

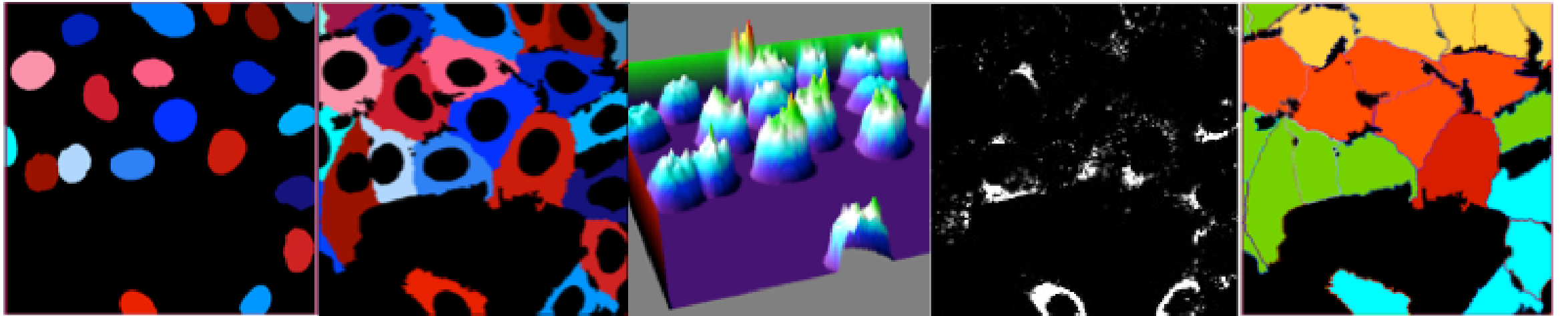
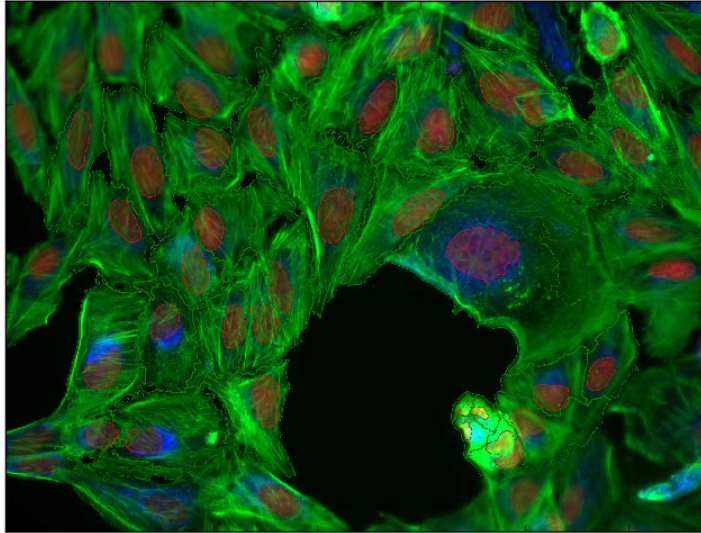
A fluorescence microscopy image showing a dense population of cells. The cytoplasm of the cells is stained green, while the nuclei are stained red. There are also blue-stained structures, possibly representing specific organelles or protein localization. The overall appearance is that of a tissue microenvironment.

Imaging Data Provenance and Reproducibility

*Tissue Microenvironment Group
Cardiff University, School of Medicine*

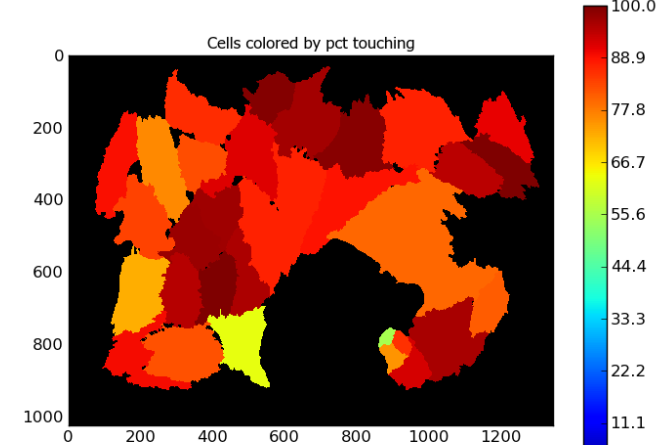
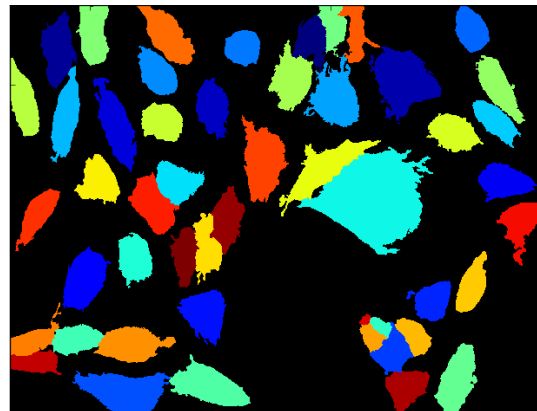
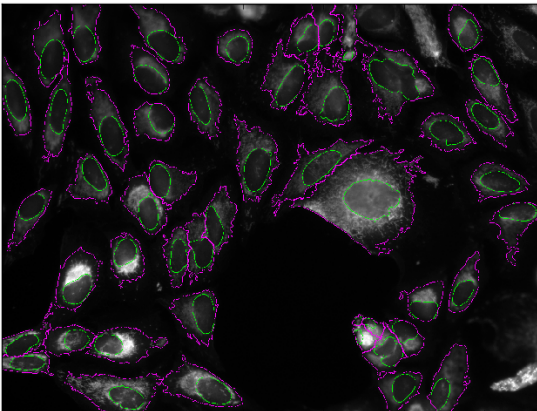
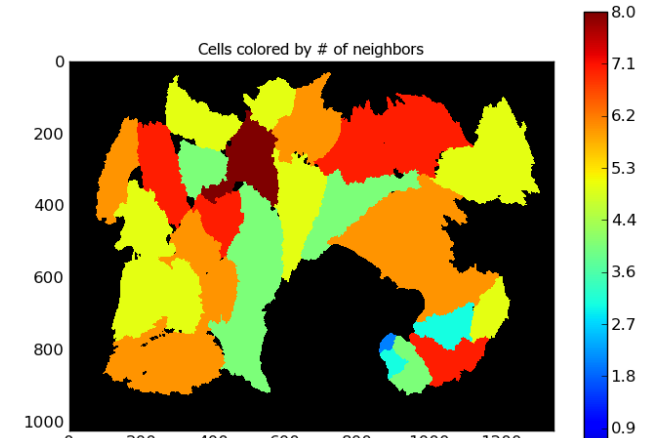
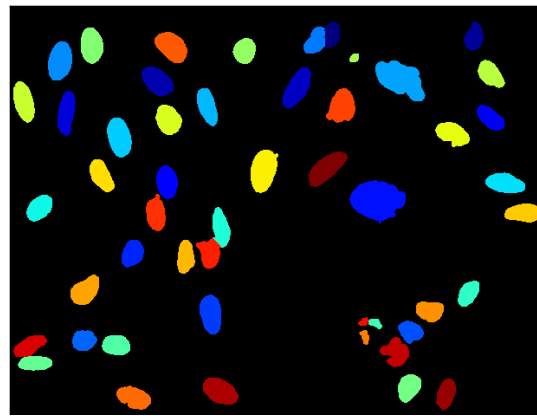
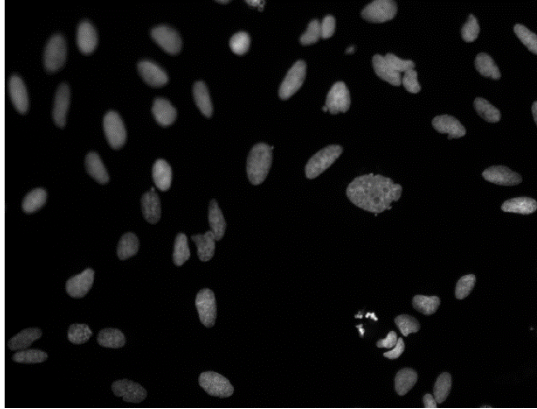
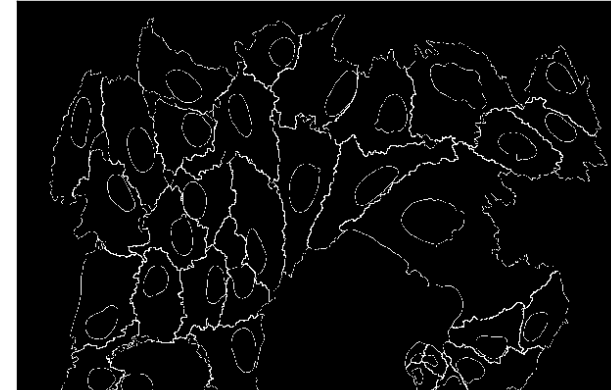
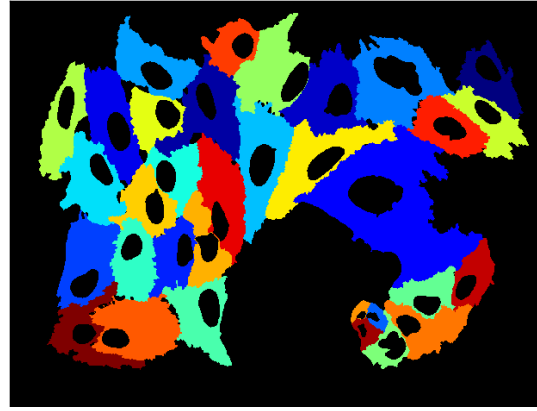
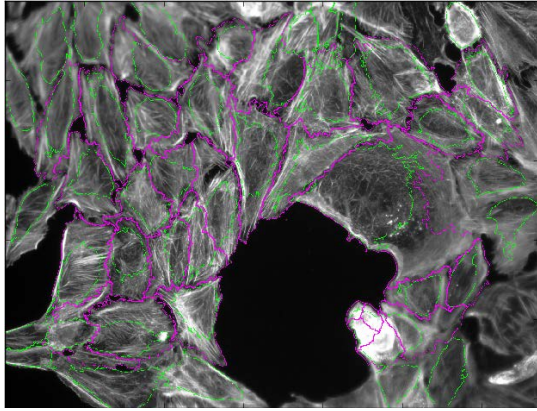
Dimitris Parthimos

Typical analysis pipeline

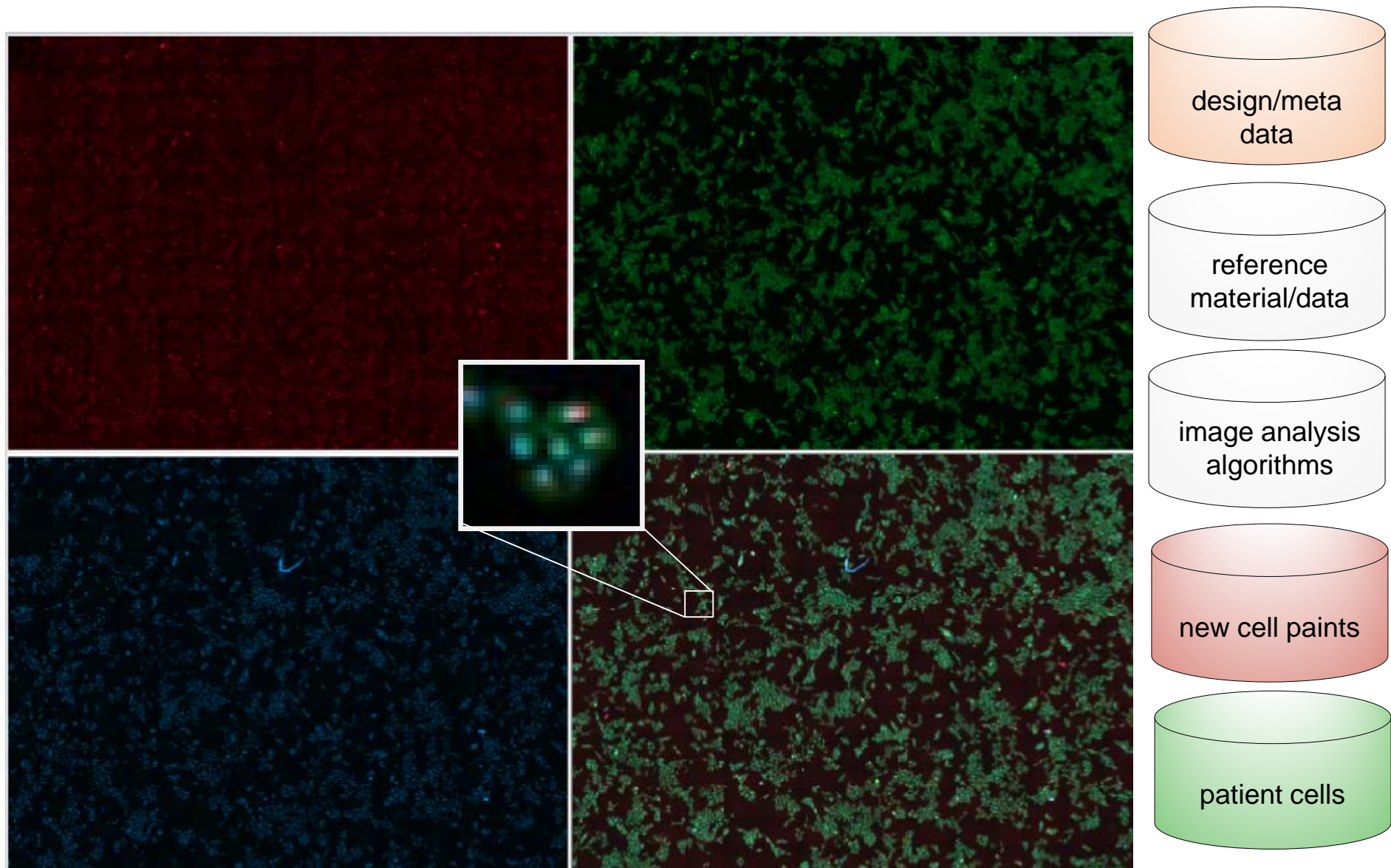


Counts, Sizes, Shapes, Intensities, Textures, Correlations, Neighborhoods

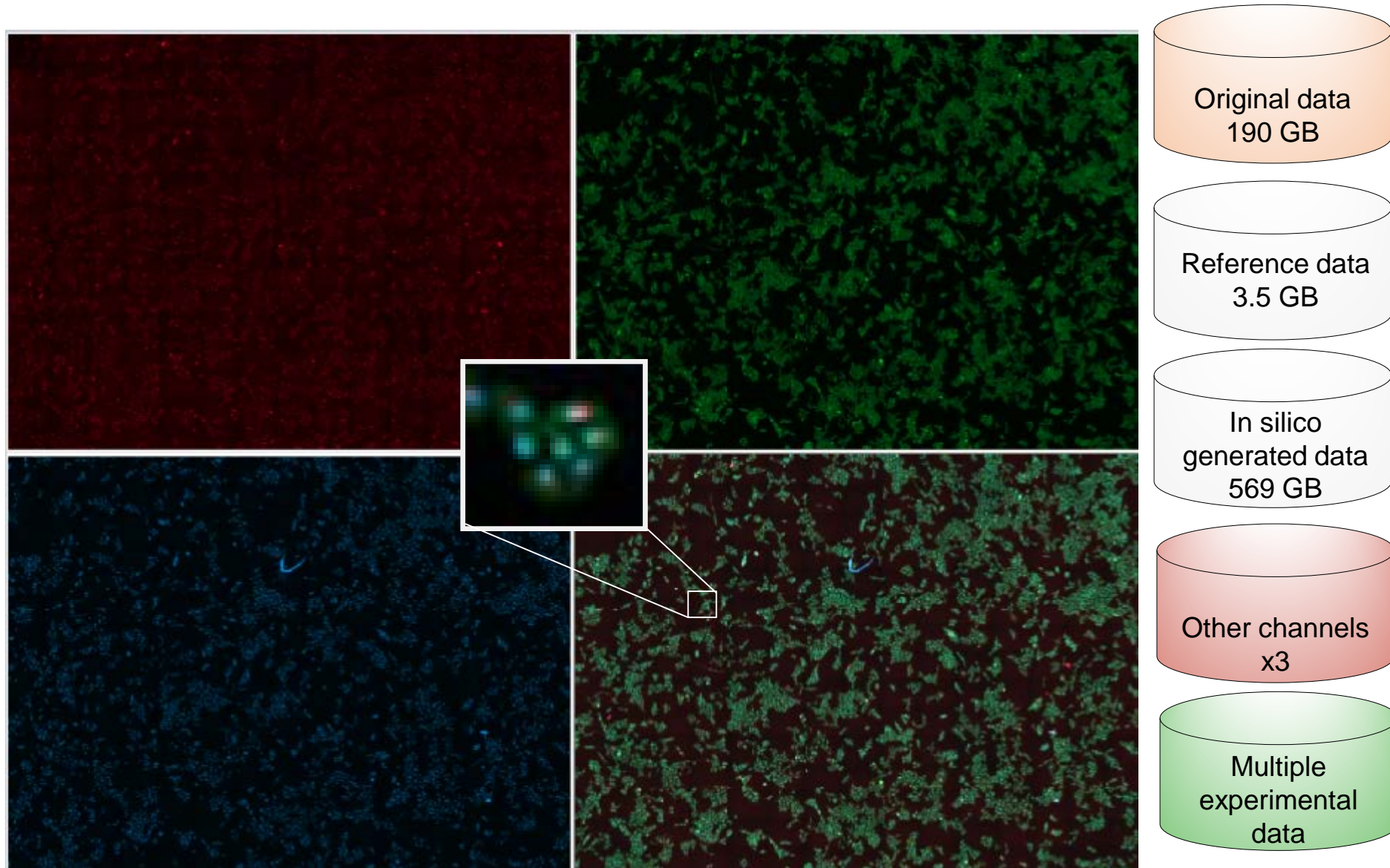
Image analysis - Pipeline in Action (CellProfiler)



Data collected at NIST: Static data



Data collected at NIST: Testing stability of data



Case Study 2

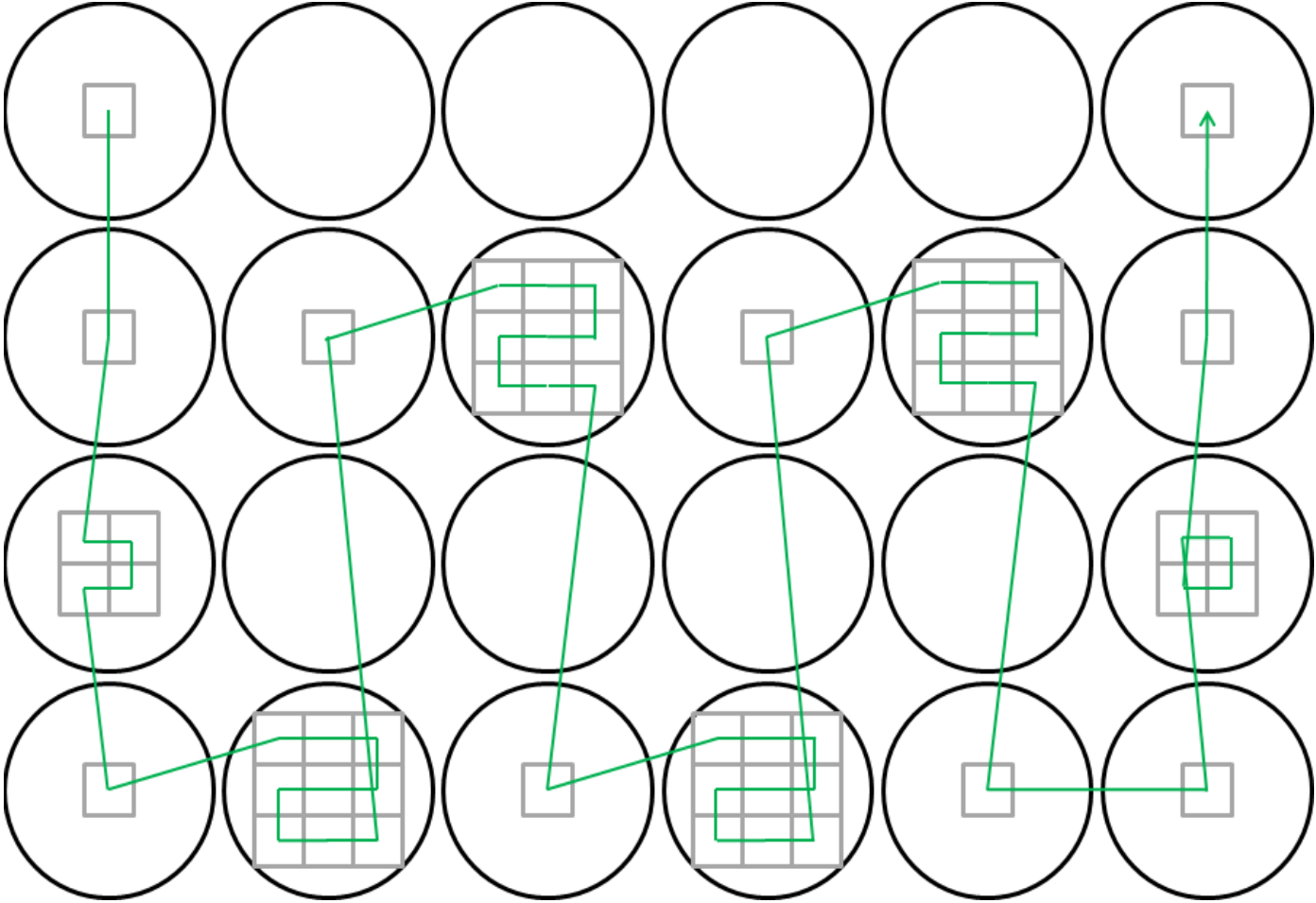
Timelapse microscopy of live cells responding to drug

R.A. Howard-Jones, Marie Wiltshire, A.J. Sloan and R.J. Errington
Cardiff University, School of Medicine,
M.R. Brown and Matthieu Duteil
Swansea University, School of Engineering

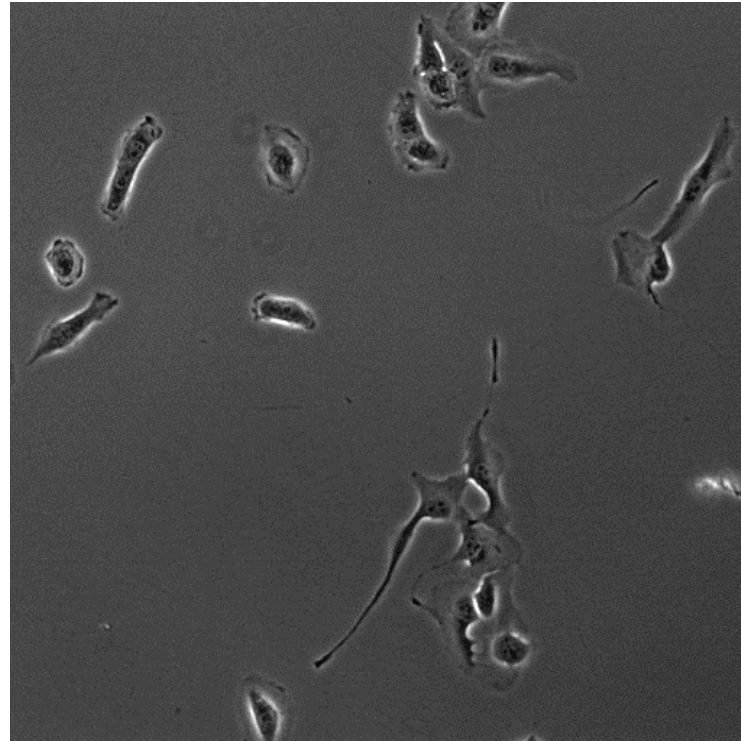
*J Elliott and Michael Halter
Cellular Biosystems, NIST*

*Peter Bajcsy and team
Information technology, NIST*

Collected on a high-throughput screening instrument designed to automatically collect an image sequence as a movie



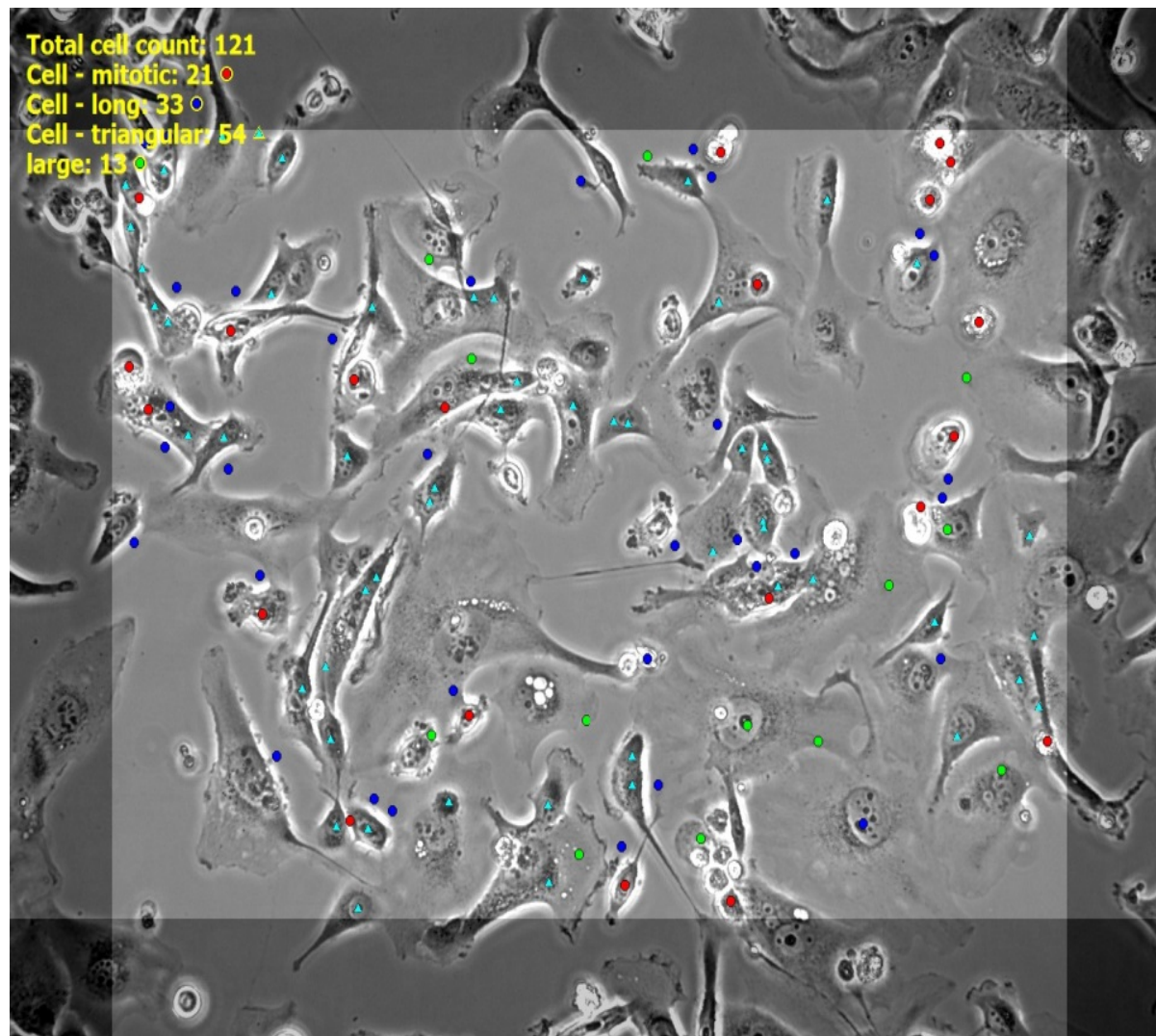
Typical video time lapse of cancer cell line



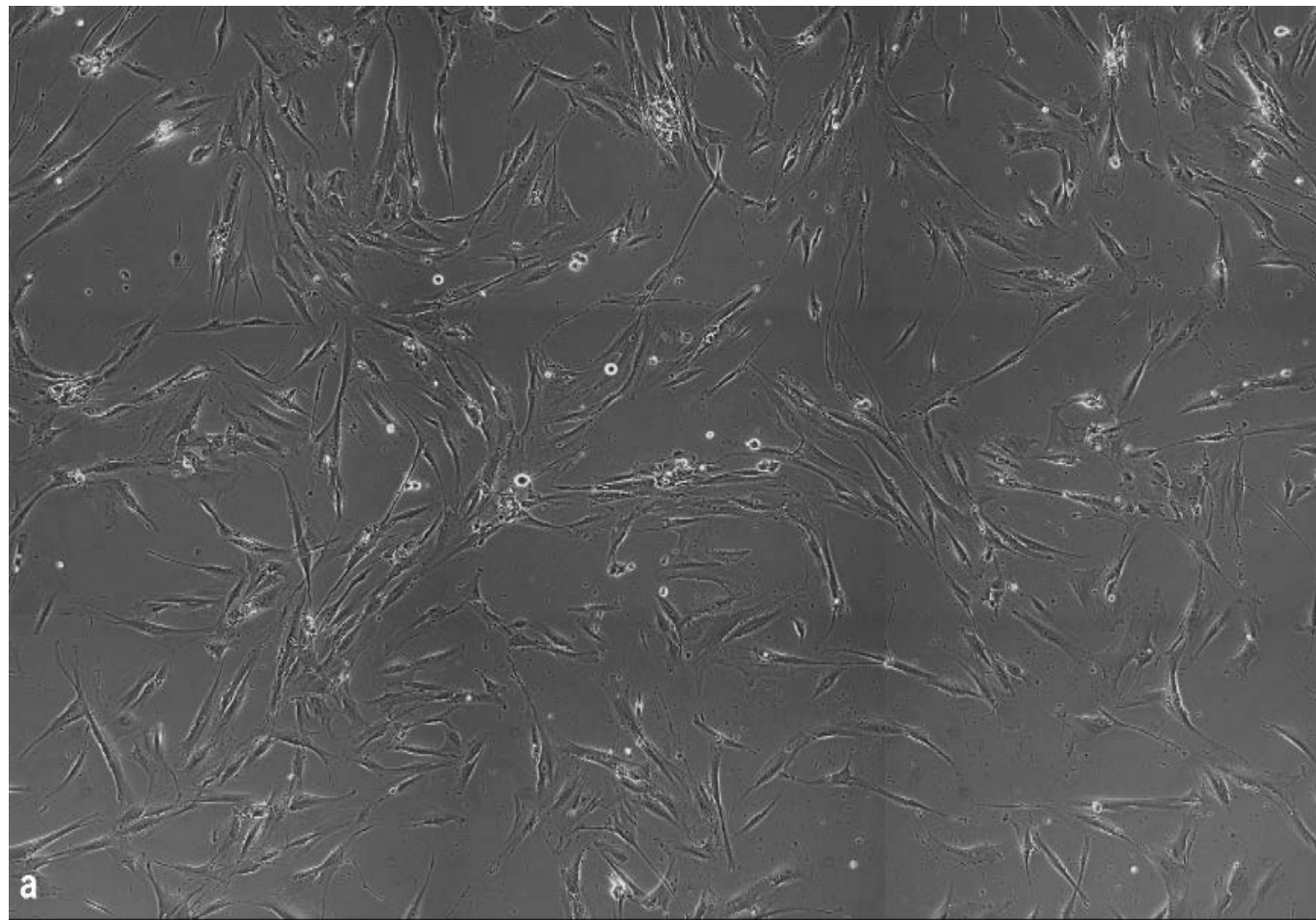
What do we want to extract from such timelapse image data

- Derive behavioural features of cells in control and perturbed conditions
- Obtain features of clonal expansion and understand this expansion at different granularities, including lineages
- In cancer cells: understand the derivation (dynamics) of the polyploidy phenotype, and measure the neighbourhood in which these cells survive.
- Patterns of drug resistance, temporal and spatial characteristics of which are poorly understood

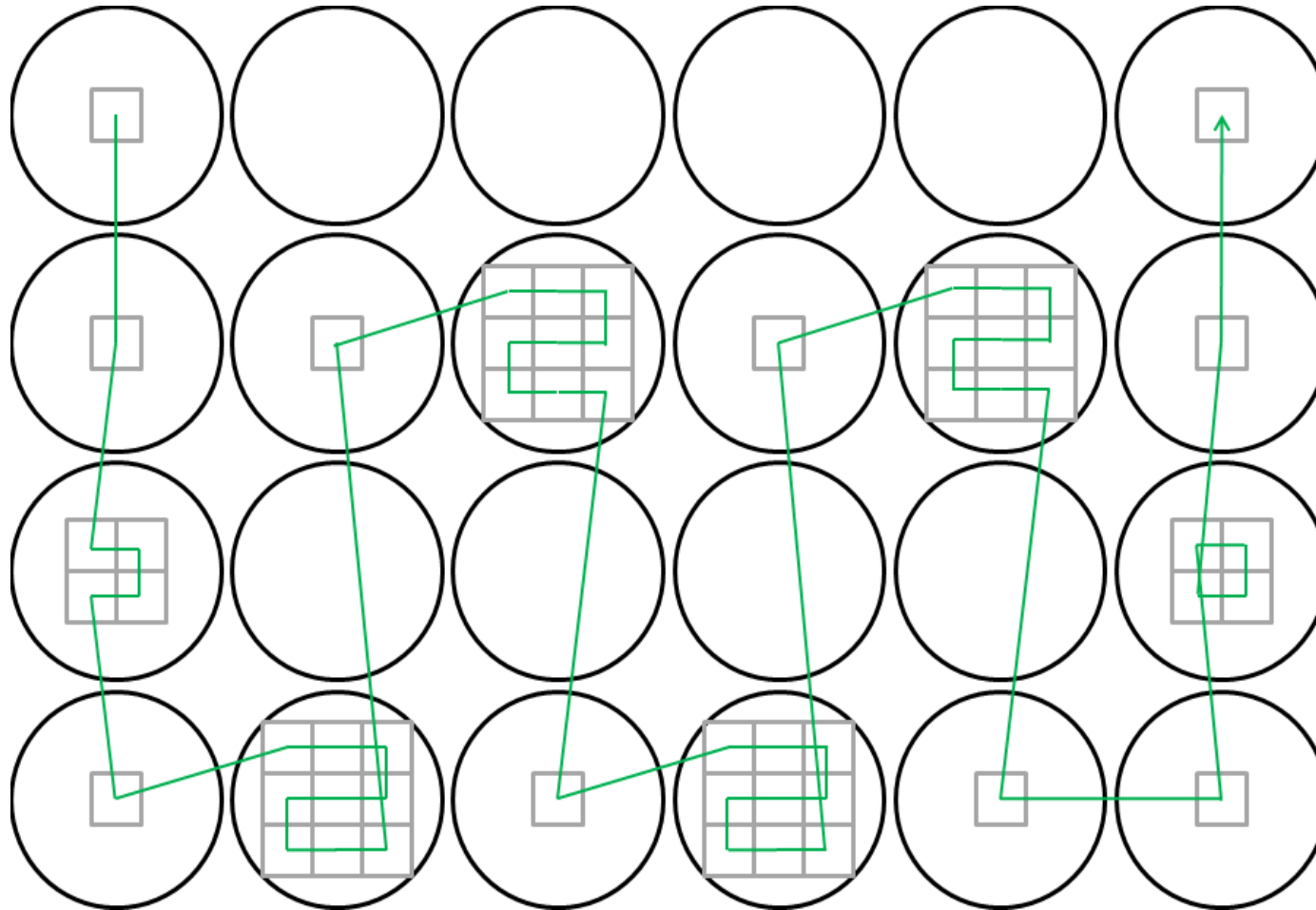
Automated cell detection and tracking



Tiled 3x3 FOV



Collected on a high-throughput screening instrument designed to automatically collect an image sequence a movie



For this one experiment: 108 FOV collected every 30 minutes for 120.5 hours: 2 channels
Each FOV = 1392×1040 therefore entire dataset is 72 Gigabytes

2 datasets one for cancer cell line and one for primary mesenchymal stem cells – 144GB

Case Study 3

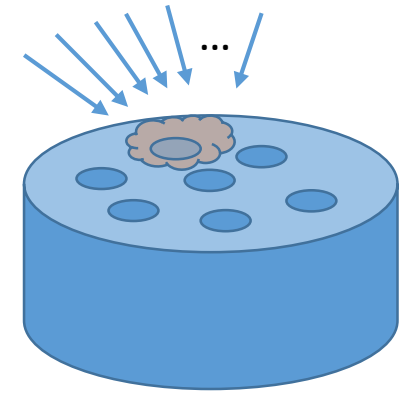
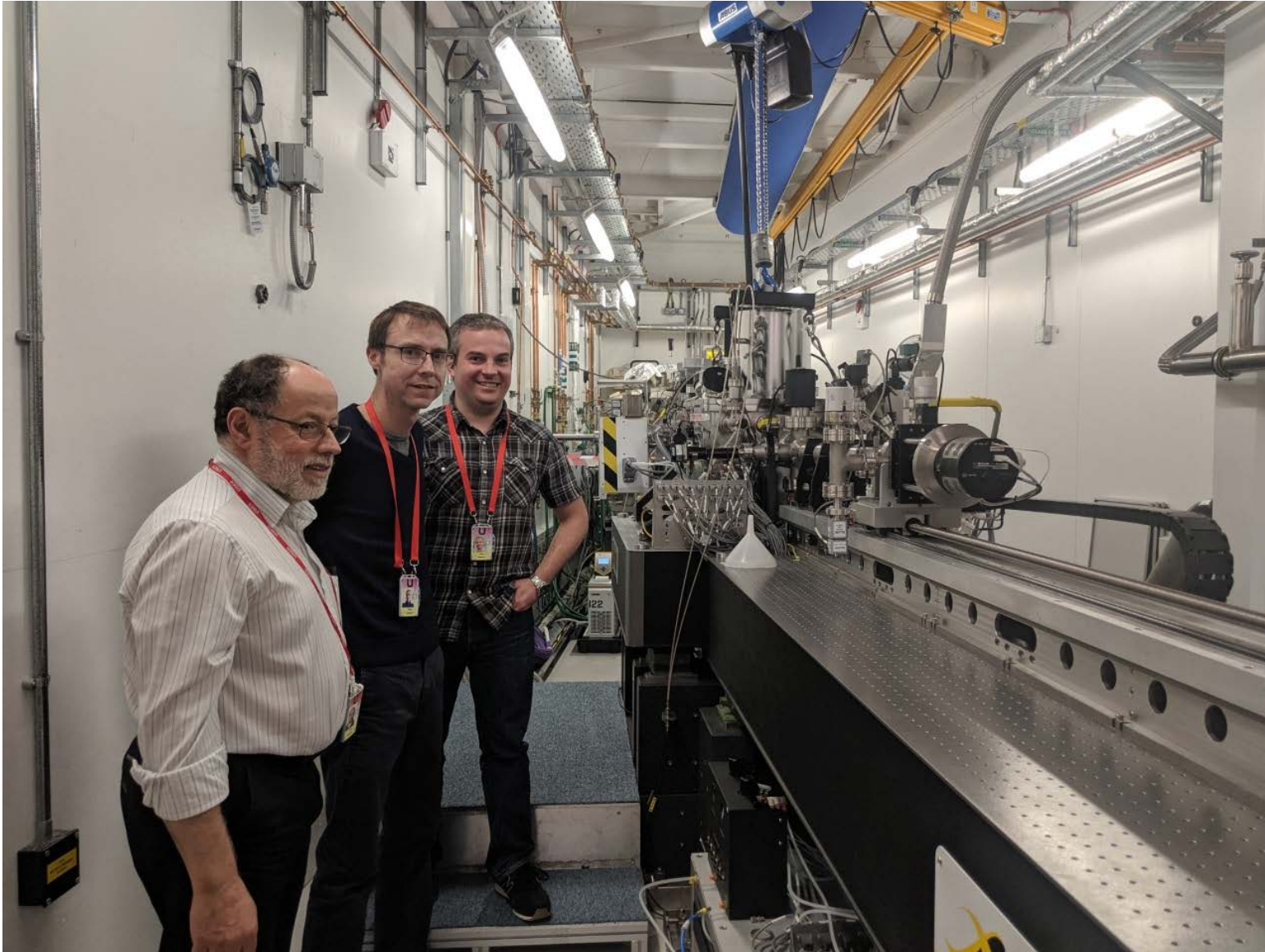
Extracellular vesicle activity associated with cancer resistance

Aled Clayton, Jason Webber
Cardiff University, School of Medicine,

Diamond

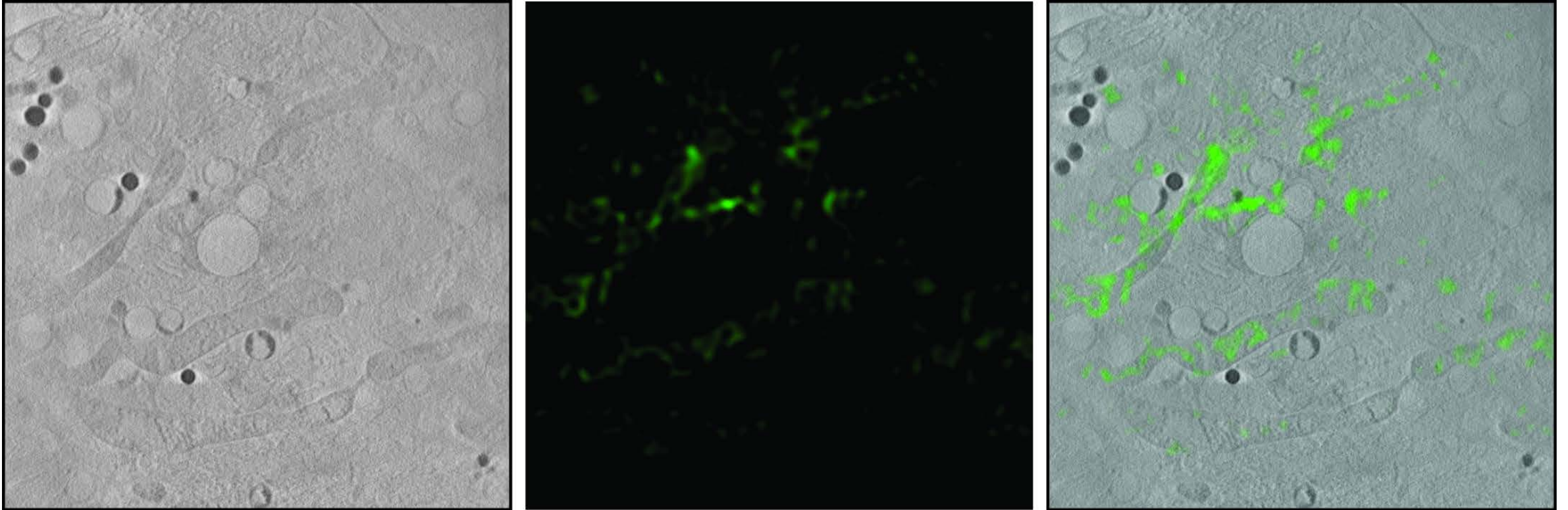


Fluorescent microscope overlaid with x-ray beam



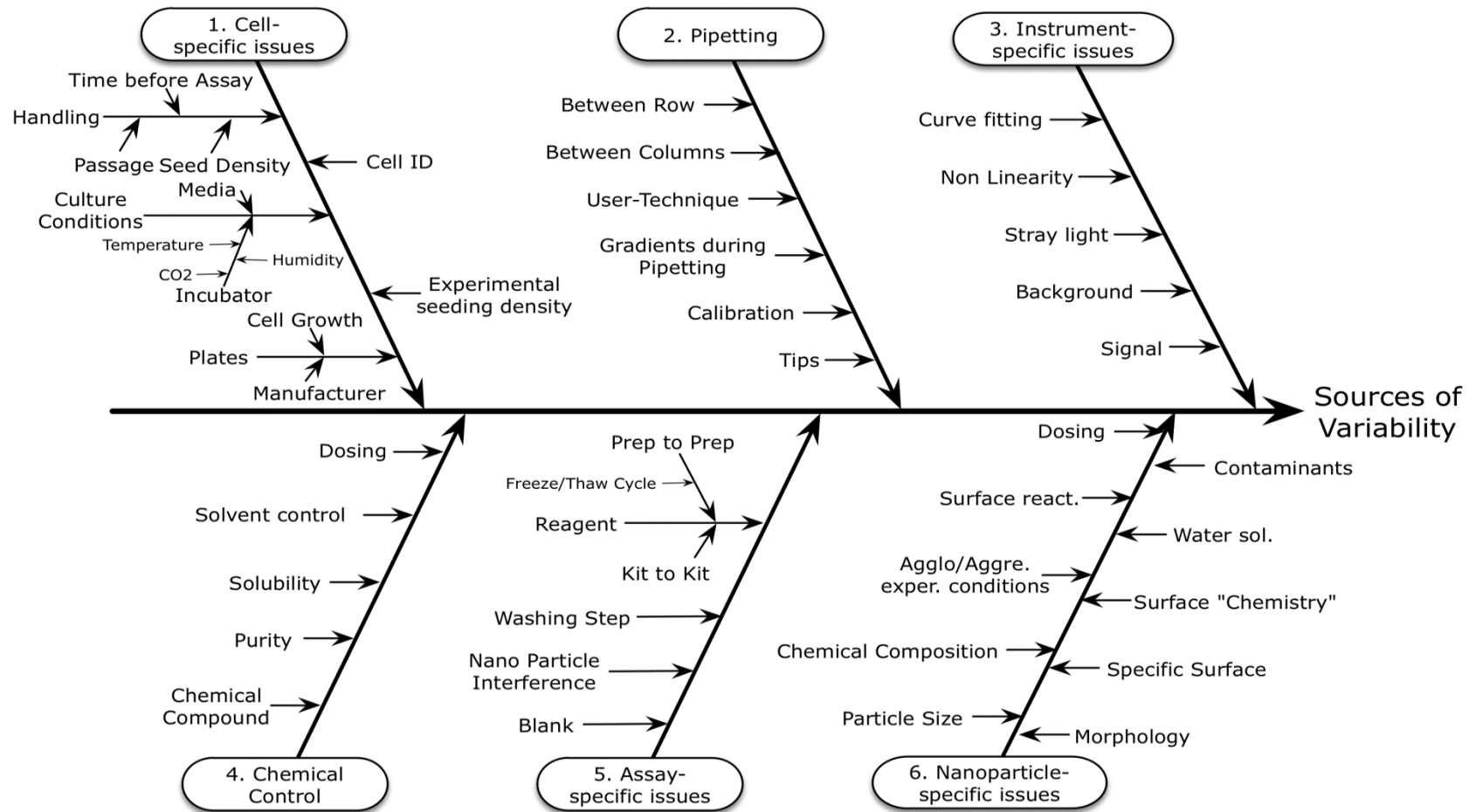
- *Cryo Soft X-ray Tomograph*
- *Cryo Structured Illumination Microscope*

Fluorescent microscopy overlaid with x-ray beam image



$\pm 60^\circ$ at 0.2° equivalent to 1.2 GB data size (x 100 cells x treatments)

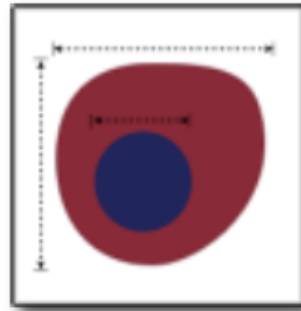
Identify sources of variability in assay



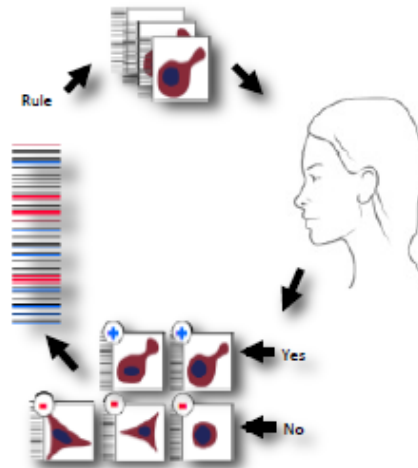
Cause and effect diagram

Three waves of quantitative image analysis

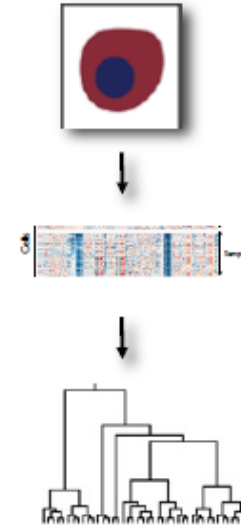
1 Measure known phenotypes



2 Train for known phenotypes



3 Profile to characterize samples



Measure everything, then use machine learning to distinguish a phenotype of interest



NIST- PABL

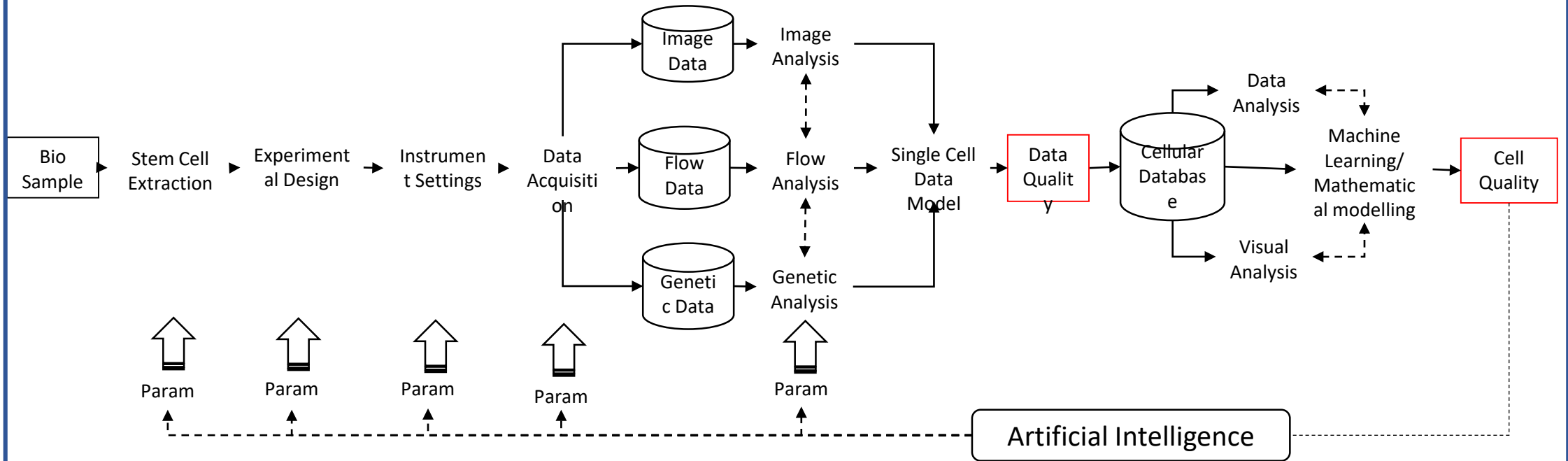


ProgeniDB
A progeny based cell lineage database

Cell-o-Pane

WET LAB

DRY LAB



Summary

- Bioimaging community faces an explosive growth in data size and complexity
- Reproducibility is a key concern; traceability of algorithm pipelines
- Fundamental need for tools such as Web Image Processing Pipeline (WIPP) and Automated Bio-Imaging Laboratories (NIST)
- Interactive measurements and AI-assisted discoveries over large image banks