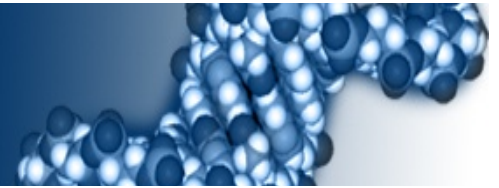


Parameterization of an *in-silico* DNA pipeline with & laboratory-specific experimental data allows for efficient & validation of the DNA analysis process &

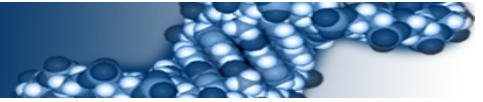
Boston University School of Medicine
Program in Biomedical Forensic Sciences
72 E. Concord Street, Boston, MA 02118



Harish Swaminathan &

Forensic Science Error Management &
International Forensics Symposium &
NIST, Gaithersburg, MD &
July 27, 2017 &

Quality of the electropherogram affects DNA mixture interpretation &

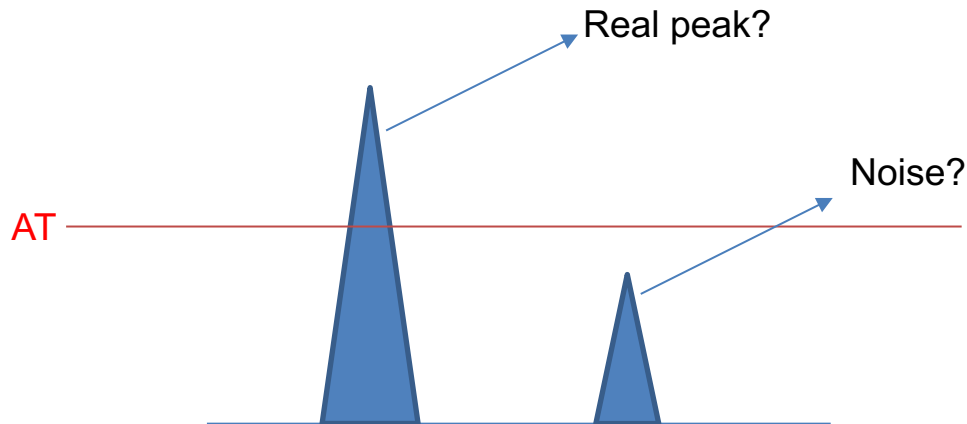


- Electropherogram (EPG): Allele signal + Background noise + Artifacts
- Interpretation can be challenging
- Mixtures with major and **minor**
- Low copy number samples typically exhibit signal loss
 - Sampling effects
 - **Detection effects**

$$\textit{Likelihood Ratio (LR)} = \frac{\Pr(\mathbf{E}|H_1)}{\Pr(\mathbf{E}|H_2)}$$

- By improving the information content of E , one can expect a more informative LR
 - For e.g. a large LR for a true contributor and a small LR for a non-contributor
- Focus of the talk is on development of a validation scheme to **improve signal-to-noise resolution** and to **minimize detection error rates**

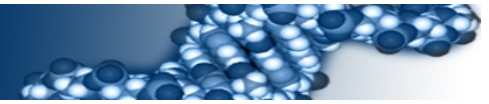
Optimal AT is necessary to minimize detection errors &



- Analytical Threshold (AT): the minimum height requirement at and above which detected peaks can be reliably distinguished from background noise*
- Errors can occur while applying an AT
- False Positive or Type I error: Noise peaks are mislabeled as real peaks
- False Negative or Type II error: Real peaks are not labelled (dropout)
- Ideally, the chosen AT minimizes both types of errors
- **AT impacts downstream interpretation process, including the match statistic**

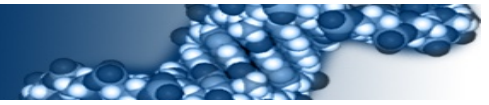
*SWGAM Interpretation Guidelines for Autosomal STR Typing by Forensic DNA Testing Laboratories – APPROVED 01/12/2017 &

Combined simulation + experimental approach &



- **Time** and **cost** are limiting factors in validation
 - For e.g. AT should be determined by large-scale in-house validation studies using negatives, dilution series, etc.
- *In-silico* execution of the forensic DNA analysis process allows for **fast, easy, inexpensive** generation of **representative** large-scale EPG data
- **Quickly evaluate optimal laboratory conditions under various scenarios &**
- Improve detection rates:
 - Determine optimal **AT** to minimize Type I and Type II error rates
- Improve signal-to-noise resolution:
 - Explore optimal values for parameters such as **number of PCR cycles, time of injection**, etc.

RESOLVEit: Resolve Evidentiary Signal by Optimizing Laboratory's Validation &



RESOLVEit

Calibration file:

Frequency file:

Stutter file:

Output files directory:

Number of simulations: DNA conc (ng/uL):

Injection time (for simulation): Prob (Observing noise):

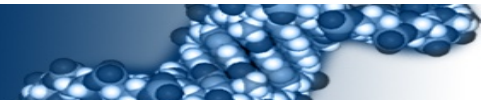
PCR cycles (for calibration): Volume (Amp):

PCR cycles (for simulation): Volume (CE):

PCR efficiency (for calibration): Final IT:

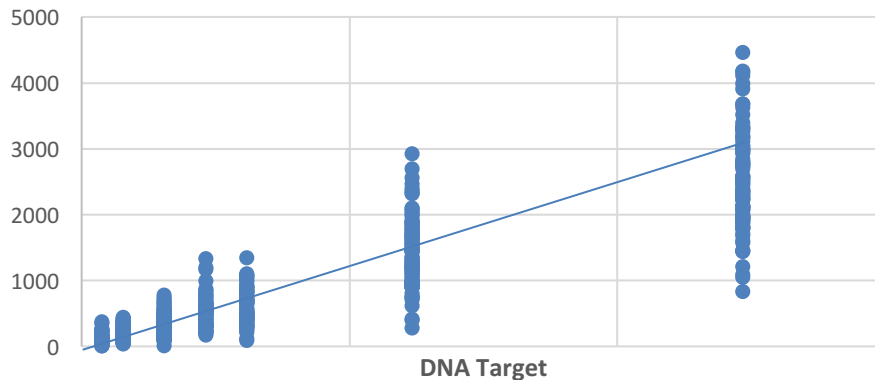
PCR efficiency (for simulation):

Step I: Parametrization

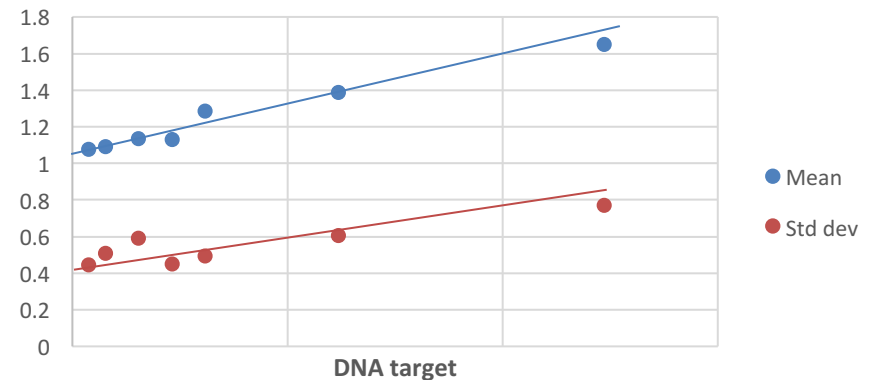


- **Laboratory-specific data:** Large number of single source samples of known genotypes at different targets and injection times
- Calculate **CE sensitivity α**
 - Describes increase in signal wrt target concentration of DNA
- Calculate **noise parameters - mean μ and std dev σ** - at each target concentration, assuming a lognormal distribution*

D8S1179 - 10s – heights of peaks at allele and stutter positions

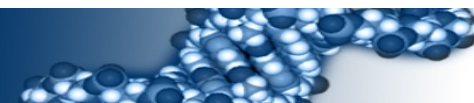


D8S1179 - 10s – heights of peaks at noise positions



*Probabilistic characterisation of baseline noise in STR profiles, Monich et al, Forensic Science International Genetics 19 (2015) 107-122. &

Step II: Simulation of signal and noise* &



- Signal is simulated starting from a **single target copy number**
- Noise is simulated from a **specified target mass**

Module 1

Genotype sampling

- 2 alleles are sampled at each locus
- $N_0 = 1$ for each allele

Module 2

DNA amplification (PCR)

- At each of the N_{PCR} cycles:
 - **Amp efficiency:**
Each True amplicon:
 - a) Gets copied or
 - b) Doesn't get copied.
 - **Stutter slippage probability:**
Each copied amplicon:
 - a) Causes stutter or
 - b) Doesn't cause stutter.

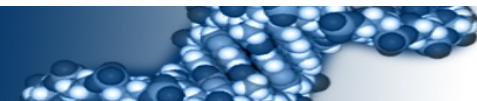
Module 3

Capillary electrophoresis and fluorescence calculation

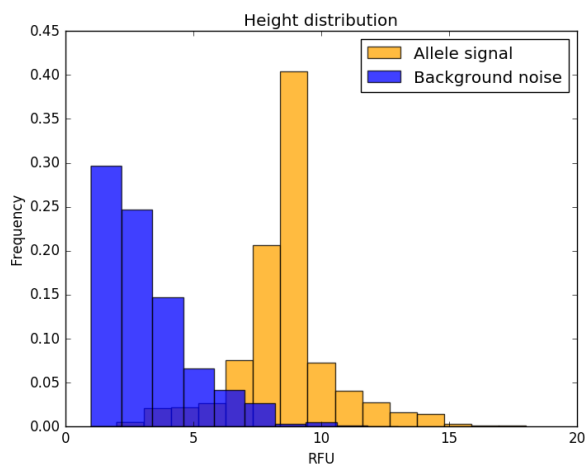
- $H_{allele} \sim B(N_{PCR}, V_{CE}/V_{AMP}) * \alpha$
- $H_{noise} \sim \text{Lognormal}(\mu, \sigma)$

*Exploring STR signal in the single- and multicopy number regimes: Deductions from an in silico model of the entire DNA laboratory process. KR Duffy et al. Electrophoresis 38 (6), 855-868, 2017.

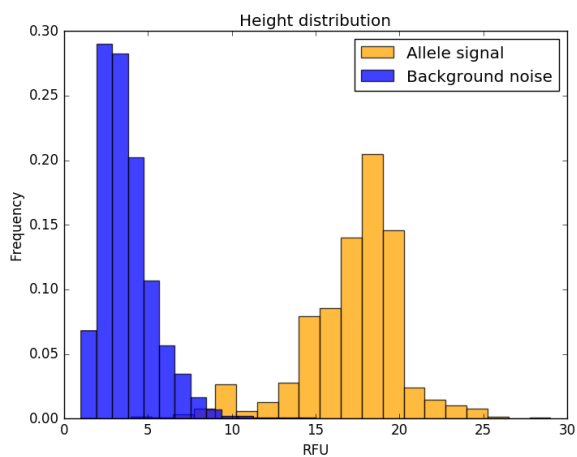
Results: D8S1179 – 1000 simulations &



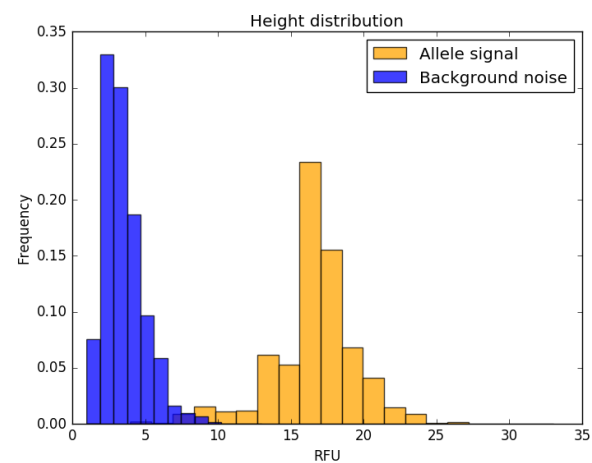
Sim 1 – 5s, 28 cycles &
Allele signal: 1 copy, Background noise: 0.008ng &



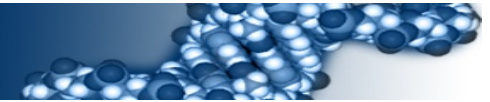
Sim 2 – 10s, 28 cycles
Allele signal: 1 copy, Background noise: 0.008ng



Sim 3 – 5s, 29 cycles
Allele signal: 1 copy, Background noise: 0.008ng



Results: D8S1179 – 1000 simulations &

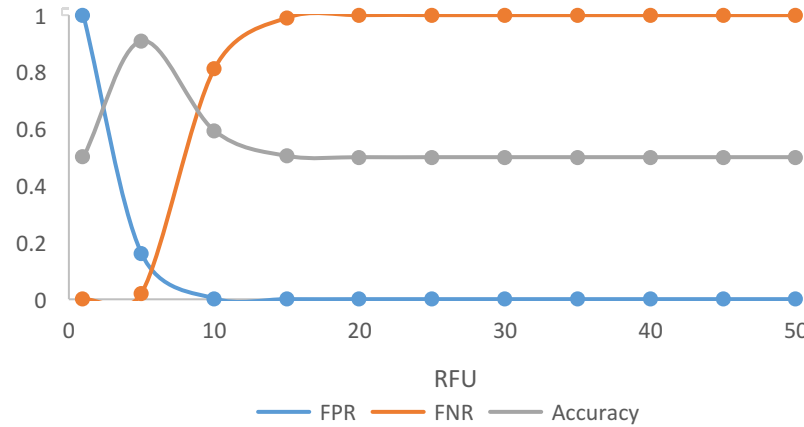


$$FPR = \frac{FP}{FP + TN}$$

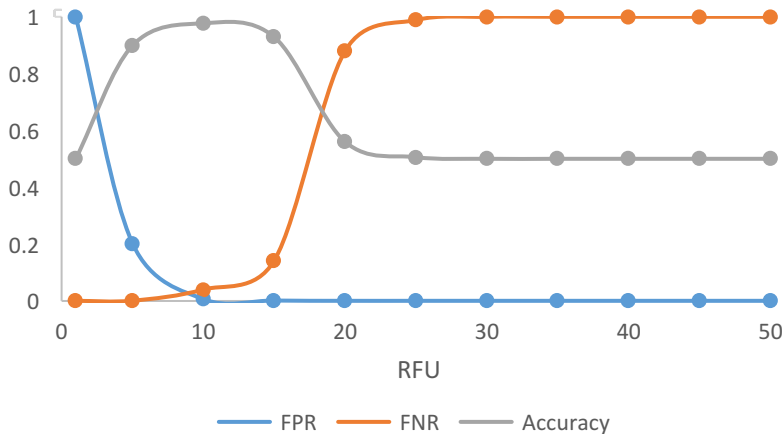
$$FNR = \frac{FN}{FN + TP}$$

$$ACC = \frac{TP + TN}{TP + FP + FN + TN}$$

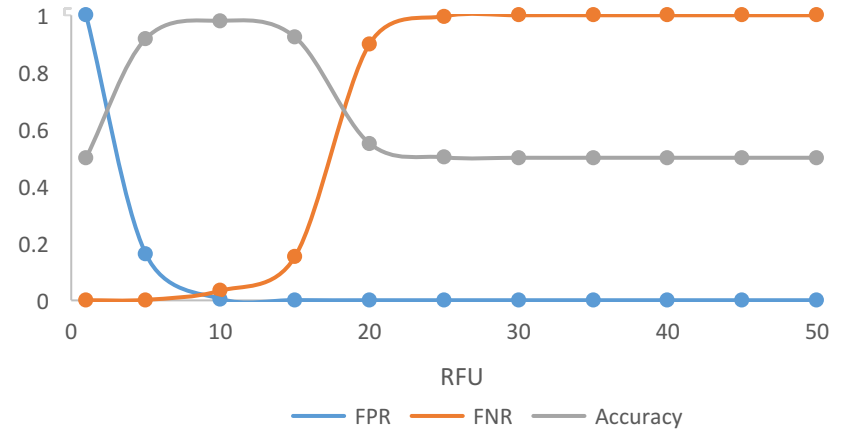
**Sim 1 – 5s, 28 cycles &
Allele signal: 1 copy, Background noise: 0.008ng &
Error rates of AT**



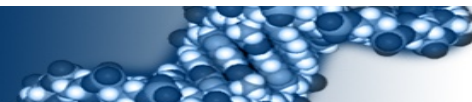
**Sim 2 – 10s, 28 cycles
Allele signal: 1 copy, Background noise: 0.008ng
Error rates of AT**



**Sim 3 – 5s, 29 cycles
Allele signal: 1 copy, Background noise: 0.008ng
Error rates of AT**

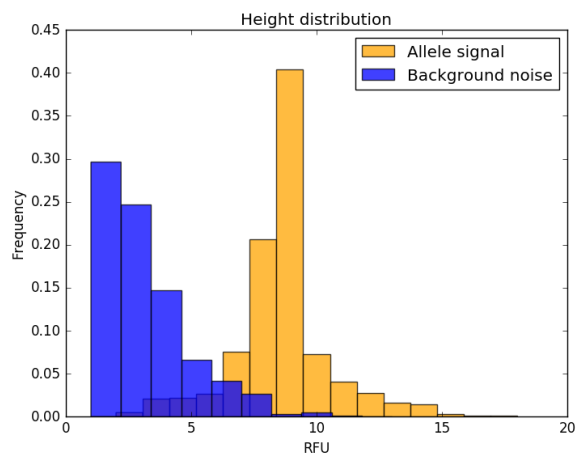


Results: D8S1179 – 1000 simulations &



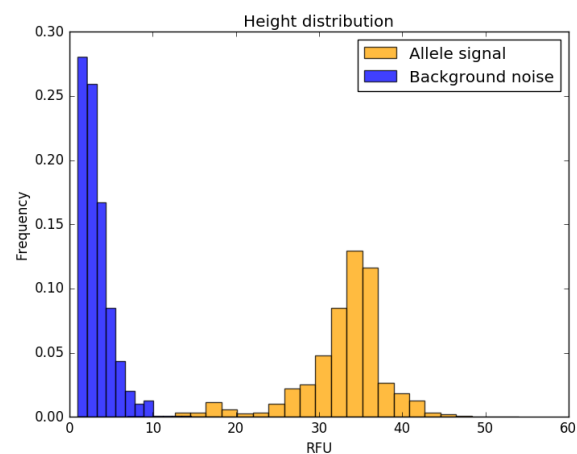
Sim 1 – 5s, 28 cycles

Allele signal: 1 copy, Background noise: 0.008ng

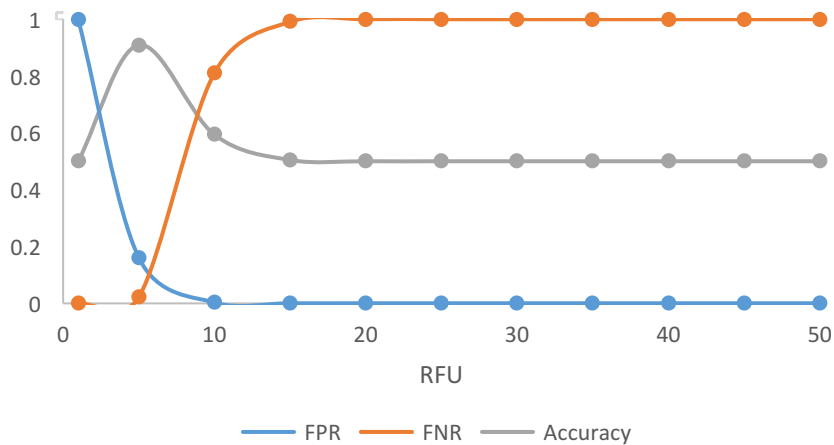


Sim 4 – 10s, 29 cycles &

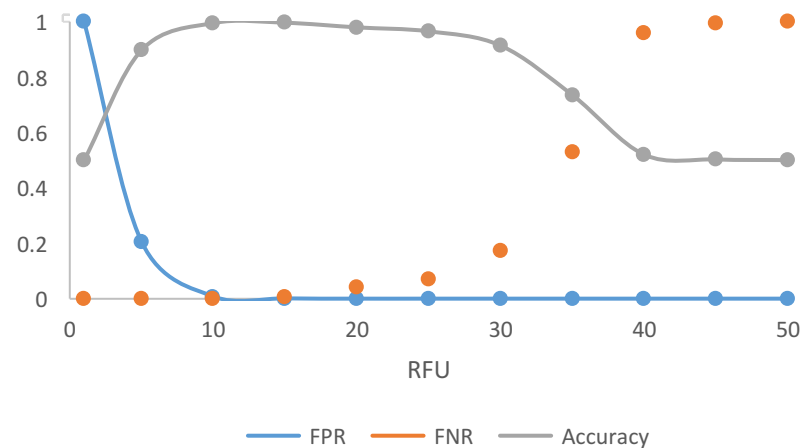
Allele signal: 1 copy, Background noise: 0.008ng &



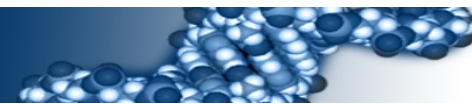
Error rates of AT



Error rates of AT

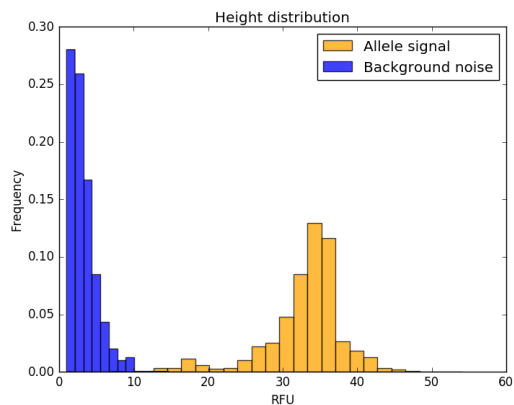


Results: D8S1179 – 1000 simulations &



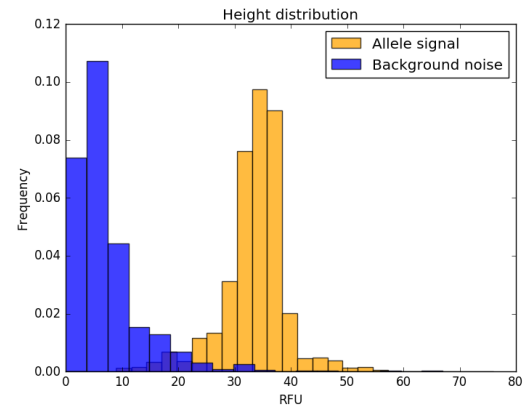
Sim 4 – 10s, 29 cycles

Allele signal: 1 copy, Background noise: 0.008ng

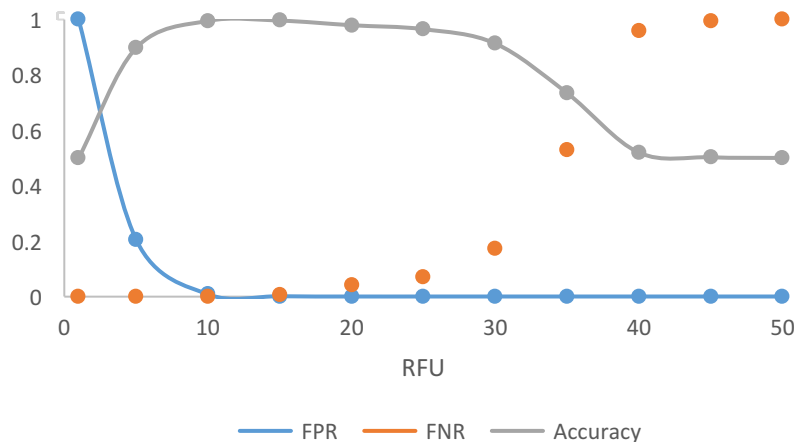


Sim 5 – 10s, 29 cycles &

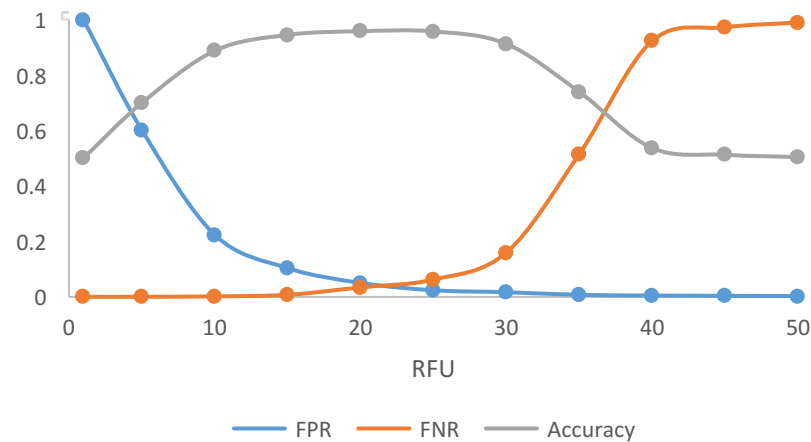
Allele signal: 1 copy, Background noise: 0.25ng



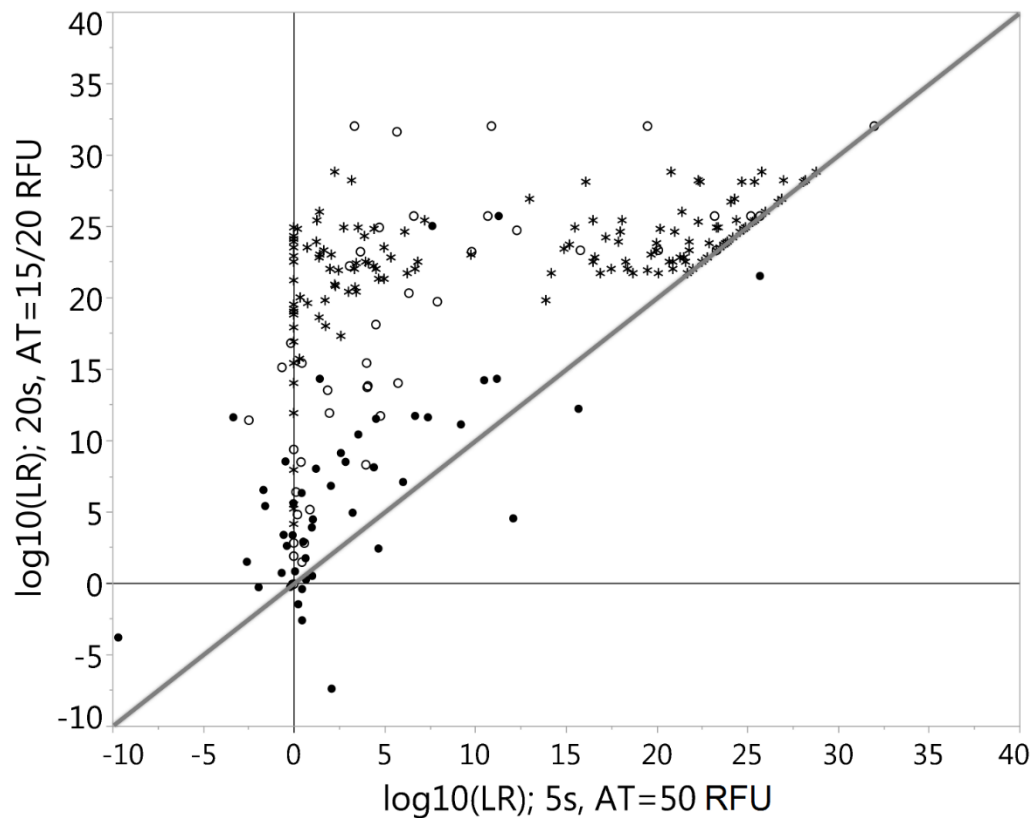
Error rates of AT



Error rates of AT

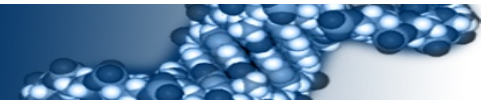


Impact of Information Content on Low-Template Probabilistic Interpretation* &



*Production of High-Fidelity Electropherograms Results in Improved and Consistent DNA Interpretation: Standardizing the Forensic Validation Process, Kelsey C Peters, et al. Forensic Sciences International: Genetics, Submitted.

Conclusions &



- Achieving signal_{1-copy}-to-noise resolution increases information content imported into LR calculation systems
- Choosing a condition-specific AT and laboratory parameters will maximize signal-to-noise resolution while simultaneously minimizing detection error rates
- A combined experimental & simulation-based approach makes the validation process fast and inexpensive