnostic tool for revealing the chemistry of the process
- Provides rich information for yield enhancement tools

# Mass Spectrum of Complex Sample: Cu plating bath solution

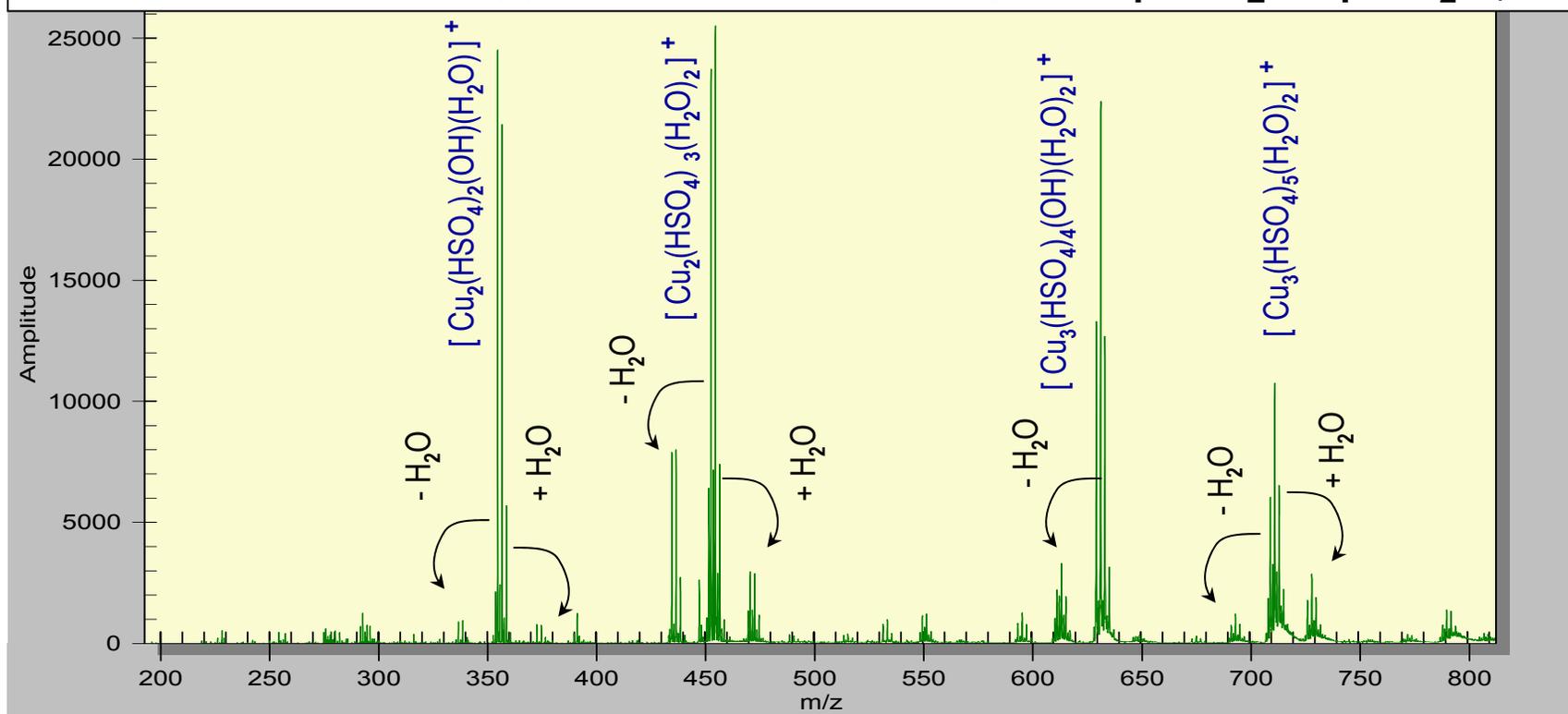


x = Organometallic cluster ions

# Cu 'reagent': Speciation Mode

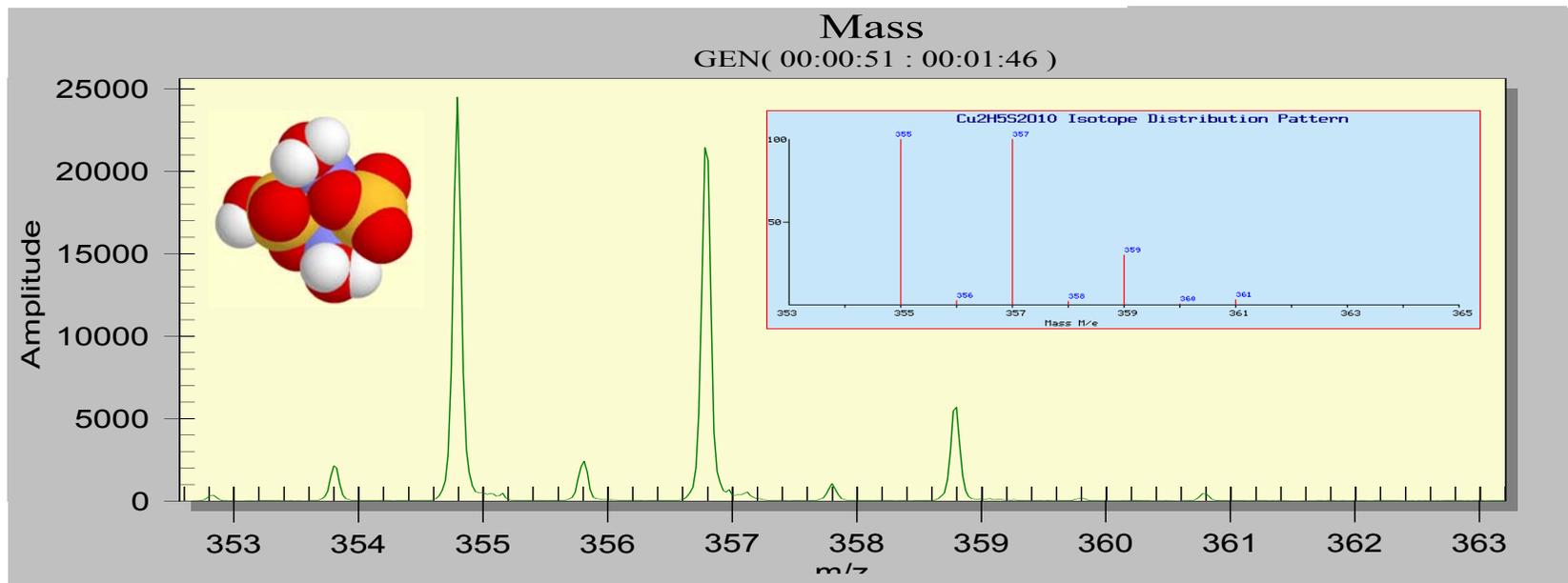
## Qualitative and Quantitative

Ionic clusters of copper, sulfate & water ( $\text{CuSO}_4 + \text{H}_2\text{SO}_4 + \text{H}_2\text{O}$ )



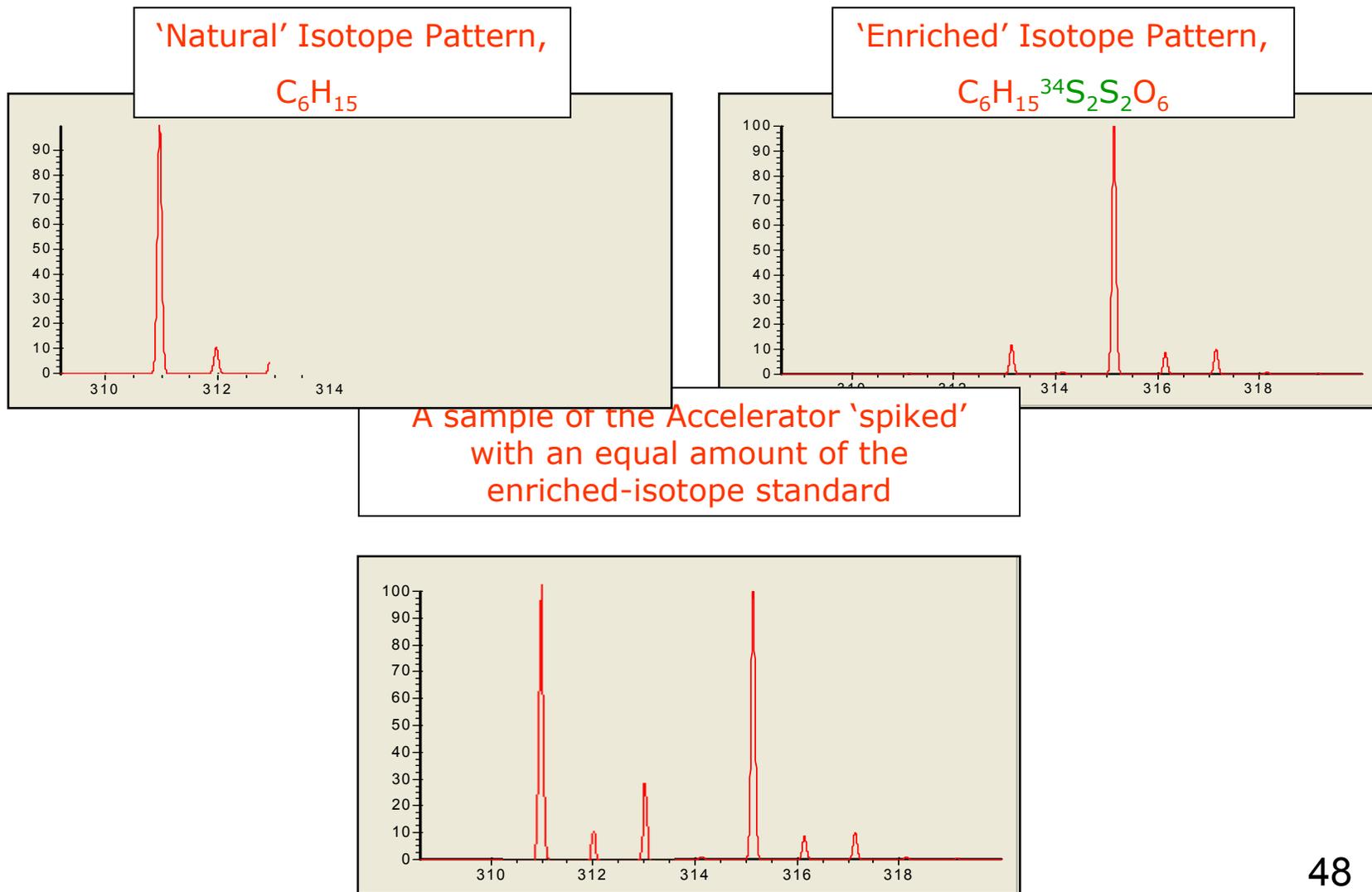
- 'Soft Ionization' in Speciation Mode encourages detection of intact clusters of copper ions with sulfate and water, by minimizing their fragmentation in the mass-spectrometer.
- These parameters have selected for copper sulfate clusters containing 2 & 3 copper ions (at below and above  $m/z \sim 500$  respectively) with a charge,  $z$ , of +1.
- The chemical formulae of the species are reliable, although the structural assignments are speculative.

# Qualitative Identification and Quantitative Analysis of one Copper species:



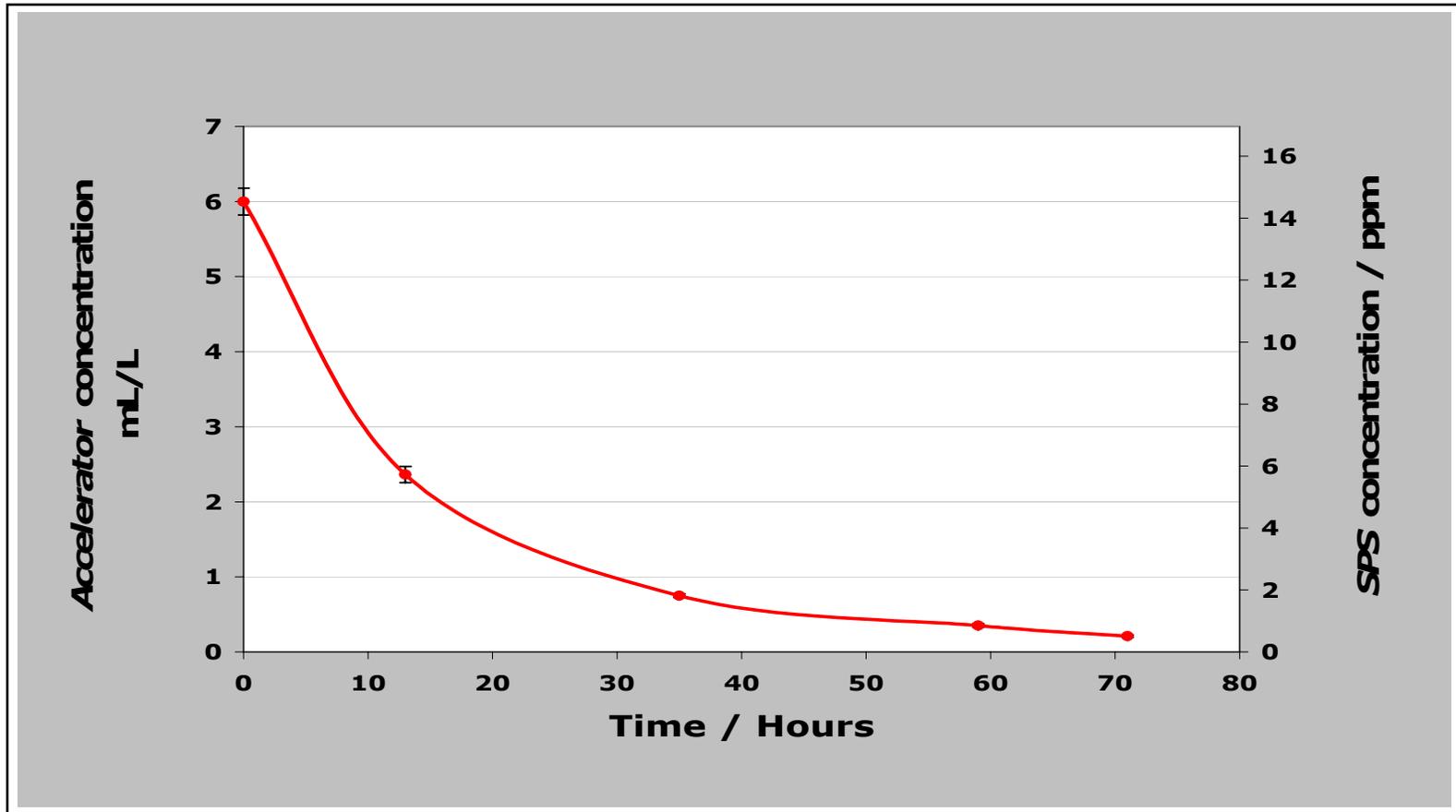
- The **isotope peak pattern** is consistent with the chemical formulation; the most abundant isotopes of both copper and sulfur differ by 2 a.m.u. ( $^{63}\text{Cu}$  &  $^{65}\text{Cu}$ , and  $^{32}\text{S}$  &  $^{34}\text{S}$ ).
- There is an excellent match between the **measured** & **calculated** masses for the major isotope peak (**354.7922** and **354.7916** a.m.u. respectively).
- The set of peaks of lower intensity offset by 1 a.m.u. is consistent with the formulation  $[\text{Cu}_2\text{S}_2\text{H}_4\text{O}_{10}]^+$ , perhaps formed by the collisional loss of a hydrogen atom in the mass-spectrometer.

# Quantitation of the Accelerator Bis(3-sulfopropyl) disulfide (SPS) by Isotopically-Enriched Spike



# Trend Data

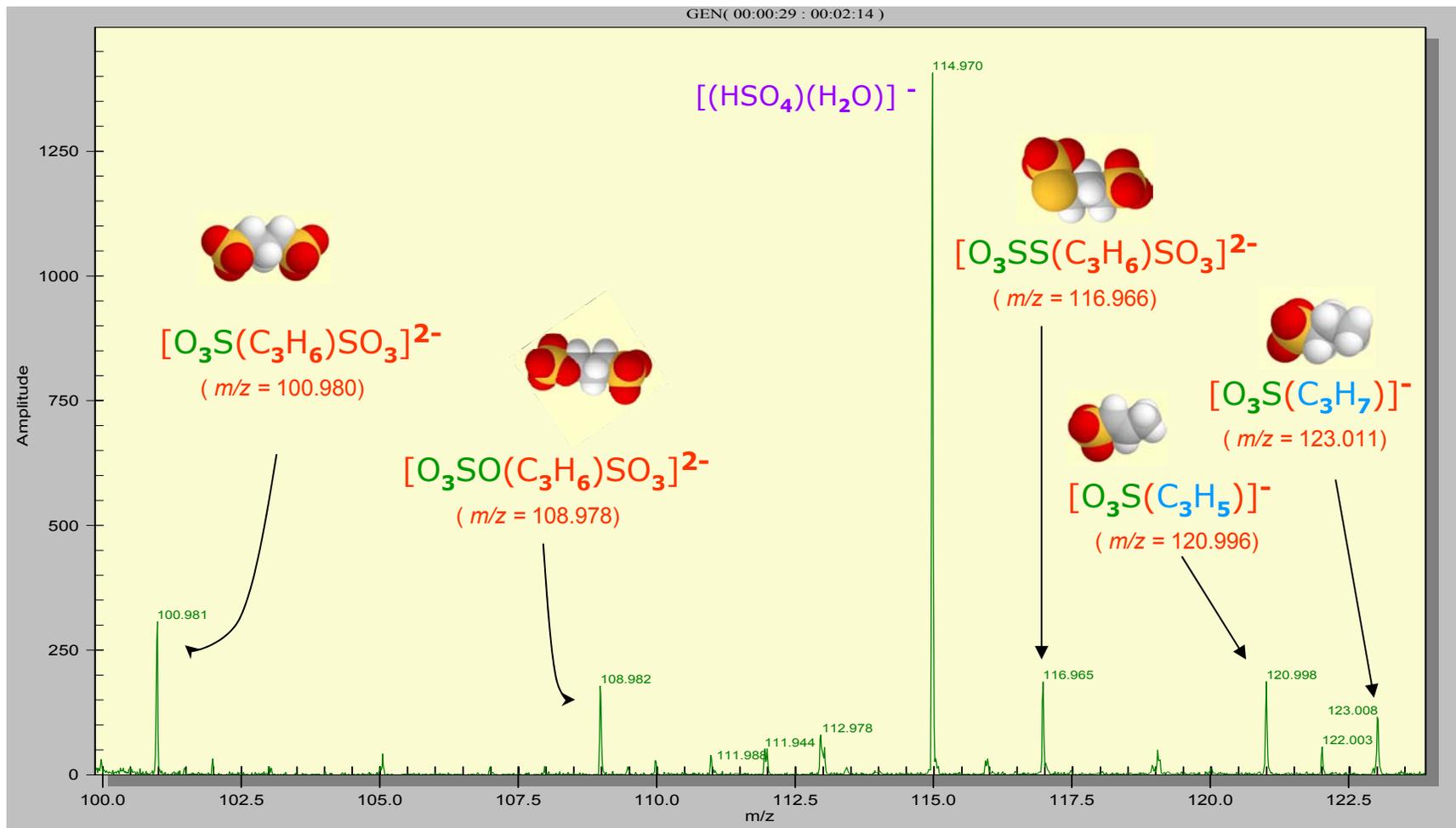
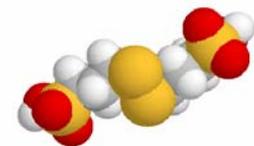
## Rate of *Accelerator* (SPS) Change in a Cu-ECD Test Facility



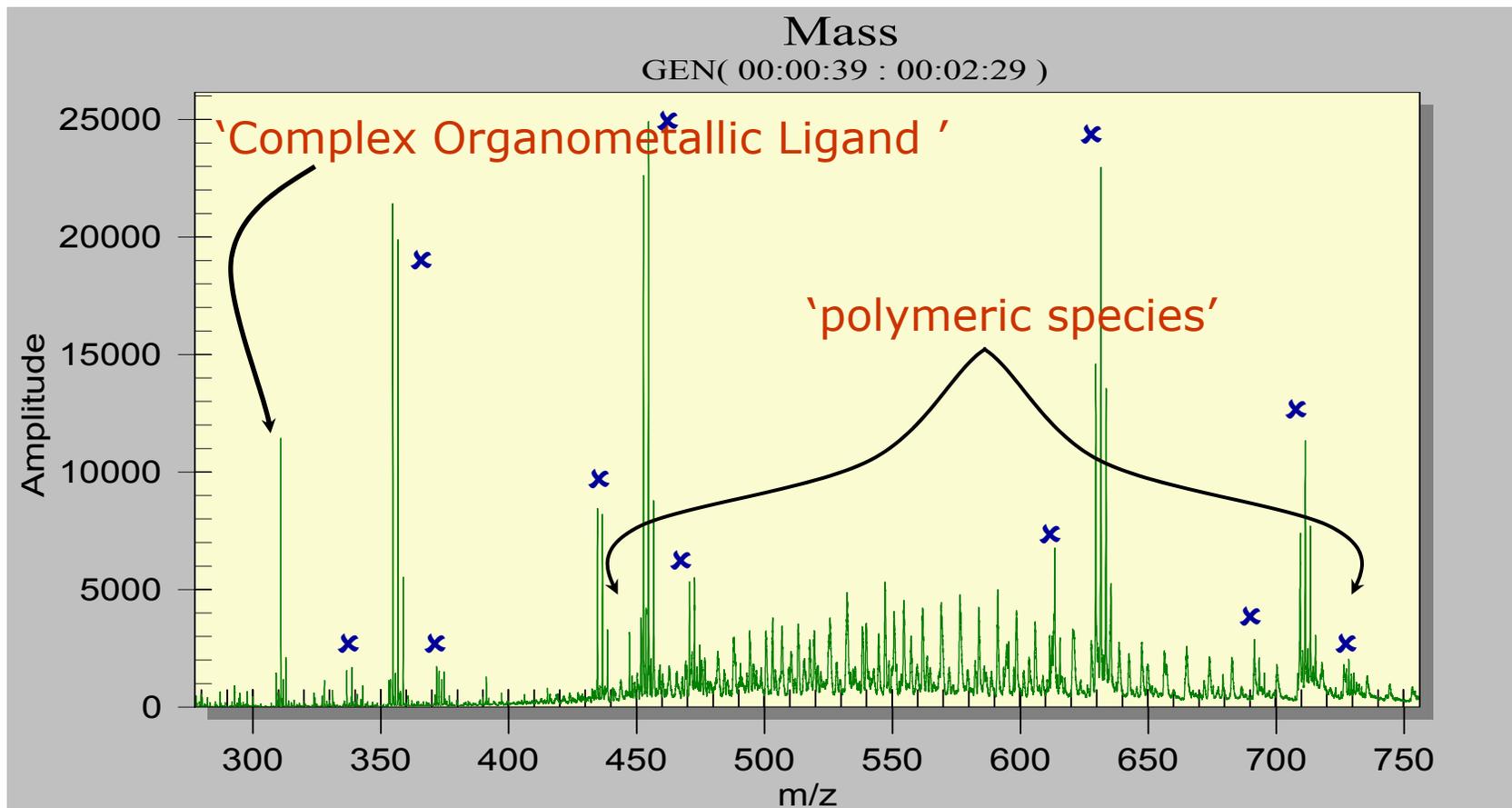
• 10 amps • 1.635 V • 2 Sabre anodes



# Decomposition of the Accelerator *SPS* in an Aged Sample



# Mass Spectrum of Complex Sample



x = Organometallic cluster ions

## What is the Challenge?

**“The most critical challenge is to find ways to determine the effects of trace impurities on device performance and yield.”**

Reference: “Examining upcoming yield enhancement challenges in the 2001 roadmap”

*-Micro*, February 2002

Authors: Christopher Long, IBM;  
Milton Godwin, Applied Materials;  
Manuela Huber, Sematech/Infineon;  
Richard Jarvis, Sematech/AMD  
Fred Lakhani, Sematech

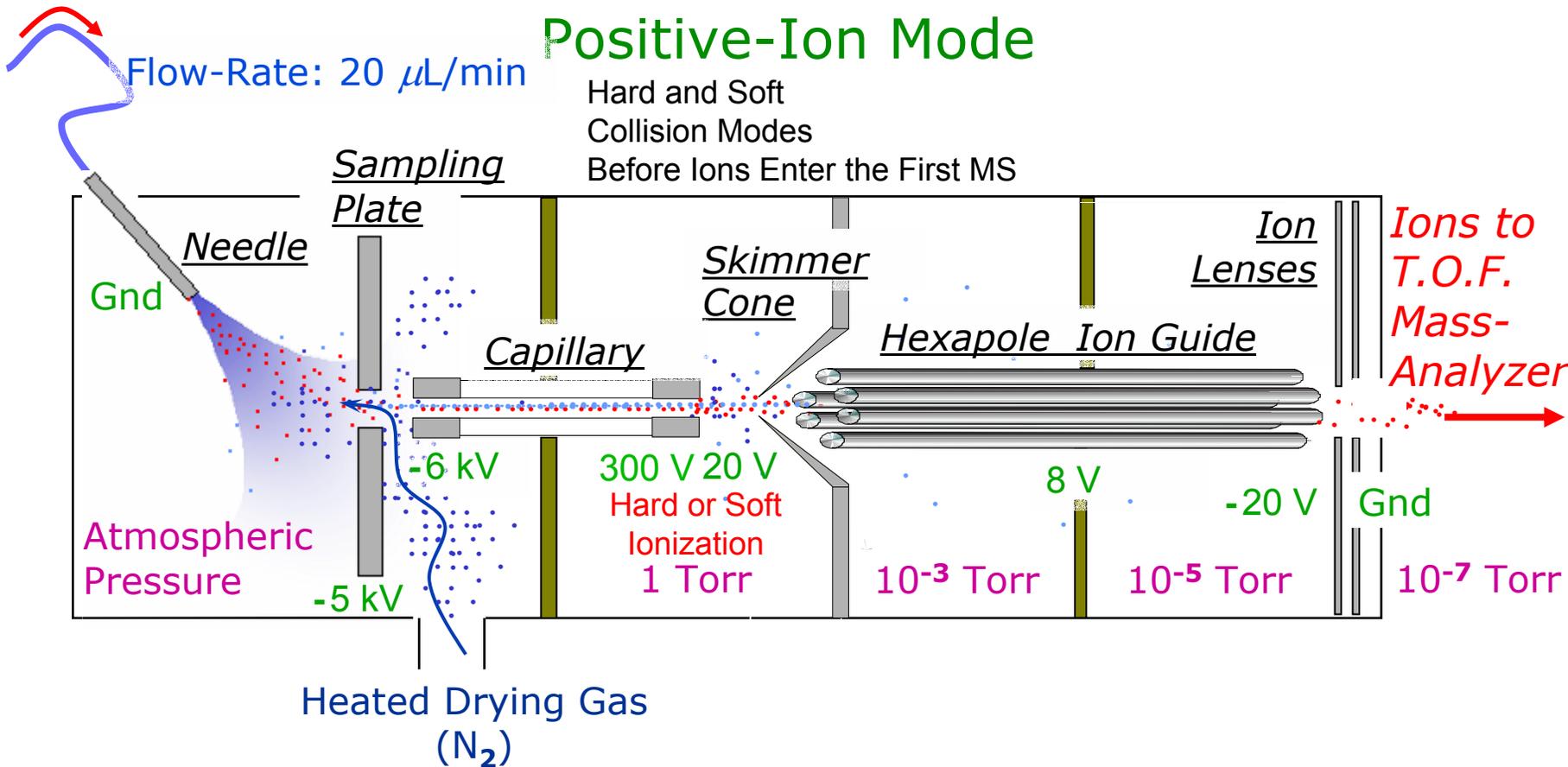


# The Integrated System



# Electrospray Ionization Interface

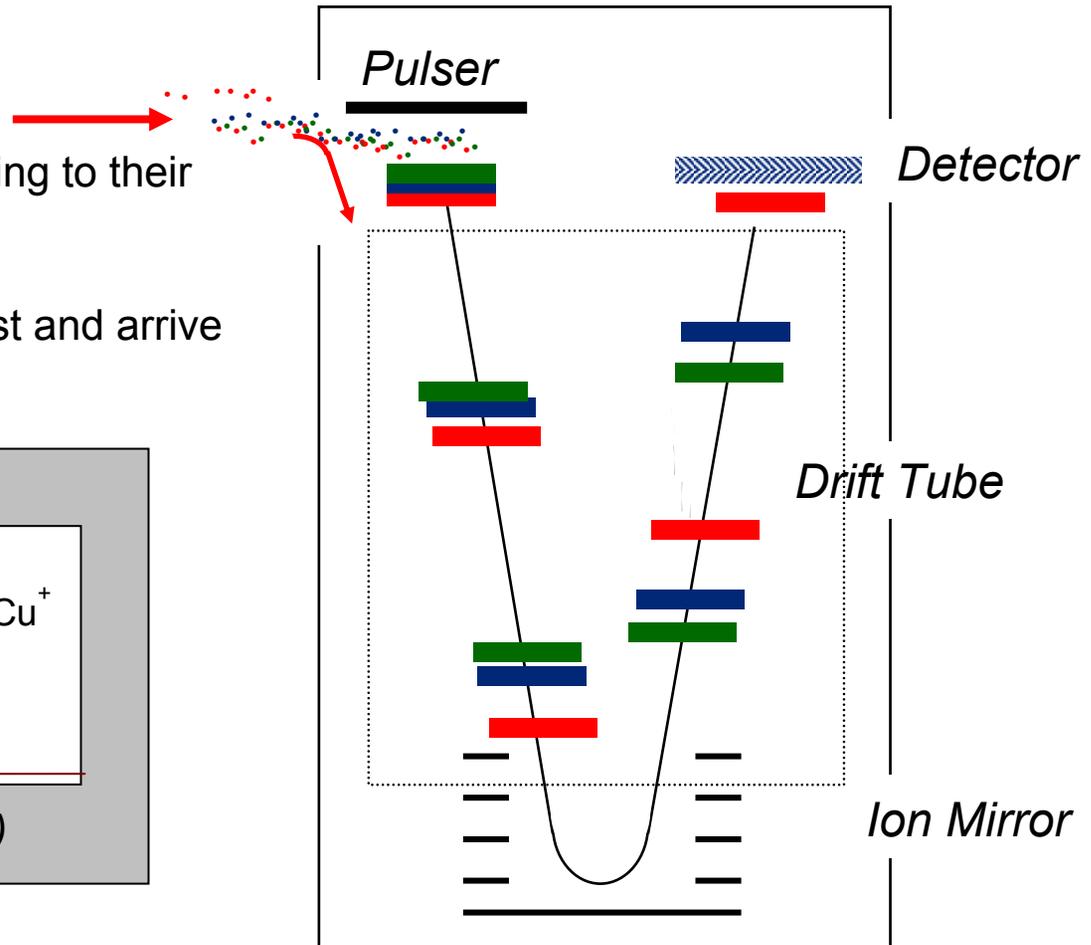
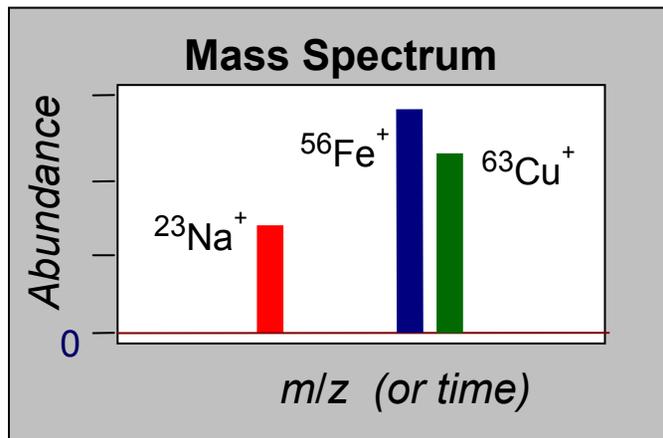
from Sample Preparation Unit



# Time-of-Flight Mass Analyzer

*Ions from Electrospray*

- Ion packets separate according to their mass-to-charge ratio ( $m/z$ ).
- The lightest ions travel fastest and arrive at the detector earliest.



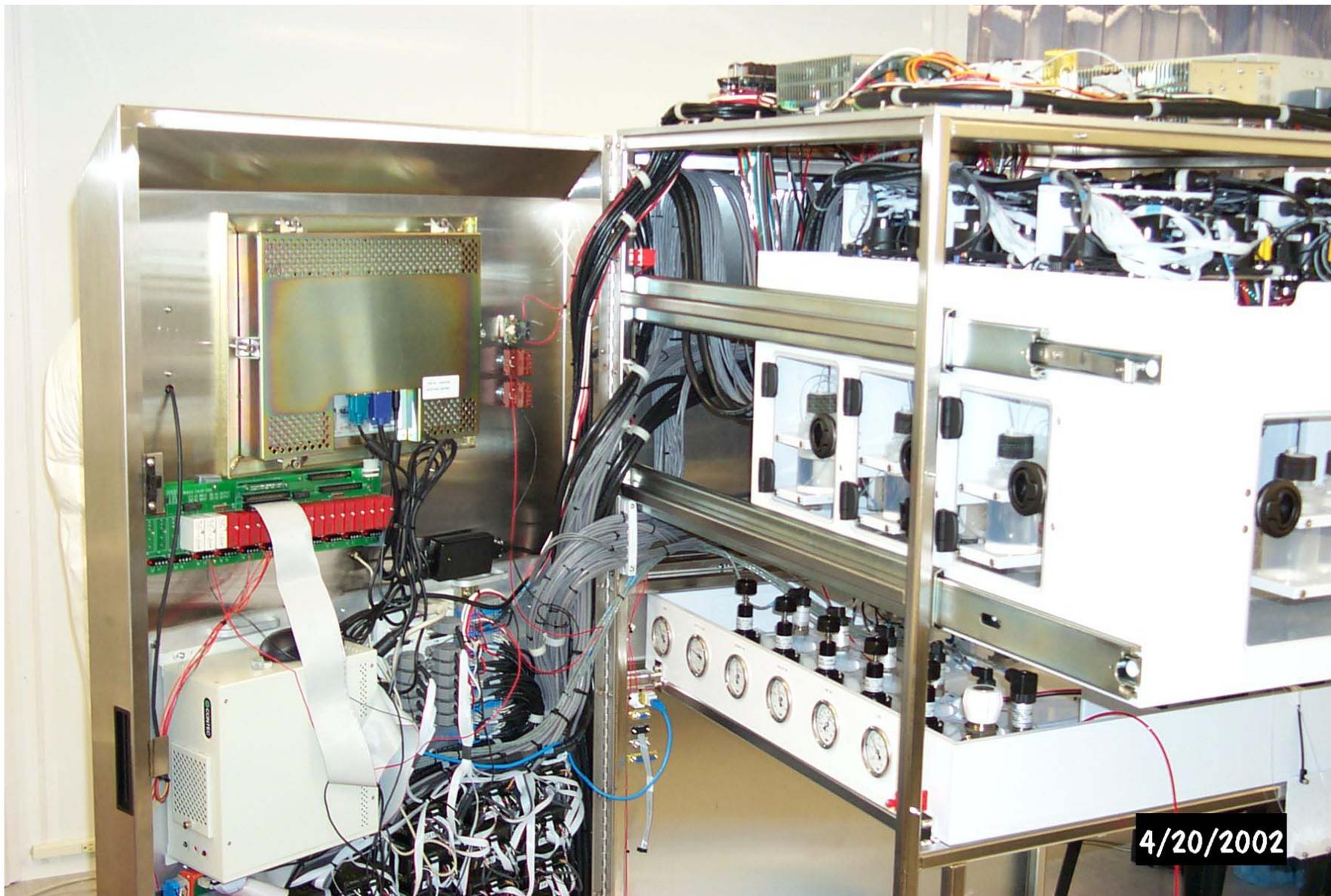
# IPMS (ES-TOF) Vs. ICP-MS

| Item                                   | LMS-300 TCA   | ICP-MS  |
|--|---|---|
| Target Application                     | Real-time, In-line Trace contamination monitoring of standard aqueous semiconductor solutions               | Lab applications  |
| Sample Matrix                          | UPW, SC1, SC2, IPA, DHF, HNO <sub>3</sub> , H <sub>2</sub> O <sub>2</sub>                                   | UPW, SC1, SC2, IPA, DHF, HNO <sub>3</sub> , H <sub>2</sub> O <sub>2</sub> |
| Monitors                               | 16 (22) Trace Contaminants<br>Cations (+) and anions (-)<br>Elements and species<br>Inorganics and Organics | Depends Upon Available Standards<br>Cations (+)<br>Elements<br>Inorganics |
| Sample Preparation                     | Automatic   | Manual  |
| Limit of Detection                     | < 20 ppt  | < 20 ppt  |
| Calibration and Quantitation Standards | Enriched isotope elemental standard (250 ppb)<br>NIST natural isotope standard (10 ppm)                     | Calibration Curves  |
| Sample Volume                          | 2 mL  | 5 mL  |
| Technology                             | In-Process Mass Spectrometry (IPMS)   | ICP-MS  |
| Throughput                             | 8 sample/hr (7.5 min/sample)  | 24 to 72 hours  |
| Accuracy                               | ± 25% at the quantitation limit   | ± 25% at the quantitation limit   |



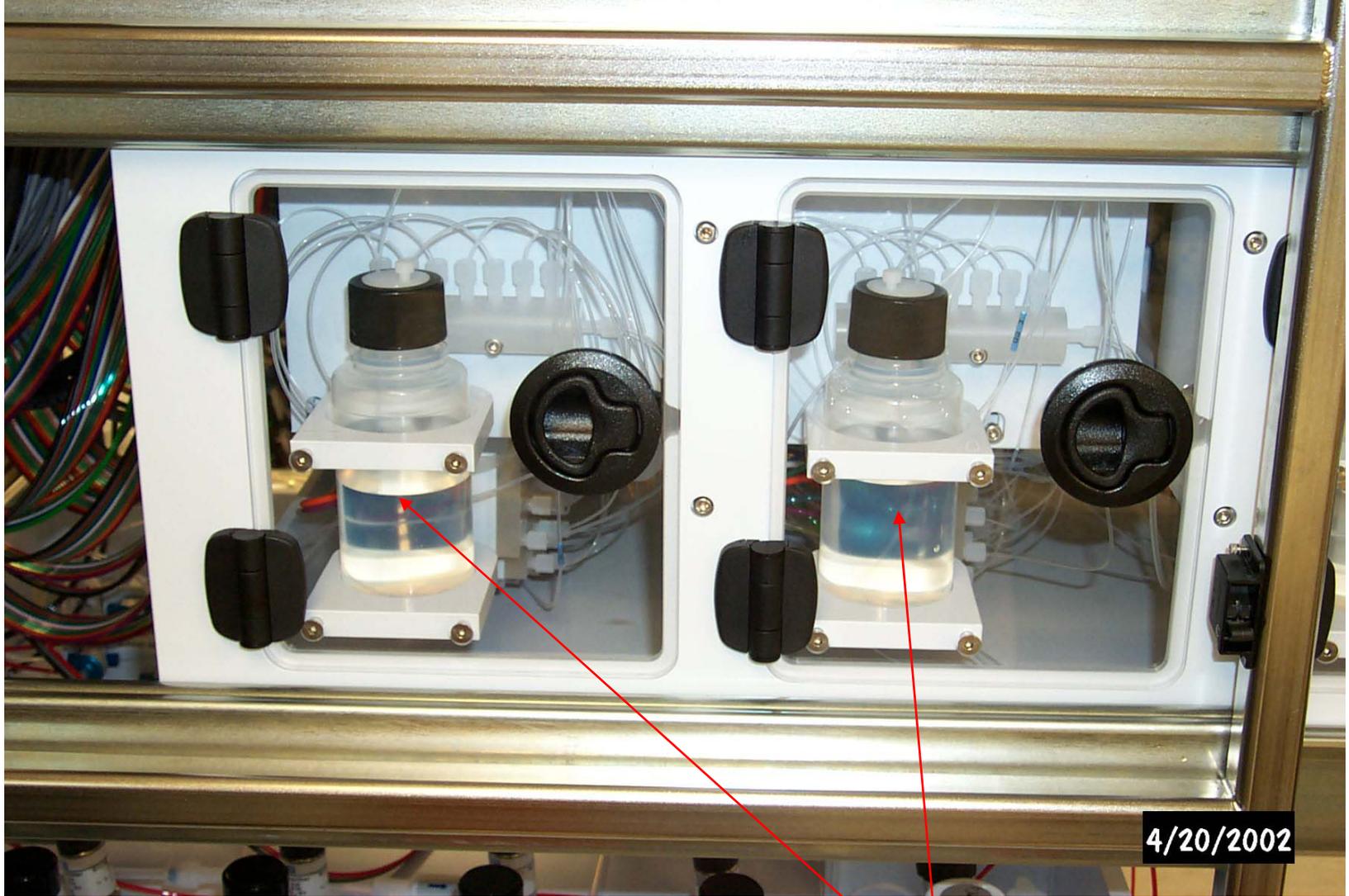
# Trace Contamination Analyzer (TCA) Semiconductor Solution Metrology





4/20/2002





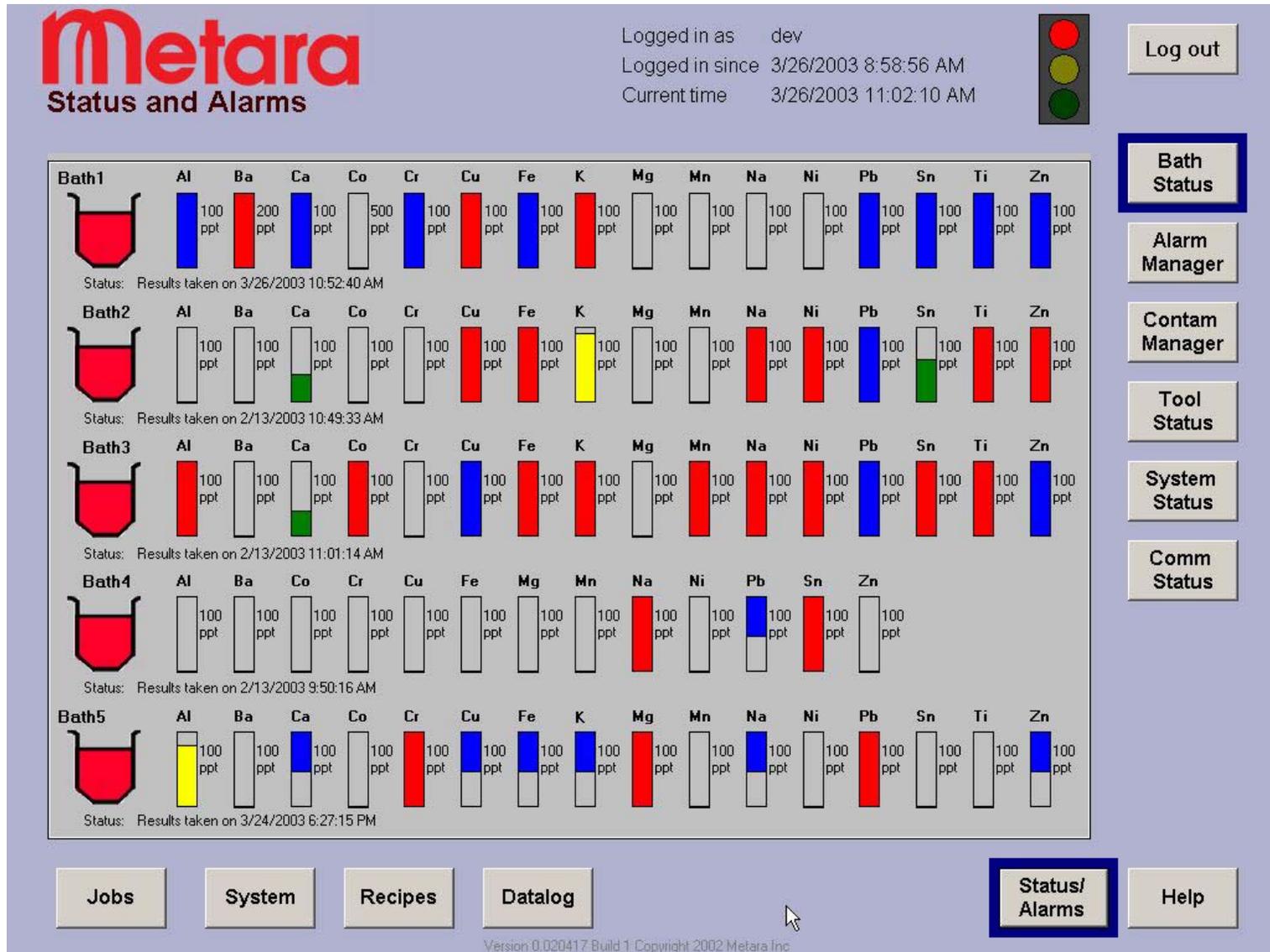
4/20/2002

“Enriched” Isotope Solutions Standards”

# One semiconductor instrument being tested and one being assembled in clean room



# Decision Information in Graphical User Interface (Beta) (Element selection individualized for a specific Fab)



# Acknowledgements

## Current and Former Research Group Member

### Duquesne University

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Ds. Helen Boylan

Ms. Sejal Iyer

Mr. Mizanur Rahman

Dr. Dan Taylor

Ms. Yusheng Lu

Dr. Dirk Link

Dr. Dengwei Huo

Dr. Stuart Chalk

Dr. Peter Walter

Dr. Robert Richter

Many others

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Dr. Harmish Seni

Mr. Bob Ormond

Dr. Kenneth Bhang

Mr. Rudy Mui

Mr. Patrick Franklin

programmers, engeneers...

Many others



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**Allegheny Energy Supply Company**

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**Metara Inc.**

**Milestone Inc.**

**Agilent Technologies**

**Fisons**

**Note:**

**Patents exist and are pending on portions of this research  
Nationally and Internationally**





*Thank YOU*

*This is not your fathers kind of chemistry any more!*

NI SI Conference 3/27/2003

H. M. Skip Kingston



**Micromachines of the future will be on an unprecedented scale with atomic and molecular domains where constituent and contaminant analysis must be on these same scales**

