



# Overview of the Ohio Wastewater Monitoring Network

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# Ohio Wastewater Monitoring Network (OWMN)

## Goal

- Monitor trend of SARS-CoV-2 RNA at specific locations (vs compare sites)
- Serve as early indicator of COVID-19 community spread
- Prioritize resources

## Statewide network

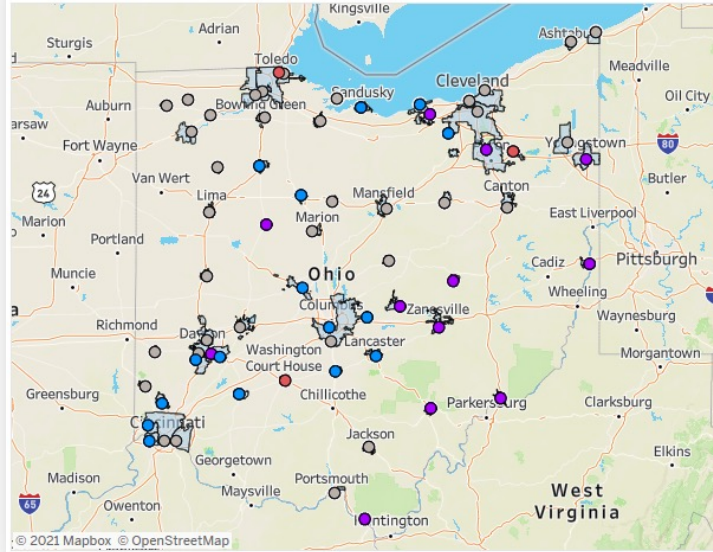
- Started July 2020
- leveraged expertise and resources
  - Ohio Universities
  - US EPA-ORD

## 67 locations twice a week

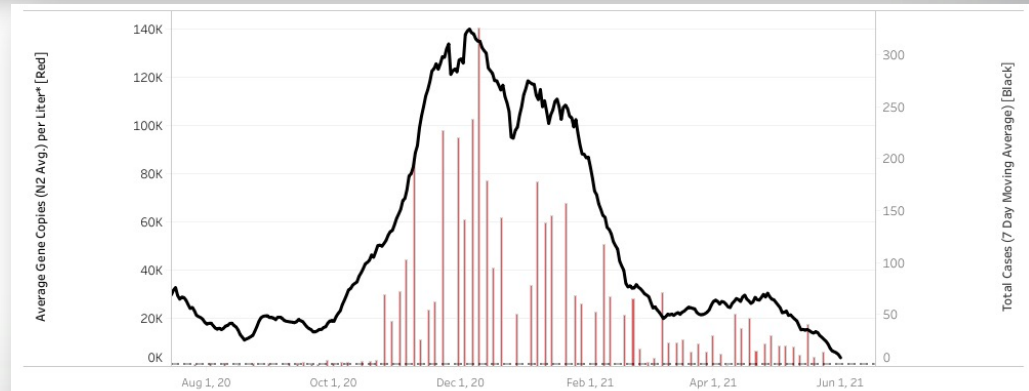
## Sequencing to screen for possible presence of SARS-CoV-2 variants

- Variants of Concern (VOC)
- Variants of Interest (VOI)

Wastewater Treatment Plant Locations and Boundaries



Athens WTP	↑
Eastern Regional WRF	↑
Newark WWTP	↑
Zanesville WWTP	↑
Mount Vernon WWTP	↑
Washington Court House WWTP	↑
Pataskala WWTP	↑
Franklin Hills WWTP	↑
Village of Ottawa WWTP	↑
City of Sidney WWTP	↑
City of Wilmington WWTP	↑
City of Eaton WWTP	↑
Lorain Black River WWTP	↑
Conneaut WWTP	↔
Elyria WWTP	↔
NEORS Southernly WWTC	↔
Toledo Bay View Park WWTP	↔



<https://coronavirus.ohio.gov/wps/portal/gov/covid-19/dashboards/other-resources/wastewater>



# Public Health Application

- **To serve as an early warning of infection in communities and an understanding of case trends**
- **The focus is on trends or significant changes in the number of viral gene copies detected.**
- **Currently action is taken when at least 3 samples show a sustained increase of at least 10-fold (1 log)**
- **State actions when increases are observed:**
  - Notify the local health district and utility
  - Provide information on how to interpret the data and link to message toolkit
  - Notify the state pandemic testing team for linkages to establish pop-up testing sites
  - Provide case data by sewer shed to local health district (this extraction to be provided soon)
- **Participation in the CDC National Wastewater Surveillance System**
- **Toolkit link: <https://coronavirus.ohio.gov/wps/portal/gov/covid-19/healthcare-providers-and-local-health-districts/for-local-health-districts-and-governments>**



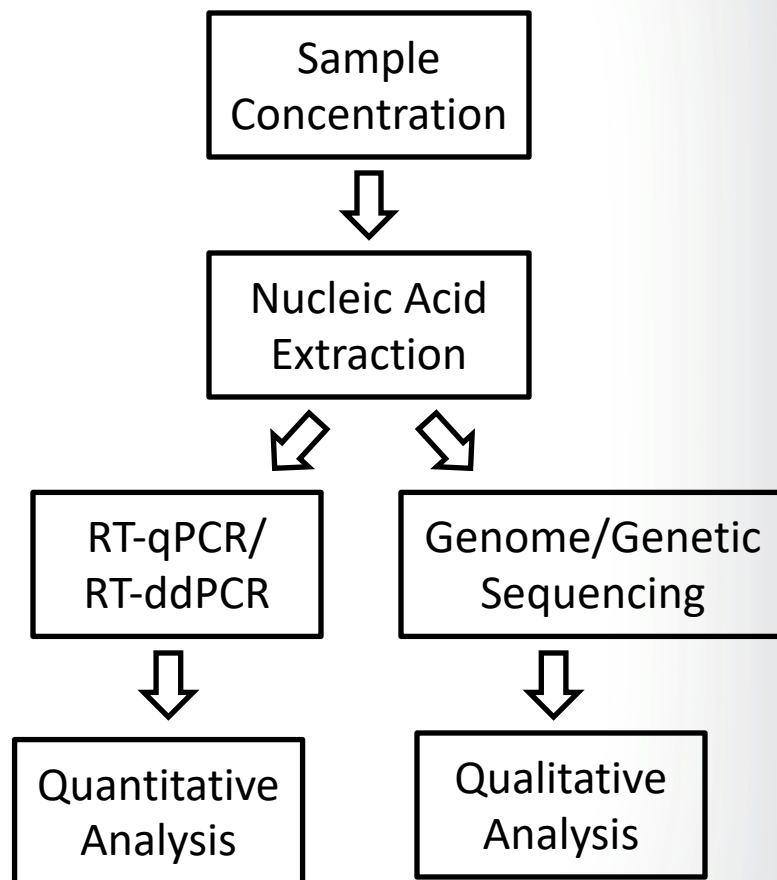
# Accomplishments

- **Built statewide network that represents wastewater flow from nearly 5 million residents**
- **Almost 1 year of weekly data collected**
- **All data is publicly available on the Ohio coronavirus dashboard and is updated daily**
- **Provided nearly 500 warnings to local health communities**
- **Expanded to include genomic sequencing of wastewater to pair with clinical data and inform public health decisions**



# Lab Method Logistics

- **Twice weekly samples**
- **Report data within 2 days of sample receipt**
- **No prescribed method; labs decide**
- **Supply chain shortages**
- **Low target concentration**
- **Sample hold time: 4°C - 72 hours**
- **No sample pasteurization**





# Quality Assurance/Quality Control

- **Matrix Spike to assess method recovery efficiency**
  - Coronavirus recommended: human (OC43), murine (MHV), bovine (BCoV)
- **Inhibition control to monitor for PCR amplification inhibition**
- **RT-qPCR standards/RT-ddPCR positive control**
- **Human fraction measurements**
  - crAssphage
  - Pepper mild mottle virus
- **Monthly Interlaboratory Method Validation**
  - Pick a site with sufficient concentration of SARS-CoV-2
  - Each lab gets 0.5 L
  - Each lab processes and analyzes sample
  - Report data to Project Coordinator





# OH Network Lab Methods

LAB	LOD (copies/L)	Processing Method	Nucleic Acid Extraction	Quantitative Analysis Method	RT-PCR Standard Curve/ Control	Inhibition Control	Matrix Spike	Fecal Indicator
A	850	Centrifugation, filtration	Qiagen RNeasy PowerWater Kit	RT-qPCR	DNA plasmid	Dilution	MHV	crAssphage
B	135	Centrifugation, filtration	Qiagen Allprep DNA/RNA Kit	RT-qPCR	DNA plasmid	Dilution	BCoV	crAssphage
C	133	Tween, solids removal hollow fiber ultrafiltration (InnovaPrep)	Qiagen PowerMicrobiome Kit	RT-ddPCR	DNA plasmid	Luciferase Control RNA	OC43	crAssphage
D	7,440	Filtration	Trizol, garnet bead beating, alcohol precipitation	RT-qPCR	DNA plasmid	Luciferase Control RNA	BCoV	PMMoV
E	500	Centrifugation, filtration	Trizol and RNA purification kit	RT-qPCR	Synthetic RNA	Luciferase Control RNA	BCoV	crAssphage
F	3,000	Promega, add protease, supernatant through GFA/silica column	Promega Wastewater Large Volume TNA Capture Kit	RT-qPCR	DNA plasmid	Promega probe	OC43	PMMoV
G	231	Centrifugation, filtration	Qiagen RNeasy PowerWater Kit/Trizol-chloroform	RT-ddPCR	SARS-CoV-2 genomic RNA	Luciferase Control RNA	OC43	crAssphage
H	891	Acidification, Filtration, extract filter	Qiagen Allprep PowerViral DNA/RNA Kit	RT-qPCR	Synthetic RNA	Mouse lung RNA	OC43	PMMoV



# OH Network Lab Methods

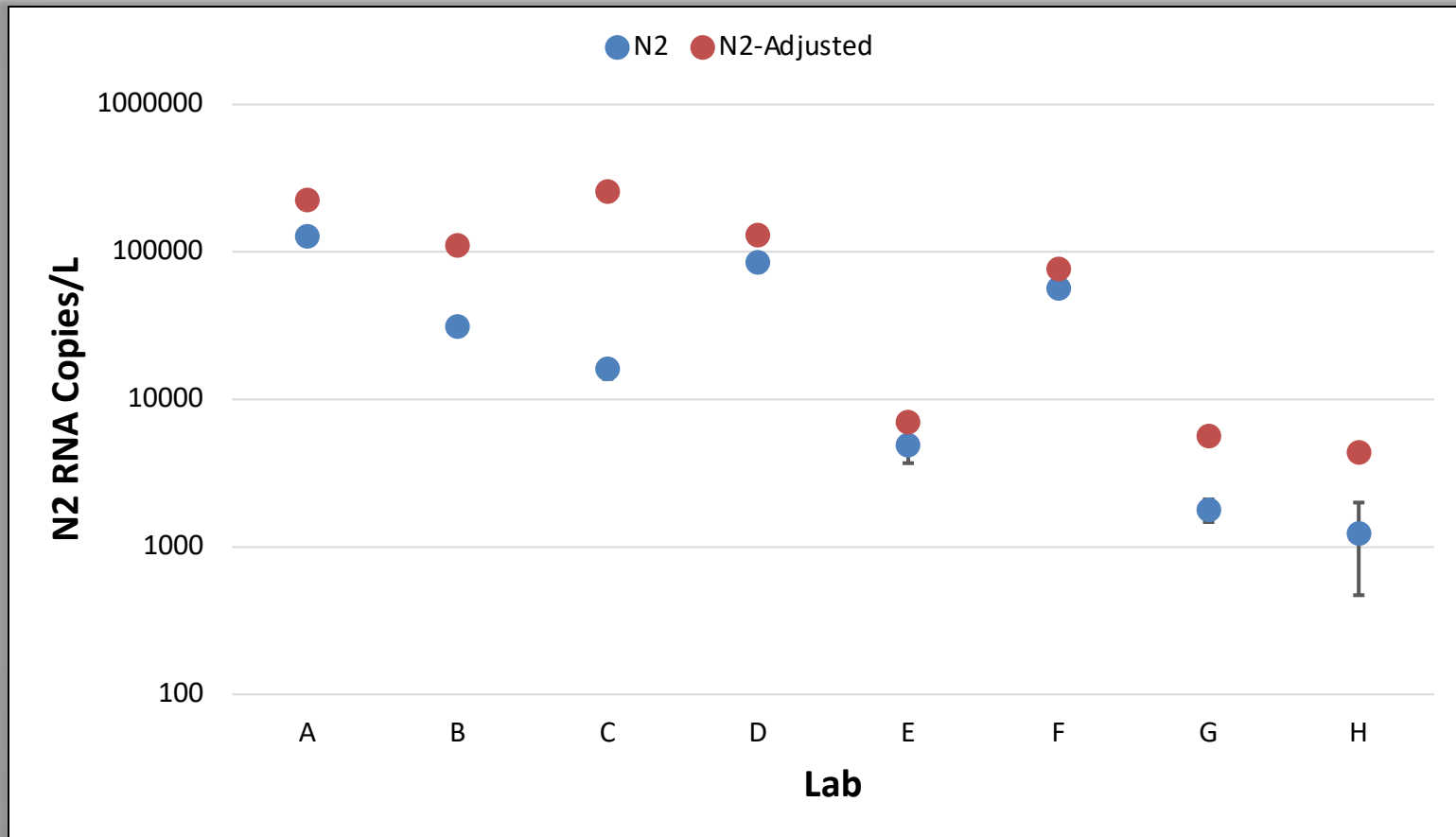
LAB	LOD (copies/L)	Processing Method	Nucleic Acid Extraction	Quantitative Analysis Method	RT-PCR Standard Curve/ Control	Inhibition Control	Matrix Spike	Fecal Indicator
A	850							
B	135							
C	133							
D	7,440							
E	500	5	8	2	3	4	3	2
F	3,000							
G	231							
H	891							

Number of different procedures employed at the various processing/analysis steps





# Validation Results (April 2021)



N2 RNA concentrations from 8 labs span >2 orders of magnitude

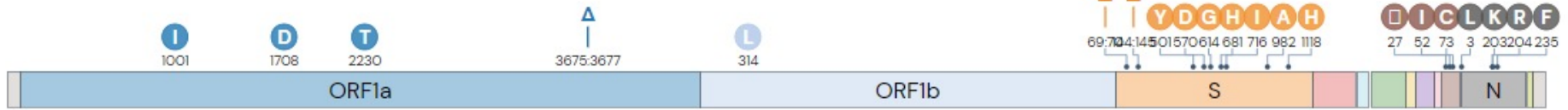


# RT-ddPCR Mutation Assays

## B.1.1.7

first identified in United Kingdom

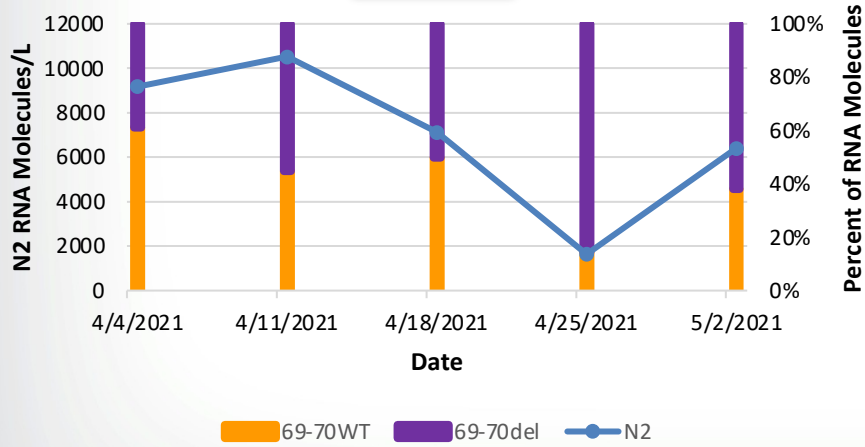
a.k.a. Variant of Concern 202012/01, VOC-202012/01, 20B/501Y.V1, 20I/501Y.V1



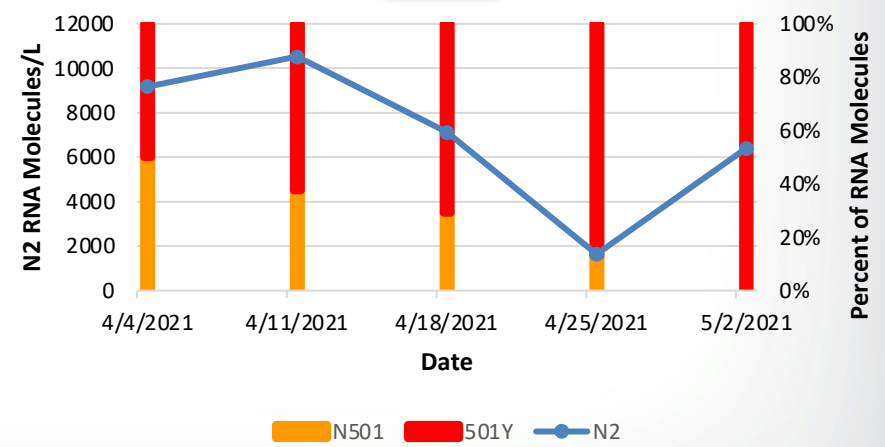
VARIANT OF CONCERN

<https://outbreak.info>

del 69-70



N501Y



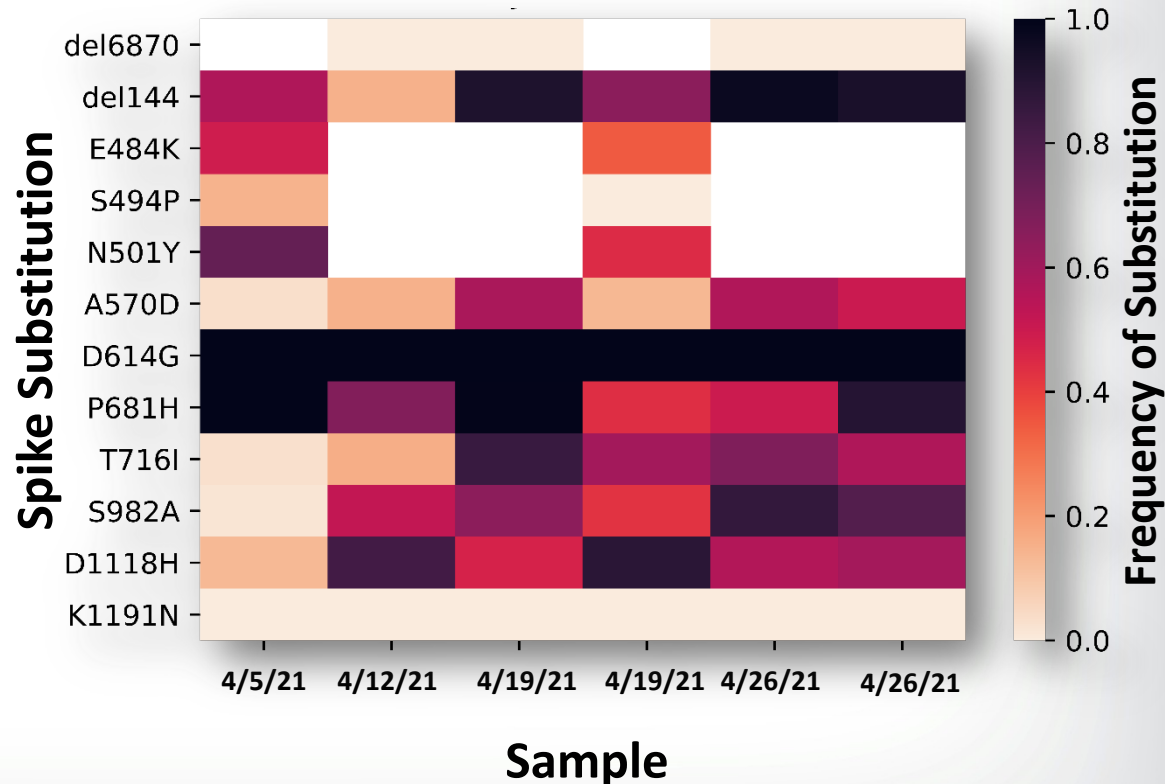
RNA sequences with deletions of nucleotides that result in absence of spike aa 69-70 increases over time

RNA sequences with nucleotides that change spike aa 501 increases over time



# Genome/Genetic Sequencing

- **Pooled sample**
  - Cannot assemble a genome
  - Focus on mutations that cause amino acid substitutions, signatures of VOC/VOI
- **Genome/Genetic Sequencing**
  - 3 labs, different methods
  - Tiled amplicon approach
  - Short read seq via Illumina
  - Short term - spike amino acid changes for CDC's VOC/VOI
  - Report (for each site)
    - Read depth
    - Number of alternative alleles





# OH Network Insights

- **How do current practices (eg. methods, protocols, technologies, best practices, etc.) successfully contribute toward comparable, high quality data/results/decisions?**
  - Using a consistent method, trends of SARS-CoV-2 RNA in a sewershed can be evaluated
  - Implementation of Quality Control parameters allow for confidence in lab measurements
  - Frequent communication/regular meetings facilitate interlab discussion and troubleshooting
- **How do current practices compromise efficiency and reduce confidence in data/results/decisions?**
  - Too many labs/methods result in measurement variation
  - Varied experience leads to measurement variation
  - Supply shortages lead to method changes
- **What is needed to increase comparability and confidence in data and results?**
  - Standardized methods/procedures
  - Standardized quality control samples/reagents
  - Statistical models to quantify uncertainty
- **What types of standards could potentially help to fill these needs?**
  - Matrix Spike
  - Extraction controls
  - RT-qPCR standards
  - RT-ddPCR controls
  - Inhibition controls
  - Sequencing controls



# Research Team and Partners

## **EPA/ORD**

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## **Utilities**

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**Montgomery County**

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**City of Hamilton**

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**City of Springfield**

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