



# SURFing with Green Tea: What Neutrons Read in Tea Leaves

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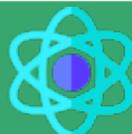
# OUTLINE



BSA & Polyphenols



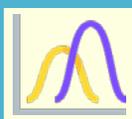
High Pressure



Neutron Scattering



Experimental Setup

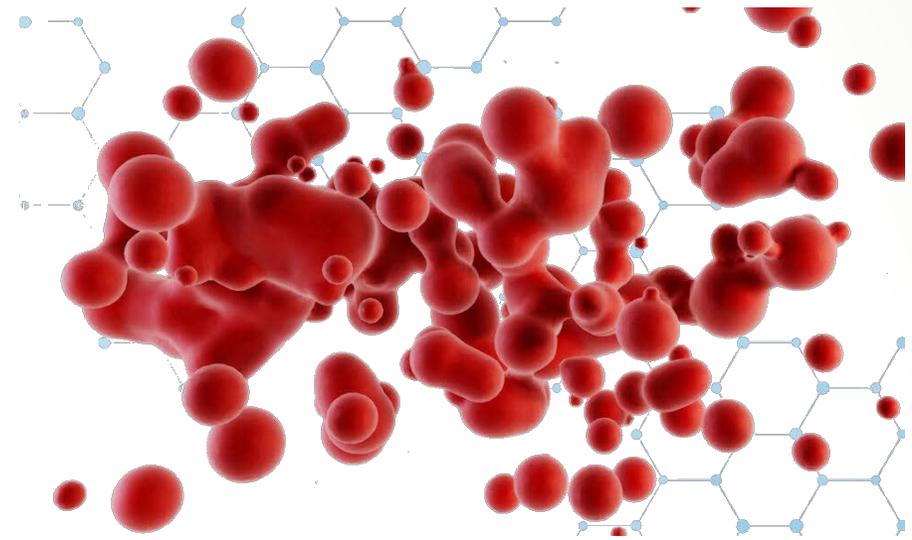


Results



Conclusion & Future Directions

# Bovine Serum Albumin

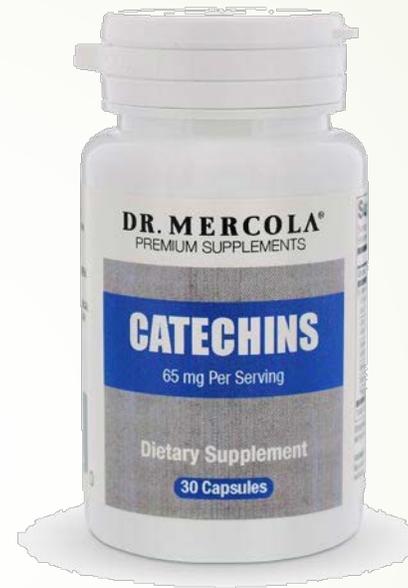
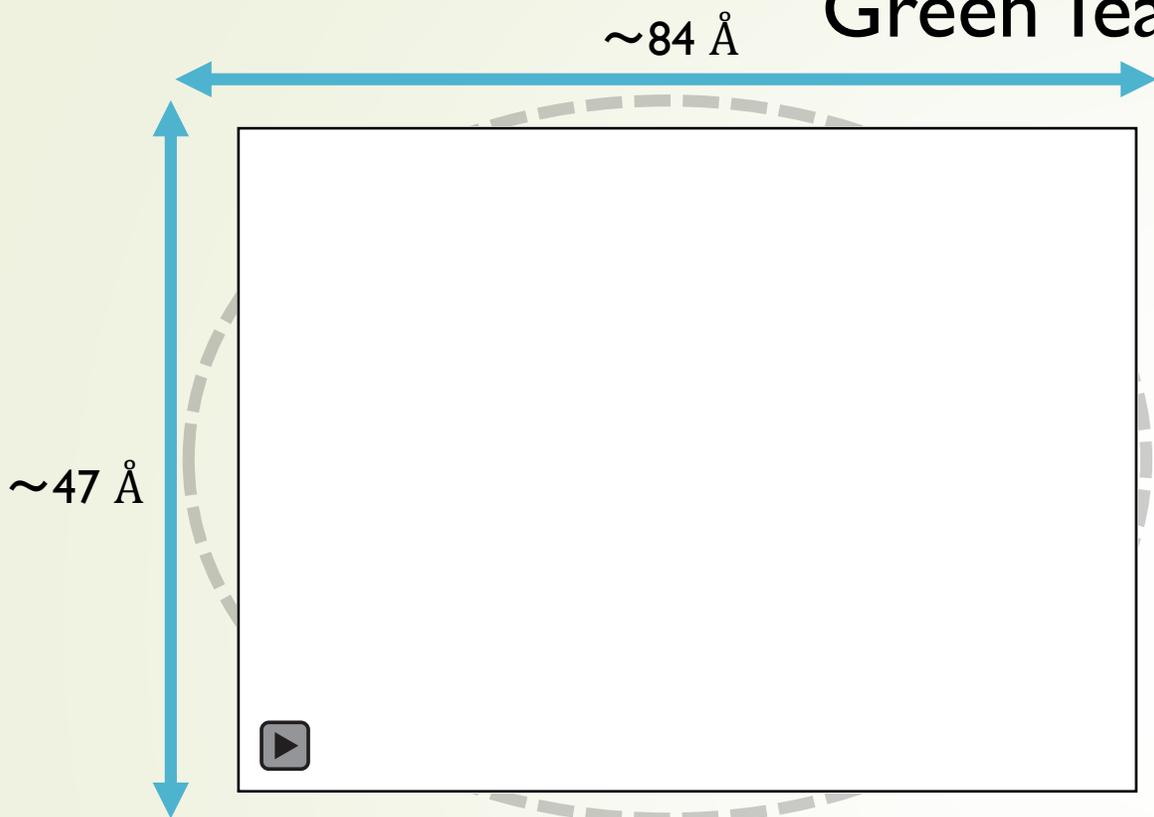


Electrostatic Potential (mV)

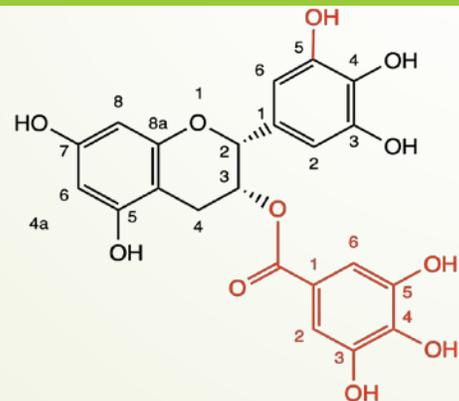
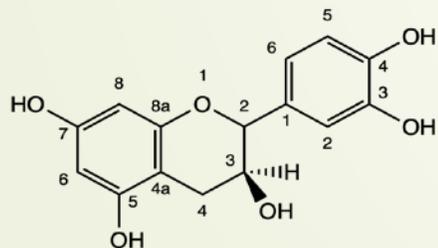
Calculated using Adaptive Poisson-Boltzmann Solver (APBS) at pH 7.

[Baker et al. (2001). PNAS 98, 10037-10041]

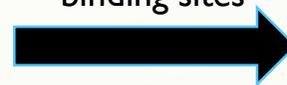
# Green Tea Polyphenols



**(+)-Catechin**    **(-)-Epigallocatechin Gallate (EGCG)**



Computational docking analysis binding sites



**Hydrophobic**  
(VDW + Nonpolar)

**Hydrogen bonding**

**Interactions**

-25.11 kCal/mol (EGCG)

-18.63 kCal/mol (Catechin)

7.49 (EGCG)

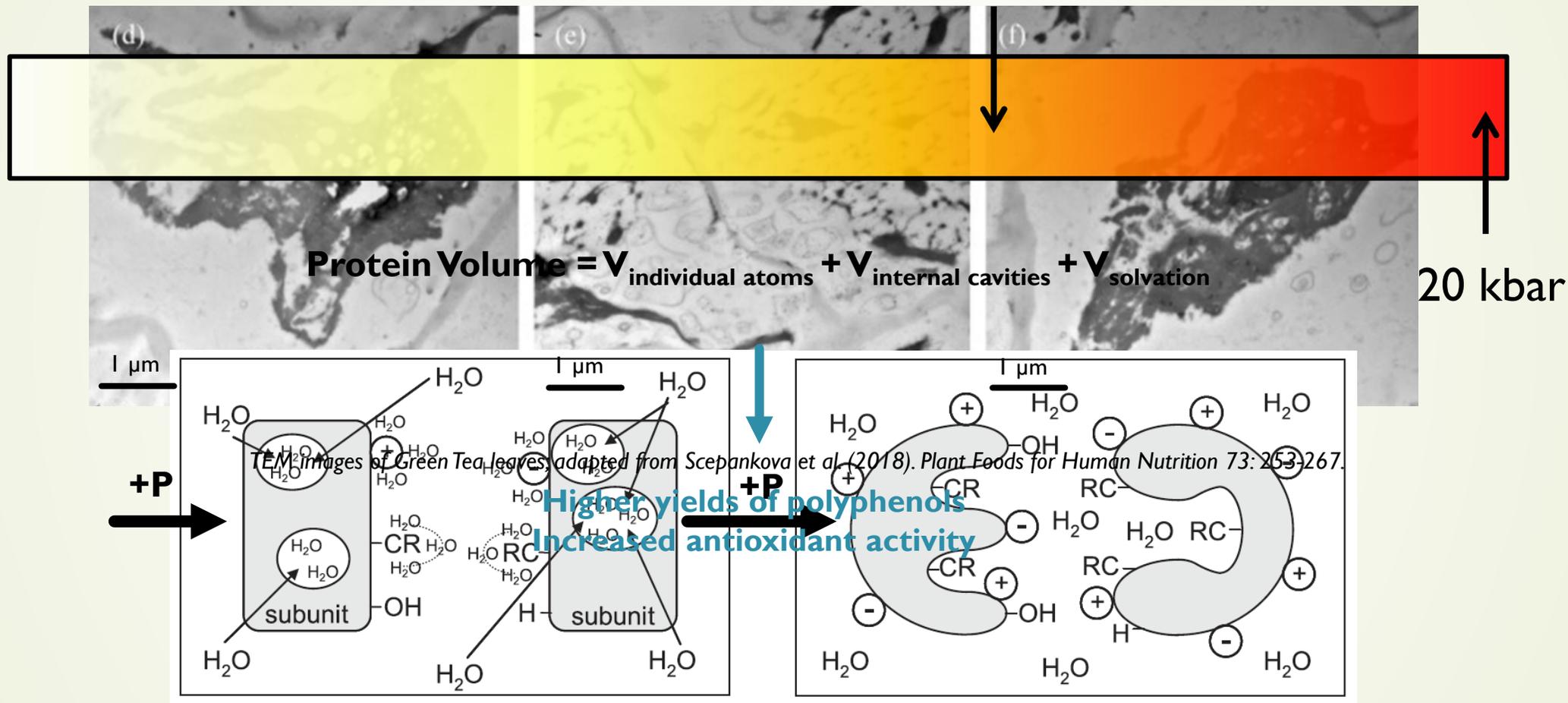
1.68 (Catechin)

[Tang et al. (2017). J. Agric. Food Chem. 65: 656-665]

# High Pressure Effects

No Treatment

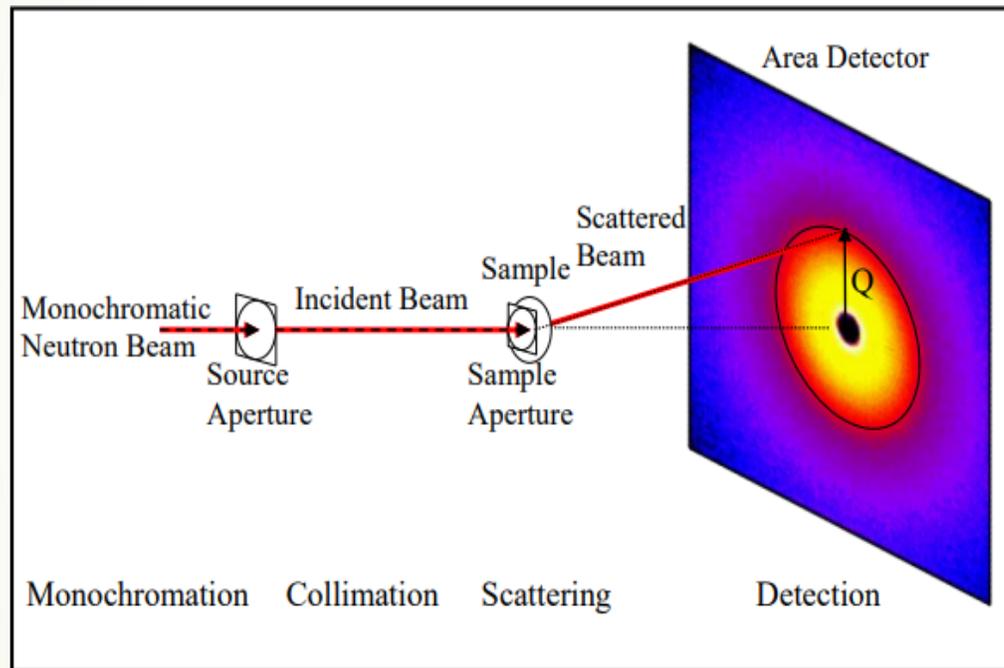
High Pressure 4 kbar High Temperature



Adapted from: R. Winter et al. (2007). *J. Non-Equilib. Thermodyn.* 32, 41–97.

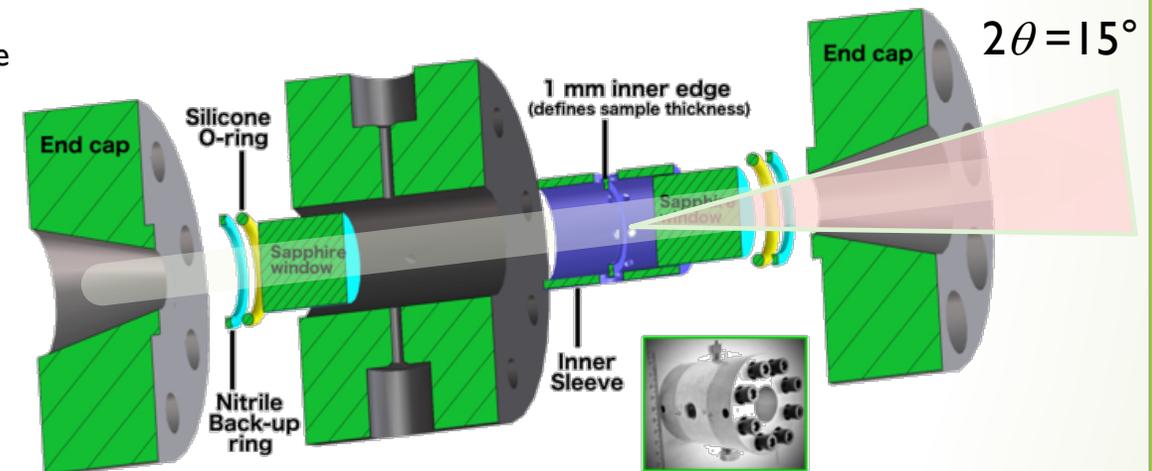
# Why SANS?

Structure, conformation of biomaterials  $\rightarrow$  Function and behavior



Reciprocal space

Real space



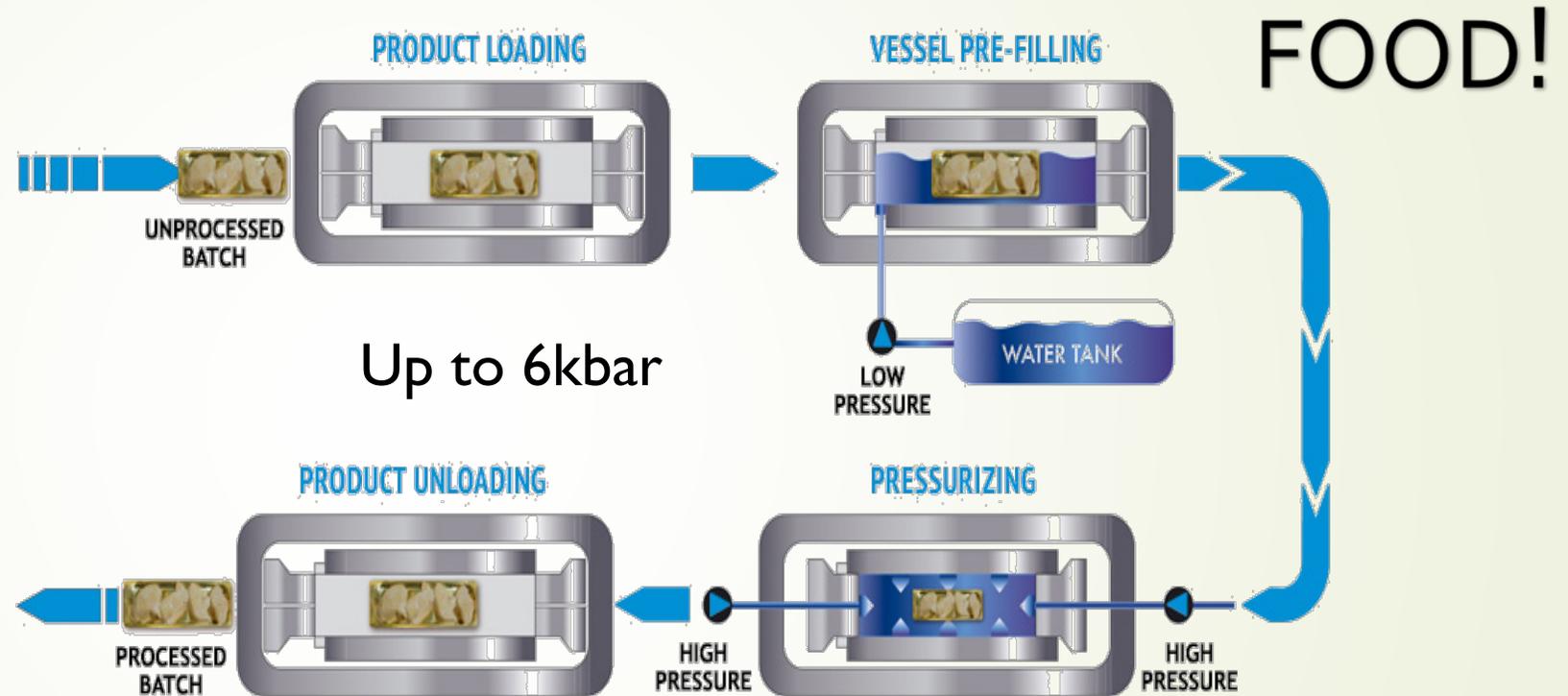
Pressure: up to 3.5 kbar.  
Temperature :  $-15^\circ\text{C}$  to  $+65^\circ\text{C}$   
 $q$ -Range:  $0.001$ - $0.3 \text{ \AA}^{-1}$   
Sample Volume: 2.5 mL per load.  
Typical Sample: 3-10 mg/mL

Adapted from Teixeira et al. (2018) *J. Neutron Res.* 20(1), 13-23.

# GOALS

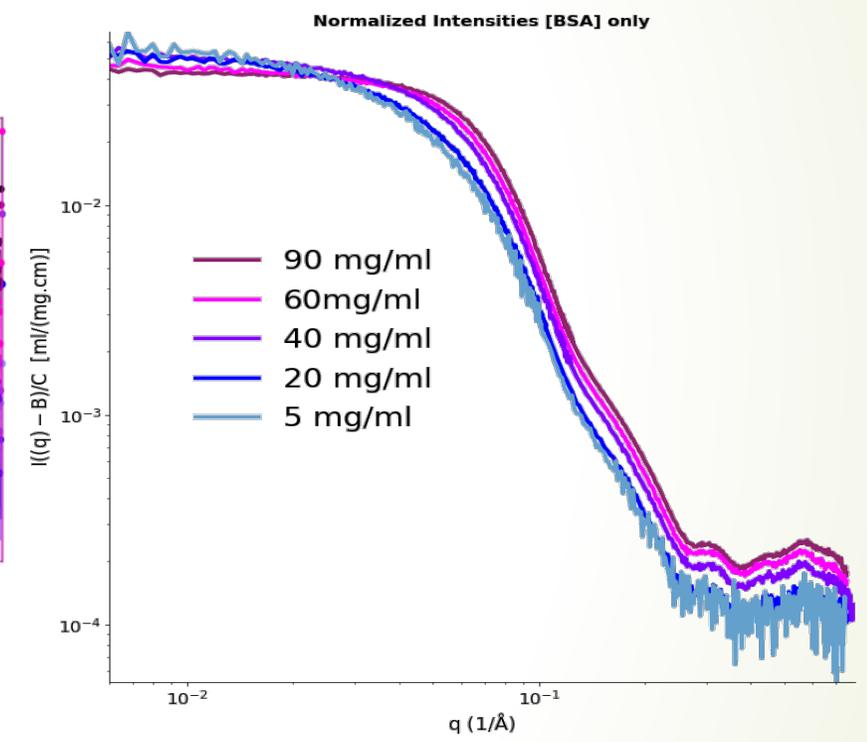
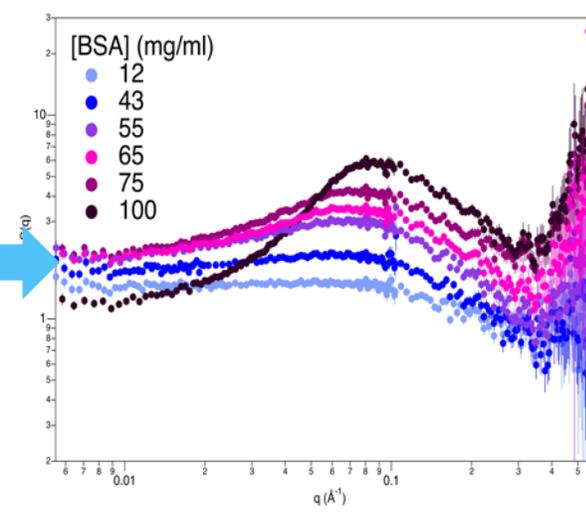
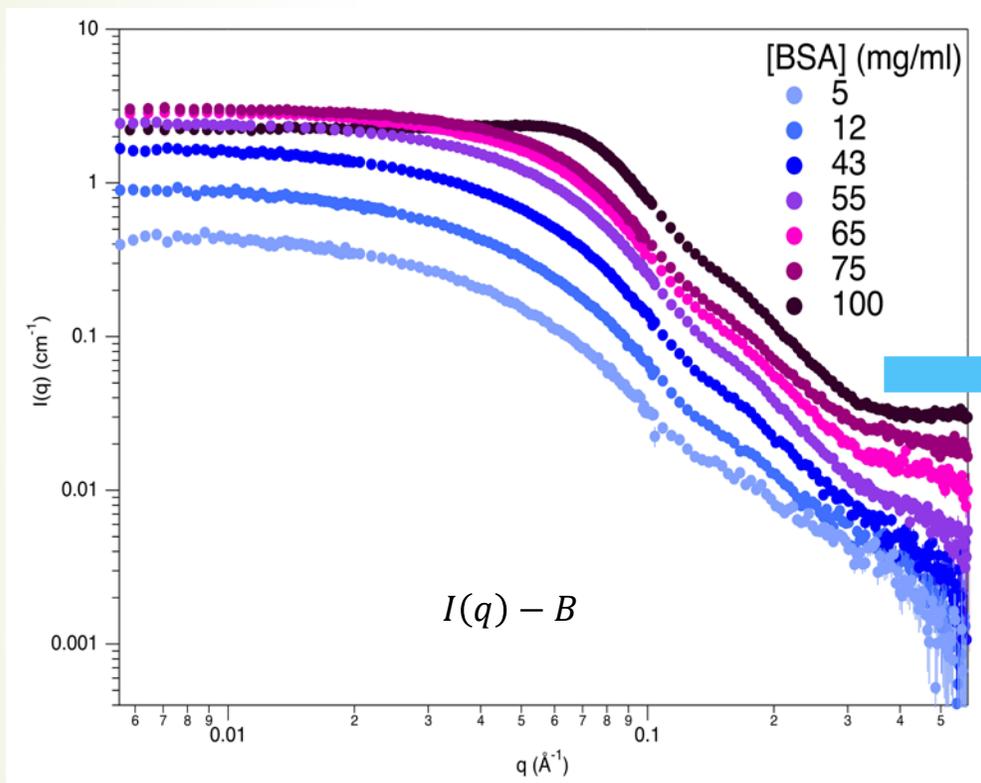
## Investigate:

- Structure and stability of BSA-polyphenol complexes under conditions relevant to food processing
- Effects of pressure on conformation and binding of polyphenols to BSA



Adapted from *Hiperbaric High Pressure Processing*.

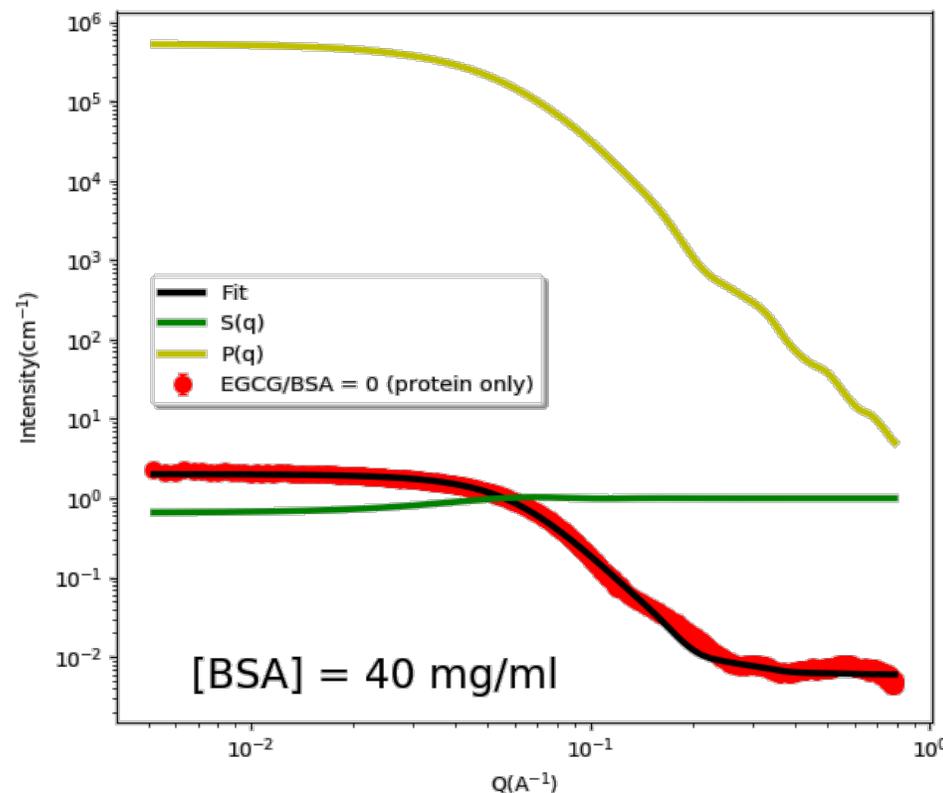
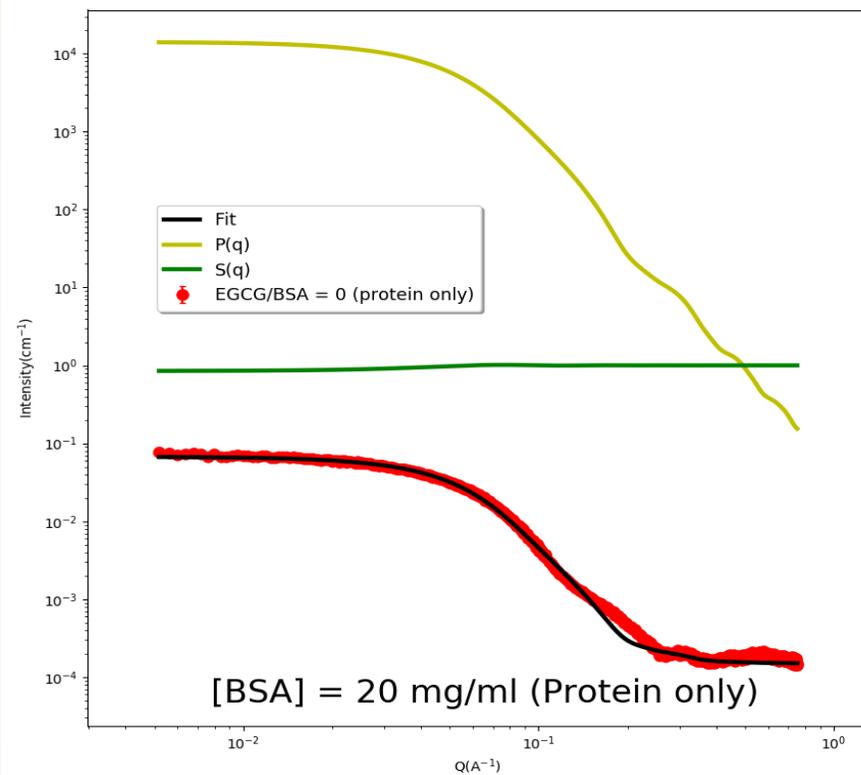
# Concentration Screening & Structure Factor



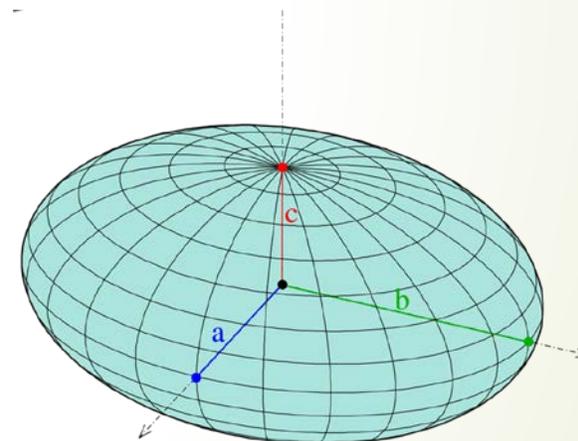
SANS (NG7 & NGB30 instruments)

SAXS (IBBR In-House Source)

# BSA Ellipsoid Model - Rough Fitting



$$I(q) = P(q) * S(q)$$

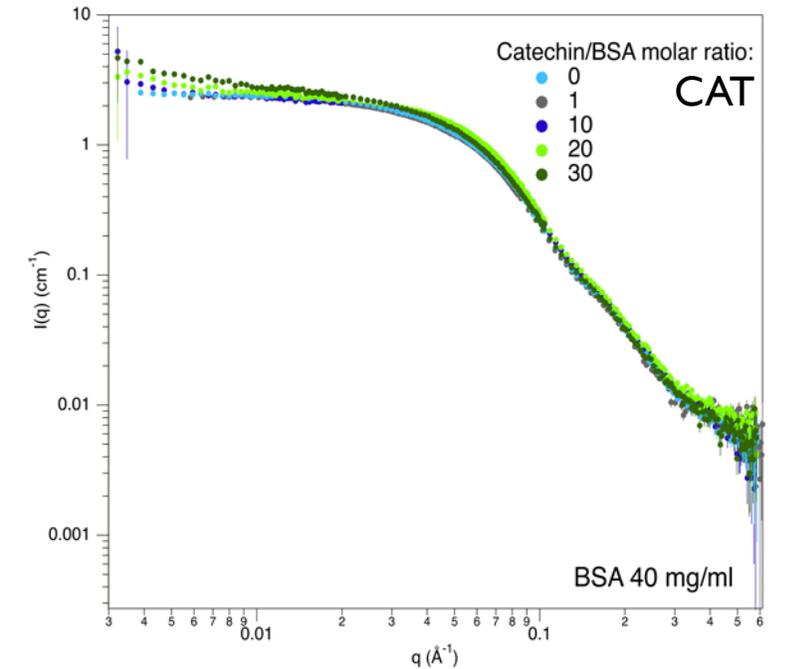
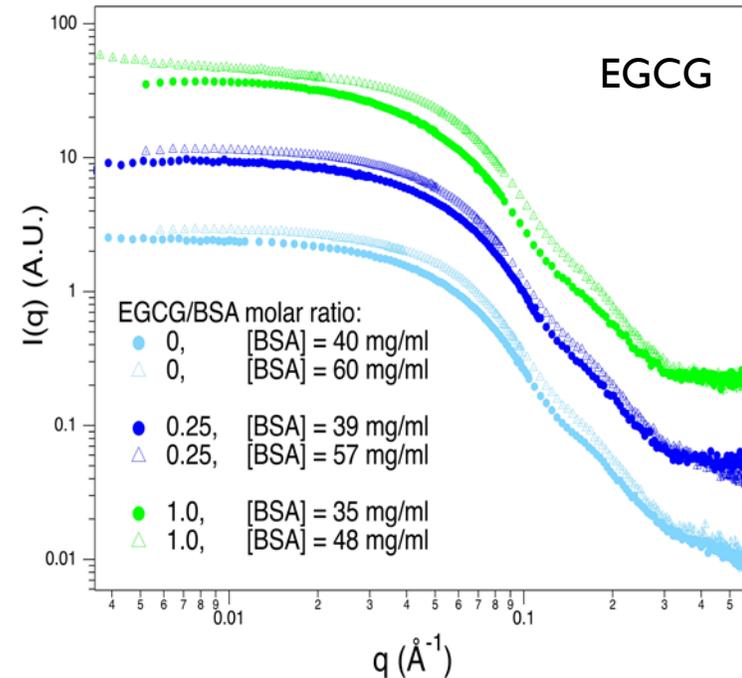
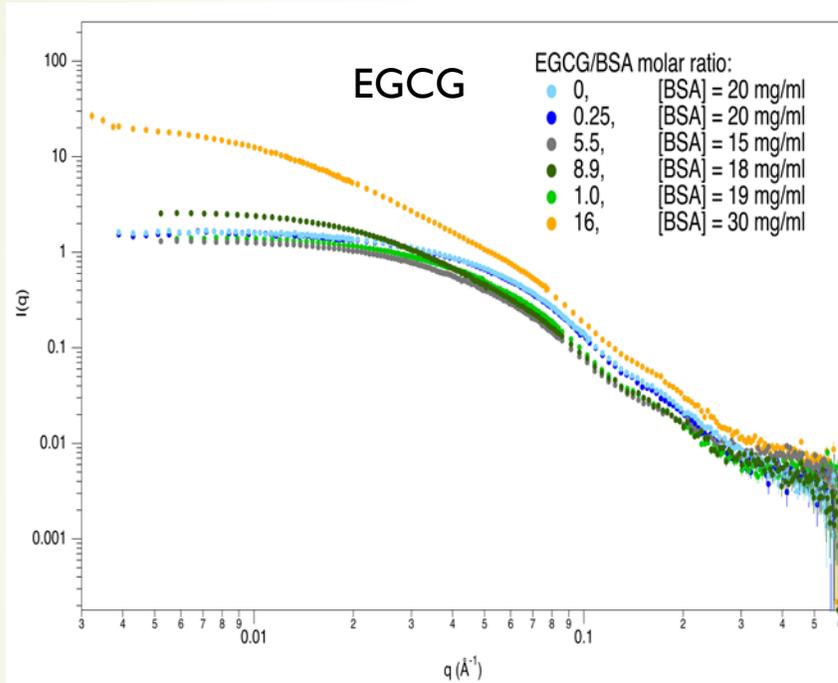


[Protein]	<b>a</b> Minor Eq. Radius (Å)	<b>b</b> Major Eq. Radius (Å)	<b>c</b> Polar Radius (Å)
20 mg/ml	42.6 ± 0.3	49.5 ± 0.3	17.7 ± 0.1
40 mg/ml	37.7 ± 0.1	64.9 ± 1.3	17.0 ± 0.1

Tri-axial model: Finnigan, J.A., Jacobs, D.J., 1971. *J. Phys. D: Appl. Phys.* 4, 72-77.

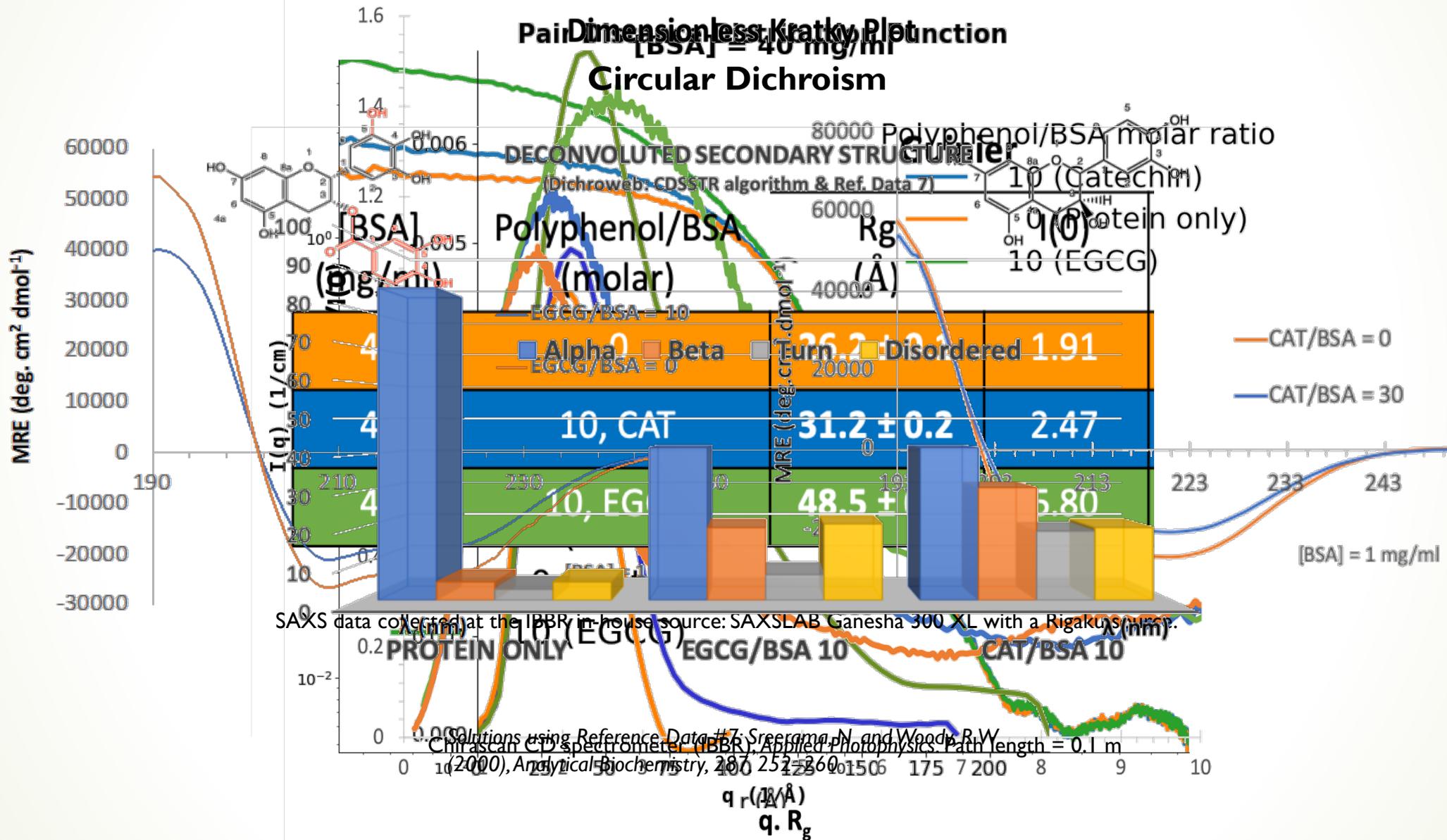
Hayter MSA Structure Factor model: J B Hayter & J Penfold, 1981. *Molecular Physics* 42, 109-118.

# Complexes of Polyphenols/BSA – Parameters Probed

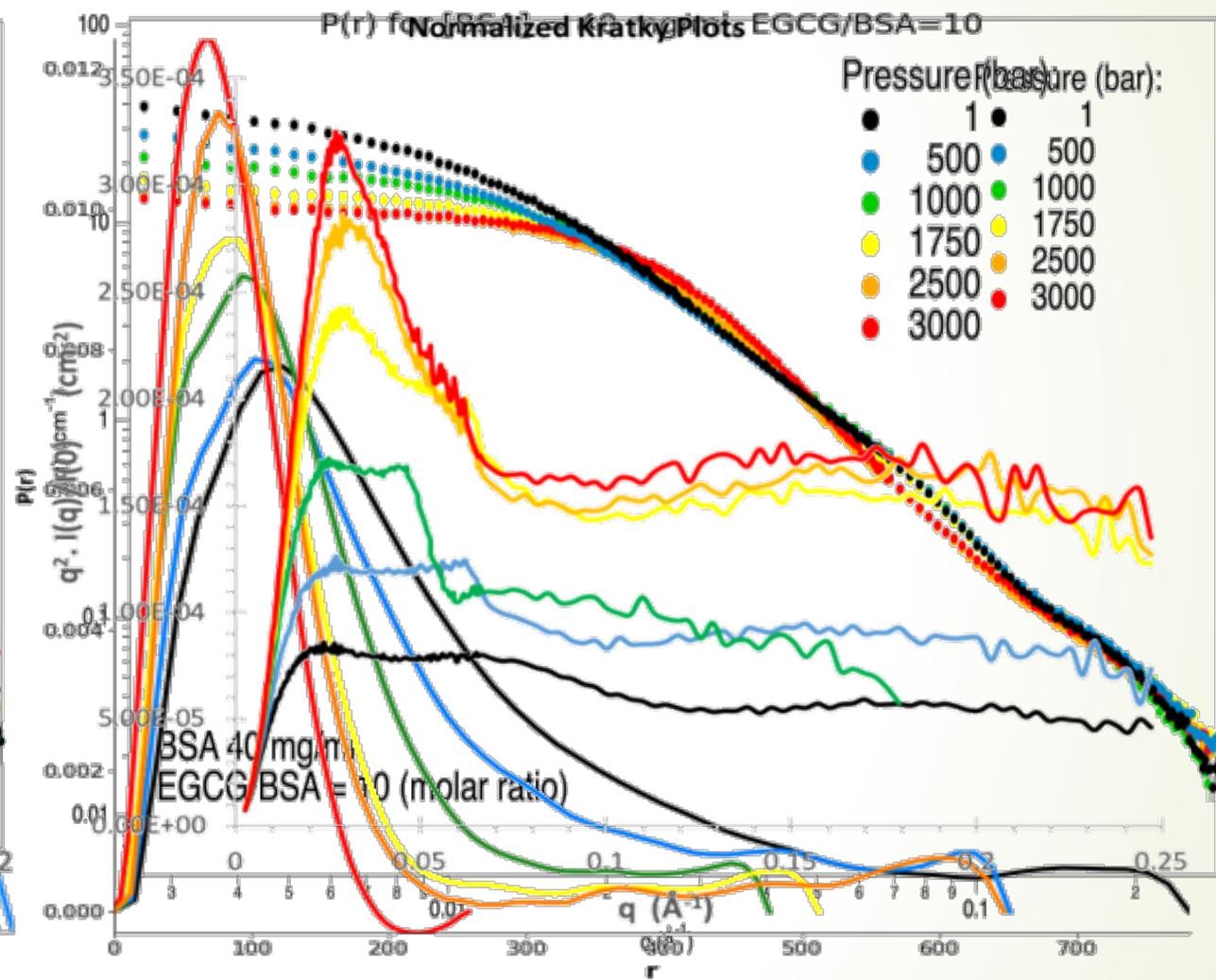
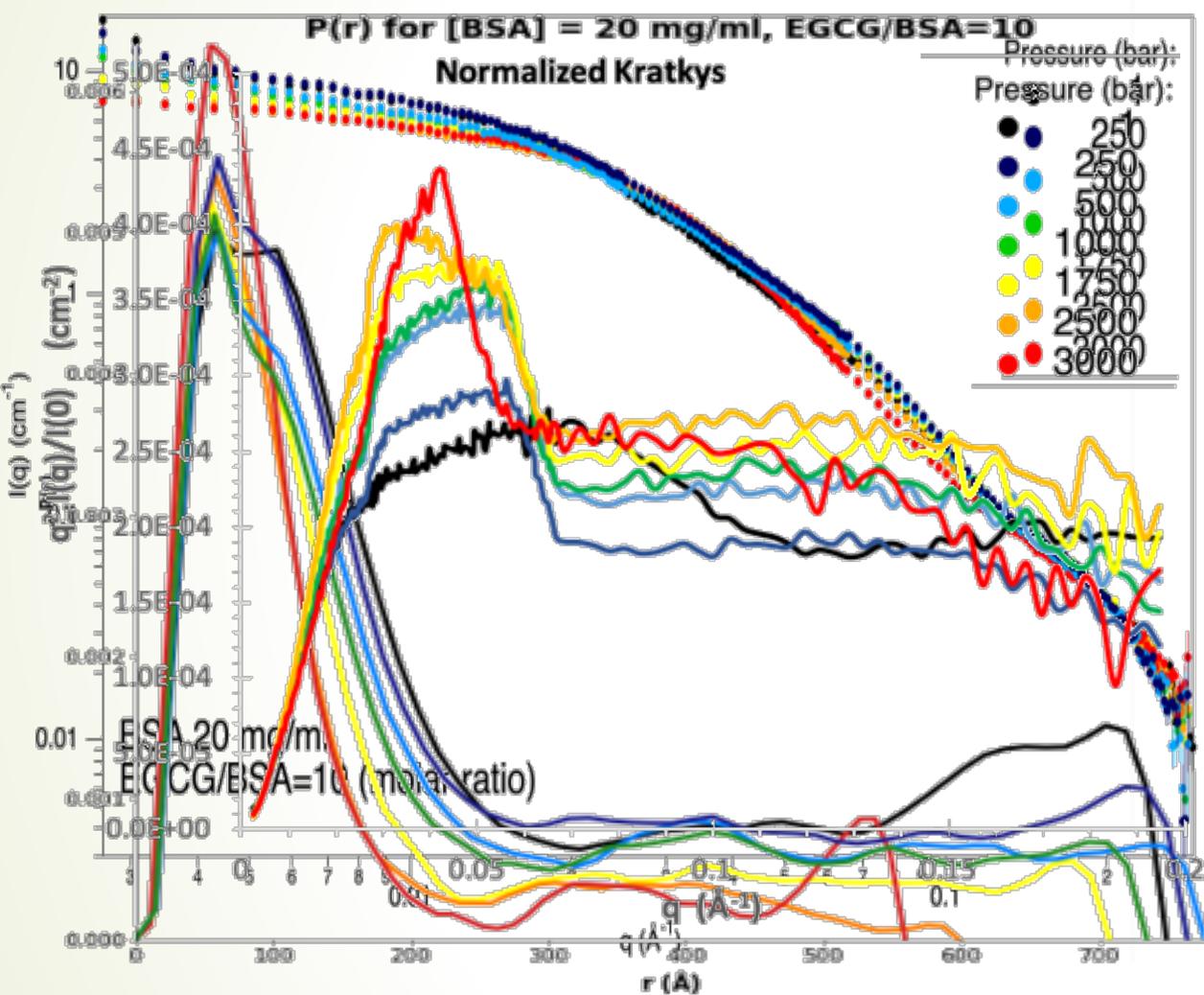


Parameters	
BSA Concentration	10 mg/mL – 60 mg/mL
Molar Ratio (Polyphenol: BSA)	0 – 30 mol
Time	2 hrs – 4 weeks
H <sub>2</sub> O/D <sub>2</sub> O Solvation	100% H <sub>2</sub> O – 100% D <sub>2</sub> O

# Catechin vs EGCG Effects on BSA Structure – SAXS and CD Data



# HP-SANS – 20 & 40 mg/mL BSA Complexed with EGCG



# Conclusions

- Established the onset of  $S(q)$  for BSA solutions
- Established SAS profiles for BSA/polyphenol complexes
- Determined the effects of polyphenols on the secondary structure of BSA
- Determined the effects of HP on the complex structure
- Established the use of HP SANS as a viable tool to monitor the effects and their reversibility of HP *in situ*

- Molecular Dynamics (MD) on the BSA crystal structure to determine models for the solution structure of the protein at different pressures
- Near-UV (250-320 nm) CD profiles of complexes under HP-SANS conditions to monitor binding effects on specific types of amino acids and disulfide bonds
- Probe HP effects vs Temperature, pH



## Future Directions

# Acknowledgements

THANK YOU!!!

- NSF - CHRNS
- Susana Teixeira (SURF Mentor, U. Delaware/NIST)
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- NCNR Neutron Condensed Matter Science Group
- Fellow SURFers (Shark Tank, Fish Bowl, Submarine, & the Exiles)



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